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(54) Title: TRANSPORTERS AND ION CHANNELS

(57) Abstract: Various embodiments of the invention provide human transporters and ion channels (TRICH) and polynucleotides which identify and encode TRICH. Embodiments of the invention also provide expression vectors, host cells, antibodies, agonists, and antagonists. Other embodiments provide methods for diagnosing, treating, or preventing disorders associated with aberrant expression of TRICH.





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TRANSPORTERS AND ION CHANNELS

TECHNICAL FIELD

The invention relates to novel nucleic acids, transporters and ion channels encoded by these nucleic acids, and to the use of these nucleic acids and proteins in the diagnosis, treatment, and prevention of transport, neurological, muscle, immunological and cell proliferative disorders. The invention also relates to the assessment of the effects of exogenous compounds on the expression of nucleic acids and transporters and ion channels.

BACKGROUND OF THE INVENTION

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Eukaryotic cells are surrounded and subdivided into functionally distinct organelles by hydrophobic lipid bilayer membranes which are highly impermeable to most polar molecules. Cells and organelles require transport proteins to import and export essential nutrients and metal ions including K⁺, NH₄⁺, P_i, SO₄²⁻, sugars, and vitamins, as well as various metabolic waste products. Transport proteins also play roles in antibiotic resistance, toxin secretion, ion balance, synaptic neurotransmission, kidney function, intestinal absorption, tumor growth, and other diverse cell functions (Griffith, J. and C. Sansom (1998) The Transporter Facts Book, Academic Press, San Diego CA, pp. 3-29). Transport can occur by a passive concentration-dependent mechanism, or can be linked to an energy source such as ATP hydrolysis or an ion gradient. Proteins that function in transport include carrier proteins, which bind to a specific solute and undergo a conformational change that translocates the bound solute across the membrane, and channel proteins, which form hydrophilic pores that allow specific solutes to diffuse through the membrane down an electrochemical solute gradient.

Carrier proteins which transport a single solute from one side of the membrane to the other are called uniporters. In contrast, coupled transporters link the transfer of one solute with simultaneous or sequential transfer of a second solute, either in the same direction (symport) or in the opposite direction (antiport). For example, intestinal and kidney epithelium contains a variety of symporter systems driven by the sodium gradient that exists across the plasma membrane. Sodium moves into the cell down its electrochemical gradient and brings the solute into the cell with it. The sodium gradient that provides the driving force for solute uptake is maintained by the ubiquitous Na⁺/K⁺ ATPase system. Sodium-coupled transporters include the mammalian glucose transporter (SGLT1), iodide transporter (NIS), and multivitamin transporter (SMVT). All three transporters have twelve putative transmembrane segments, extracellular glycosylation sites, and cytoplasmically-oriented N- and C-termini. NIS plays a crucial role in the evaluation, diagnosis, and treatment of

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various thyroid pathologies because it is the molecular basis for radioiodide thyroid-imaging techniques and for specific targeting of radioisotopes to the thyroid gland (Levy, O. et al. (1997) Proc. Natl. Acad. Sci. USA 94:5568-5573). SMVT is expressed in the intestinal mucosa, kidney, and placenta, and is implicated in the transport of the water-soluble vitamins, e.g., biotin and pantothenate (Prasad, P.D. et al. (1998) J. Biol. Chem. 273:7501-7506).

One of the largest families of transporters is the major facilitator superfamily (MFS), also called the uniporter-symporter-antiporter family. MFS transporters are single polypeptide carriers that transport small solutes in response to ion gradients. Members of the MFS are found in all classes of living organisms, and include transporters for sugars, oligosaccharides, phosphates, nitrates, nucleosides, monocarboxylates, and drugs. MFS transporters found in eukaryotes all have a structure comprising 12 transmembrane segments (Pao, S.S. et al. (1998) Microbiol. Molec. Biol. Rev. 62:1-34). The largest family of MFS transporters is the sugar transporter family, which includes the seven glucose transporters (GLUT1-GLUT7) found in humans that are required for the transport of glucose and other hexose sugars. These glucose transport proteins have unique tissue distributions and physiological functions. GLUT1 provides many cell types with their basal glucose requirements and transports glucose across epithelial and endothelial barrier tissues; GLUT2 facilitates glucose uptake or efflux from the liver; GLUT3 regulates glucose supply to neurons; GLUT4 is responsible for insulinregulated glucose disposal; and GLUT5 regulates fructose uptake into skeletal muscle. Defects in glucose transporters are involved in a recently identified neurological syndrome causing infantile seizures and developmental delay, as well as glycogen storage disease, Fanconi-Bickel syndrome, and non-insulin-dependent diabetes mellitus (Mueckler, M. (1994) Eur. J. Biochem. 219:713-725; Longo, N. and L.J. Elsas (1998) Adv. Pediatr. 45:293-313).

Monocarboxylate anion transporters are proton-coupled symporters with a broad substrate specificity that includes L-lactate, pyruvate, and the ketone bodies acetate, acetoacetate, and beta-hydroxybutyrate. At least seven isoforms have been identified to date. The isoforms are predicted to have twelve transmembrane (TM) helical domains with a large intracellular loop between TM6 and TM7, and play a critical role in maintaining intracellular pH by removing the protons that are produced stoichiometrically with lactate during glycolysis. The best characterized H⁺-monocarboxylate transporter is that of the erythrocyte membrane, which transports L-lactate and a wide range of other aliphatic monocarboxylates. Other cells possess H⁺-linked monocarboxylate transporters with differing substrate and inhibitor selectivities. In particular, cardiac muscle and tumor cells have transporters that differ in their K_m values for certain substrates, including stereoselectivity for L- over D-lactate, and in their sensitivity to inhibitors. There are Na⁺-monocarboxylate

cotransporters on the luminal surface of intestinal and kidney epithelia, which allow the uptake of lactate, pyruvate, and ketone bodies in these tissues. In addition, there are specific and selective transporters for organic cations and organic anions in organs including the kidney, intestine and liver. Organic anion transporters are selective for hydrophobic, charged molecules with electron-attracting side groups. Organic cation transporters, such as the ammonium transporter, mediate the secretion of a variety of drugs and endogenous metabolites, and contribute to the maintenance of intercellular pH (Poole, R.C. and A.P. Halestrap (1993) Am. J. Physiol. 264:C761-C782; Price, N.T. et al. (1998) Biochem. J. 329:321-328; and Martinelle, K. and I. Haggstrom (1993) J. Biotechnol. 30:339-350).

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ATP-binding cassette (ABC) transporters are members of a superfamily of membrane proteins that transport substances ranging from small molecules such as ions, sugars, amino acids, peptides, and phospholipids, to lipopeptides, large proteins, and complex hydrophobic drugs. ABC transporters consist of four modules: two nucleotide-binding domains (NBD), which hydrolyze ATP to supply the energy required for transport, and two membrane-spanning domains (MSD), each containing six putative transmembrane segments. These four modules may be encoded by a single gene, as is the case for the cystic fibrosis transmembrane regulator (CFTR), or by separate genes. When encoded by separate genes, each gene product contains a single NBD and MSD. These "halfmolecules" form homo- and heterodimers, such as Tap1 and Tap2, the endoplasmic reticulum-based major histocompatibility (MHC) peptide transport system. Several genetic diseases are attributed to defects in ABC transporters, such as the following diseases and their corresponding proteins: cystic fibrosis (CFTR, an ion channel), adrenoleukodystrophy (adrenoleukodystrophy protein, ALDP), Zellweger syndrome (peroxisomal membrane protein-70, PMP70), and hyperinsulinemic hypoglycemia (sulfonylurea receptor, SUR). Overexpression of the multidrug resistance (MDR) protein, another ABC transporter, in human cancer cells makes the cells resistant to a variety of cytotoxic drugs used in chemotherapy (Taglicht, D. and S. Michaelis (1998) Meth. Enzymol. 292:130-162).

A number of metal ions such as iron, zinc, copper, cobalt, manganese, molybdenum, selenium, nickel, and chromium are important as cofactors for a number of enzymes. For example, copper is involved in hemoglobin synthesis, connective tissue metabolism, and bone development, by acting as a cofactor in oxidoreductases such as superoxide dismutase, ferroxidase (ceruloplasmin), and lysyl oxidase. Copper and other metal ions must be provided in the diet, and are absorbed by transporters in the gastrointestinal tract. Plasma proteins transport the metal ions to the liver and other target organs, where specific transporters move the ions into cells and cellular organelles as needed. Imbalances in metal ion metabolism have been associated with a number of disease states (Danks, D.M. (1986) J. Med. Genet. 23:99-106).

Transport of fatty acids across the plasma membrane can occur by diffusion, a high capacity, low affinity process. However, under normal physiological conditions a significant fraction of fatty acid transport appears to occur via a high affinity, low capacity protein-mediated transport process. Fatty acid transport protein (FATP), an integral membrane protein with four transmembrane segments, is expressed in tissues exhibiting high levels of plasma membrane fatty acid flux, such as muscle, heart, and adipose. Expression of FATP is upregulated in 3T3-L1 cells during adipose conversion, and expression in COS7 fibroblasts elevates uptake of long-chain fatty acids (Hui, T.Y. et al. (1998) J. Biol. Chem. 273:27420-27429).

The lipocalin superfamily constitutes a phylogenetically conserved group of more than forty proteins that function as extracellular ligand-binding proteins which bind and transport small hydrophobic molecules. Members of this family function as carriers of retinoids, odorants, chromophores, pheromones, allergens, and sterols, and in a variety of processes including nutrient transport, cell growth regulation, immune response, and prostaglandin synthesis. A subset of these proteins may be multifunctional, serving as either a biosynthetic enzyme or as a specific enzyme inhibitor. (Tanaka, T. et al. (1997) J. Biol. Chem. 272:15789-15795; and van't Hof, W. et al. (1997) J. Biol. Chem. 272:1837-1841.)

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Members of the lipocalin family display unusually low levels of overall sequence conservation. Pairwise sequence identity often falls below 20%. Sequence similarity between family members is limited to conserved cysteines which form disulfide bonds and three motifs which form a juxtaposed cluster that functions as a target cell recognition site. The lipocalins share an eight stranded, antiparallel beta-sheet which folds back on itself to form a continuously hydrogen-bonded beta-barrel. The pocket formed by the barrel functions as an internal ligand binding site. Seven loops (L1 to L7) form short beta-hairpins, except loop L1 which is a large omega loop that forms a lid to partially close the internal ligand-binding site (Flower (1996) Biochem. J. 318:1-14).

Lipocalins are important transport molecules. Each lipocalin associates with a particular ligand and delivers that ligand to appropriate target sites within the organism. Retinol-binding protein (RBP), one of the best characterized lipocalins, transports retinol from stores within the liver to target tissues. Apolipoprotein D (apo D), a component of high density lipoproteins (HDLs) and low density lipoproteins (LDLs), functions in the targeted collection and delivery of cholesterol throughout the body. Lipocalins are also involved in cell regulatory processes. Apo D, which is identical to gross-cystic-disease-fluid protein (GCDFP)-24, is a progesterone/pregnenolone-binding protein expressed at high levels in breast cyst fluid. Secretion of apo D in certain human breast cancer cell lines is accompanied by reduced cell proliferation and progression of cells to a more differentiated phenotype.

Similarly, apo D and another lipocalin, α_1 -acid glycoprotein (AGP), are involved in nerve cell regeneration. AGP is also involved in anti-inflammatory and immunosuppressive activities. AGP is one of the positive acute-phase proteins (APP); circulating levels of AGP increase in response to stress and inflammatory stimulation. AGP accumulates at sites of inflammation where it inhibits platelet and neutrophil activation and inhibits phagocytosis. The immunomodulatory properties of AGP are due to glycosylation. AGP is 40% carbohydrate, making it unusually acidic and soluble. The glycosylation pattern of AGP changes during acute-phase response, and deglycosylated AGP has no immunosuppressive activity (Flower (1994) FEBS Lett. 354:7-11; Flower (1996) *supra*).

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The lipocalin superfamily also includes several animal allergens, including the mouse major urinary protein (mMUP), the rat α-2-microgloobulin (rA2U), the bovine β-lactoglobulin (βlg), the cockroach allergen (Bla g4), bovine dander allergen (Bos d2), and the major horse allergen, designated *Equus caballus* allergen 1 (Equ c1). Equ c1 is a powerful allergen responsible for about 80% of anti-horse IgE antibody response in patients who are chronically exposed to horse allergens. It appears that lipocalins may contain a common structure that is able to induce the IgE response (Gregoire, C. et al., (1996) J. Biol. Chem. 271:32951-32959).

Lipocalins are used as diagnostic and prognostic markers in a variety of disease states. The plasma level of AGP is monitored during pregnancy and in diagnosis and prognosis of conditions including cancer chemotherapy, renal disfunction, myocardial infarction, arthritis, and multiple sclerosis. RBP is used clinically as a marker of tubular reabsorption in the kidney, and apo D is a marker in gross cystic breast disease (Flower (1996) supra). Additionally, the use of lipocalin animal allergens may help in the diagnosis of allergic reactions to horses (Gregoire supra), pigs, cockroaches, mice and rats.

Mitochondrial carrier proteins are transmembrane-spanning proteins which transport ions and charged metabolites between the cytosol and the mitochondrial matrix. Examples include the ADP, ATP carrier protein; the 2-oxoglutarate/malate carrier; the phosphate carrier protein; the pyruvate carrier; the dicarboxylate carrier which transports malate, succinate, fumarate, and phosphate; the tricarboxylate carrier which transports citrate and malate; and the Grave's disease carrier protein, a protein recognized by IgG in patients with active Grave's disease, an autoimmune disorder resulting in hyperthyroidism. Proteins in this family consist of three tandem repeats of an approximately 100 amino acid domain, each of which contains two transmembrane regions (Stryer, L. (1995)

Biochemistry, W.H. Freeman and Company, New York NY, p. 551; PROSITE PDOC00189

Mitochondrial energy transfer proteins signature; Online Mendelian Inheritance in Man (OMIM)

*275000 Graves Disease).

This class of transporters also includes the mitochondrial uncoupling proteins, which create proton leaks across the inner mitochondrial membrane, thus uncoupling oxidative phosphorylation from ATP synthesis. The result is energy dissipation in the form of heat. Mitochondrial uncoupling proteins have been implicated as modulators of thermoregulation and metabolic rate, and have been proposed as potential targets for drugs against metabolic diseases such as obesity (Ricquier, D. et al. (1999) J. Int. Med. 245:637-642).

Ion Channels

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The electrical potential of a cell is generated and maintained by controlling the movement of ions across the plasma membrane. The movement of ions requires ion channels, which form ion-selective pores within the membrane. There are two basic types of ion channels, ion transporters and gated ion channels. Ion transporters utilize the energy obtained from ATP hydrolysis to actively transport an ion against the ion's concentration gradient. Gated ion channels allow passive flow of an ion down the ion's electrochemical gradient under restricted conditions. Together, these types of ion channels generate, maintain, and utilize an electrochemical gradient that is used in 1) electrical impulse conduction down the axon of a nerve cell, 2) transport of molecules into cells against concentration gradients, 3) initiation of muscle contraction, and 4) endocrine cell secretion.

Ion Transporters

Ion transporters generate and maintain the resting electrical potential of a cell. Utilizing the energy derived from ATP hydrolysis, they transport ions against the ion's concentration gradient. These transmembrane ATPases are divided into three families. The phosphorylated (P) class ion transporters, including Na⁺-K⁺ ATPase, Ca²⁺-ATPase, and H⁺-ATPase, are activated by a phosphorylation event. P-class ion transporters are responsible for maintaining resting potential distributions such that cytosolic concentrations of Na⁺ and Ca²⁺ are low and cytosolic concentration of K⁺ is high. The vacuolar (V) class of ion transporters includes H⁺ pumps on intracellular organelles, such as lysosomes and Golgi. V-class ion transporters are responsible for generating the low pH within the lumen of these organelles that is required for function. The coupling factor (F) class consists of H⁺ pumps in the mitochondria. F-class ion transporters utilize a proton gradient to generate ATP from ADP and inorganic phosphate (P_i).

The P-ATPases are hexamers of a 100 kD subunit with ten transmembrane domains and several large cytoplasmic regions that may play a role in ion binding (Scarborough, G.A. (1999) Curr. Opin. Cell Biol. 11:517-522). The V-ATPases are composed of two functional domains: the V_1 domain, a peripheral complex responsible for ATP hydrolysis; and the V_0 domain, an integral complex responsible for proton translocation across the membrane. The F-ATPases are structurally and

evolutionarily related to the V-ATPases. The F-ATPase F₀ domain contains 12 copies of the c subunit, a highly hydrophobic protein composed of two transmembrane domains and containing a single buried carboxyl group in TM2 that is essential for proton transport. The V-ATPase V₀ domain contains three types of homologous c subunits with four or five transmembrane domains and the essential carboxyl group in TM4 or TM3. Both types of complex also contain a single a subunit that may be involved in regulating the pH dependence of activity (Forgac, M. (1999) J. Biol. Chem. 274:12951-12954).

The resting potential of the cell is utilized in many processes involving carrier proteins and gated ion channels. Carrier proteins utilize the resting potential to transport molecules into and out of the cell. Amino acid and glucose transport into many cells is linked to sodium ion co-transport (symport) so that the movement of Na⁺ down an electrochemical gradient drives transport of the other molecule up a concentration gradient. Similarly, cardiac muscle links transfer of Ca²⁺ out of the cell with transport of Na⁺ into the cell (antiport).

Gated Ion Channels

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Gated ion channels control ion flow by regulating the opening and closing of pores. The ability to control ion flux through various gating mechanisms allows ion channels to mediate such diverse signaling and homeostatic functions as neuronal and endocrine signaling, muscle contraction, fertilization, and regulation of ion and pH balance. Gated ion channels are categorized according to the manner of regulating the gating function. Mechanically-gated channels open their pores in response to mechanical stress; voltage-gated channels (e.g., Na⁺, K⁺, Ca²⁺, and Cl⁻ channels) open their pores in response to changes in membrane potential; and ligand-gated channels (e.g., acetylcholine-, serotonin-, and glutamate-gated cation channels, and GABA- and glycine-gated chloride channels) open their pores in the presence of a specific ion, nucleotide, or neurotransmitter. The gating properties of a particular ion channel (i.e., its threshold for and duration of opening and closing) are sometimes modulated by association with auxiliary channel proteins and/or post translational modifications, such as phosphorylation.

Mechanically-gated or mechanosensitive ion channels act as transducers for the senses of touch, hearing, and balance, and also play important roles in cell volume regulation, smooth muscle contraction, and cardiac rhythm generation. A stretch-inactivated channel (SIC) was recently cloned from rat kidney. The SIC channel belongs to a group of channels which are activated by pressure or stress on the cell membrane and conduct both Ca²⁺ and Na⁺ (Suzuki, M. et al. (1999) J. Biol. Chem. 274:6330-6335).

The pore-forming subunits of the voltage-gated cation channels form a superfamily of ion

channel proteins. The characteristic domain of these channel proteins comprises six transmembrane domains (S1-S6), a pore-forming region (P) located between S5 and S6, and intracellular amino and carboxy termini. In the Na⁺ and Ca²⁺ subfamilies, this domain is repeated four times, while in the K⁺ channel subfamily, each channel is formed from a tetramer of either identical or dissimilar subunits. The P region contains information specifying the ion selectivity for the channel. In the case of K⁺ channels, a GYG tripeptide is involved in this selectivity (Ishii, T.M. et al. (1997) Proc. Natl. Acad. Sci. USA 94:11651-11656).

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Voltage-gated Na⁺ and K⁺ channels are necessary for the function of electrically excitable cells, such as nerve and muscle cells. Action potentials, which lead to neurotransmitter release and muscle contraction, arise from large, transient changes in the permeability of the membrane to Na⁺ and K⁺ ions. Depolarization of the membrane beyond the threshold level opens voltage-gated Na⁺ channels. Sodium ions flow into the cell, further depolarizing the membrane and opening more voltage-gated Na⁺ channels, which propagates the depolarization down the length of the cell. Depolarization also opens voltage-gated potassium channels. Consequently, potassium ions flow outward, which leads to repolarization of the membrane. Voltage-gated channels utilize charged residues in the fourth transmembrane segment (S4) to sense voltage change. The open state lasts only about 1 millisecond, at which time the channel spontaneously converts into an inactive state that cannot be opened irrespective of the membrane potential. Inactivation is mediated by the channel's N-terminus, which acts as a plug that closes the pore. The transition from an inactive to a closed state requires a return to resting potential.

Voltage-gated Na⁺ channels are heterotrimeric complexes composed of a 260 kDa poreforming α subunit that associates with two smaller auxiliary subunits, $\beta 1$ and $\beta 2$. The $\beta 2$ subunit is a integral membrane glycoprotein that contains an extracellular Ig domain, and its association with α and $\beta 1$ subunits correlates with increased functional expression of the channel, a change in its gating properties, as well as an increase in whole cell capacitance due to an increase in membrane surface area (Isom, L.L. et al. (1995) Cell 83:433-442).

Non voltage-gated Na⁺ channels include the members of the amiloride-sensitive Na⁺ channel/degenerin (NaC/DEG) family. Channel subunits of this family are thought to consist of two transmembrane domains flanking a long extracellular loop, with the amino and carboxyl termini located within the cell. The NaC/DEG family includes the epithelial Na⁺ channel (ENaC) involved in Na⁺ reabsorption in epithelia including the airway, distal colon, cortical collecting duct of the kidney, and exocrine duct glands. Mutations in ENaC result in pseudohypoaldosteronism type 1 and Liddle's syndrome (pseudohyperaldosteronism). The NaC/DEG family also includes the recently characterized

H⁺-gated cation channels or acid-sensing ion channels (ASIC). ASIC subunits are expressed in the brain and form heteromultimeric Na⁺-permeable channels. These channels require acid pH fluctuations for activation. ASIC subunits show homology to the degenerins, a family of mechanically-gated channels originally isolated from C. elegans. Mutations in the degenerins cause neurodegeneration. ASIC subunits may also have a role in neuronal function, or in pain perception, since tissue acidosis causes pain (Waldmann, R. and M. Lazdunski (1998) Curr. Opin. Neurobiol. 8:418-424; Eglen, R.M. et al. (1999) Trends Pharmacol. Sci. 20:337-342).

K⁺ channels are located in all cell types, and may be regulated by voltage, ATP concentration, or second messengers such as Ca²⁺ and cAMP. In non-excitable tissue, K⁺ channels are involved in protein synthesis, control of endocrine secretions, and the maintenance of osmotic equilibrium across membranes. In neurons and other excitable cells, in addition to regulating action potentials and repolarizing membranes, K⁺ channels are responsible for setting the resting membrane potential. The cytosol contains non-diffusible anions and, to balance this net negative charge, the cell contains a Na⁺-K⁺ pump and ion channels that provide the redistribution of Na⁺, K⁺, and Cl⁻. The pump actively transports Na⁺ out of the cell and K⁺ into the cell in a 3:2 ratio. Ion channels in the plasma membrane allow K⁺ and Cl⁻ to flow by passive diffusion. Because of the high negative charge within the cytosol, -Cl⁻ flows out of the cell. The flow of K⁺ is balanced by an electromotive force pulling K⁺ into the cell, and a K⁺ concentration gradient pushing K⁺ out of the cell. Thus, the resting membrane potential is primarily regulated by K⁺ flow (Salkoff, L. and T. Jegla (1995) Neuron 15:489-492).

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Potassium channel subunits of the *Shaker*-like superfamily all have the characteristic six transmembrane/1 pore domain structure. Four subunits combine as homo- or heterotetramers to form functional K channels. These pore-forming subunits also associate with various cytoplasmic β subunits that alter channel inactivation kinetics. The *Shaker*-like channel family includes the voltage-gated K⁺ channels as well as the delayed rectifier type channels such as the human ether-a-go-go related gene (HERG) associated with long QT, a cardiac dysrythmia syndrome (Curran, M.E. (1998) Curr. Opin. Biotechnol. 9:565-572; Kaczorowski, G.J. and M.L. Garcia (1999) Curr. Opin. Chem. Biol. 3:448-458).

A second superfamily of K⁺ channels is composed of the inward rectifying channels (Kir). Kir channels have the property of preferentially conducting K⁺ currents in the inward direction. These proteins consist of a single potassium selective pore domain and two transmembrane domains, which correspond to the fifth and sixth transmembrane domains of voltage-gated K⁺ channels. Kir subunits also associate as tetramers. The Kir family includes ROMK1, mutations in which lead to Bartter syndrome, a renal tubular disorder. Kir channels are also involved in regulation of cardiac pacemaker

activity, seizures and epilepsy, and insulin regulation (Doupnik, C.A. et al. (1995) Curr. Opin. Neurobiol. 5:268-277; Curran, *supra*).

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The recently recognized TWIK K⁺ channel family includes the mammalian TWIK-1, TREK-1 and TASK proteins. Members of this family possess an overall structure with four transmembrane domains and two P domains. These proteins are probably involved in controlling the resting potential in a large set of cell types (Duprat, F. et al. (1997) EMBO J 16:5464-5471).

The voltage-gated Ca²⁺ channels have been classified into several subtypes based upon their electrophysiological and pharmacological characteristics. L-type Ca²⁺ channels are predominantly expressed in heart and skeletal muscle where they play an essential role in excitation-contraction coupling. T-type channels are important for cardiac pacemaker activity, while N-type and P/Q-type channels are involved in the control of neurotransmitter release in the central and peripheral nervous system. The L-type and N-type voltage-gated Ca²⁺ channels have been purified and, though their functions differ dramatically, they have similar subunit compositions. The channels are composed of three subunits. The α_1 subunit forms the membrane pore and voltage sensor, while the $\alpha_2\delta$ and β subunits modulate the voltage-dependence, gating properties, and the current amplitude of the channel. These subunits are encoded by at least six α_1 , one $\alpha_2\delta$, and four β genes. A fourth subunit, γ , has been identified in skeletal muscle (Walker, D. et al. (1998) J. Biol. Chem. 273:2361-2367; McCleskey, E.W. (1994) Curr. Opin. Neurobiol. 4:304-312).

The high-voltage-activated Ca ²⁺ channels that have been characterized biochemically include complexes of a pore-forming alpha1 subunit of approximately 190-250 kDa; a transmembrane complex of alpha2 and delta subunits; an intracellular beta subunit; and in some cases a transmembrane gamma subunit. A variety of alpha1 subunits, alpha2delta complexes, beta subunits, and gamma subunits are known. The Cav1 family of alpha1 subunits conduct L-type Ca ²⁺ currents, which initiate muscle contraction, endocrine secretion, and gene transcription, and are regulated primarily by second messenger-activated protein phosphorylation pathways. The Cav2 family of alpha1 subunits conduct N-type, P/Q-type, and R-type Ca ²⁺ currents, which initiate rapid synaptic transmission and are regulated primarily by direct interaction with G proteins and SNARE proteins and secondarily by protein phosphorylation. The Cav3 family of alpha1 subunits conduct T-type Ca ²⁺ currents, which are activated and inactivated more rapidly and at more negative membrane potentials than other Ca ²⁺ current types. The distinct structures and patterns of regulation of these three families of Ca ²⁺ channels provide an array of Ca ²⁺ entry pathways in response to changes in membrane potential and a range of possibilities for regulation of Ca ²⁺ entry by second messenger pathways and interacting proteins (Catterall, W.A. (2000) Annu. Rev. Cell Dev. Biol. 16:521-555).

The alpha-2 subunit of the voltage-gated Ca ²⁺-channel may include one or more Cache domains. An extracellular Cache domain may be fused to an intracellular catalytic domain, such as the histidine kinase, PP2C phosphatase, GGDEF (a predicted diguanylate cyclase), HD-GYP (a predicted phosphodiesterase) or adenylyl cyclase domain, or to a noncatalytic domain, like the methyl-accepting, DNA-binding winged helix-turn-helix, GAF, PAS or HAMP (a domain found in istidine kinases, denylyl cyclases, ethyl-binding proteins and phosphatases). Small molecules are bound via the Cache domain and this signal is converted into diverse outputs depending on the intracellular domains (Anantharaman, V. and Aravind, L.(2000) Trends Biochem. Sci. 25:535-537).

The transient receptor family (Trp) of calcium ion channels are thought to mediate capacitative calcium entry (CCE). CCE is the Ca²⁺ influx into cells to resupply Ca²⁺ stores depleted by the action of inositol triphosphate (IP3) and other agents in response to numerous hormones and growth factors. Trp and Trp-like were first cloned from Drosophila and have similarity to voltage gated Ca²⁺ channels in the S3 through S6 regions. This suggests that Trp and/or related proteins may form mammalian CCE channels (Zhu, X. et al. (1996) Cell 85:661-671; Boulay, G. et al. (1997) J. Biol. Chem. 272:29672-29680). Melastatin is a gene isolated in both the mouse and human, whose expression in melanoma cells is inversely correlated with melanoma aggressiveness *in vivo*. The human cDNA transcript corresponds to a 1533-amino acid protein having homology to members of the Trp family. It has been proposed that the combined use of malastatin mRNA expression status and tumor thickness might allow for the determination of subgroups of patients at both low and high risk for developing metastatic disease (Duncan, L.M. et al (2001) J. Clin. Oncol. 19:568-576).

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Chloride channels are necessary in endocrine secretion and in regulation of cytosolic and organelle pH. In secretory epithelial cells, Cl⁻ enters the cell across a basolateral membrane through an Na⁺, K⁺/Cl⁻ cotransporter, accumulating in the cell above its electrochemical equilibrium concentration. Secretion of Cl⁻ from the apical surface, in response to hormonal stimulation, leads to flow of Na⁺ and water into the secretory lumen. The cystic fibrosis transmembrane conductance regulator (CFTR) is a chloride channel encoded by the gene for cystic fibrosis, a common fatal genetic disorder in humans. CFTR is a member of the ABC transporter family, and is composed of two domains each consisting of six transmembrane domains followed by a nucleotide-binding site. Loss of CFTR function decreases transepithelial water secretion and, as a result, the layers of mucus that coat the respiratory tree, pancreatic ducts, and intestine are dehydrated and difficult to clear. The resulting blockage of these sites leads to pancreatic insufficiency, "meconium ileus", and devastating "chronic obstructive pulmonary disease" (Al-Awqati, Q. et al. (1992) J. Exp. Biol. 172:245-266).

The voltage-gated chloride channels (CLC) are characterized by 10-12 transmembrane

domains, as well as two small globular domains known as CBS domains. The CLC subunits probably function as homotetramers. CLC proteins are involved in regulation of cell volume, membrane potential stabilization, signal transduction, and transepithelial transport. Mutations in CLC-1, expressed predominantly in skeletal muscle, are responsible for autosomal recessive generalized myotonia and autosomal dominant myotonia congenita, while mutations in the kidney channel CLC-5 lead to kidney stones (Jentsch, T.J. (1996) Curr. Opin. Neurobiol. 6:303-310).

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Ligand-gated channels open their pores when an extracellular or intracellular mediator binds to the channel. Neurotransmitter-gated channels are channels that open when a neurotransmitter binds to their extracellular domain. These channels exist in the postsynaptic membrane of nerve or muscle cells. There are two types of neurotransmitter-gated channels. Sodium channels open in response to excitatory neurotransmitters, such as acetylcholine, glutamate, and serotonin. This opening causes an influx of Na⁺ and produces the initial localized depolarization that activates the voltage-gated channels and starts the action potential. Chloride channels open in response to inhibitory neurotransmitters, such as γ -aminobutyric acid (GABA) and glycine, leading to hyperpolarization of the membrane and the subsequent generation of an action potential. Neurotransmitter-gated ion channels have four transmembrane domains and probably function as pentamers (Jentsch, *supra*). Amino acids in the second transmembrane domain appear to be important in determining channel permeation and selectivity (Sather, W.A. et al. (1994) Curr. Opin. Neurobiol. 4:313-323).

Ligand-gated channels can be regulated by intracellular second messengers. For example, calcium-activated K+ channels are gated by internal calcium ions. In nerve cells, an influx of calcium during depolarization opens K+ channels to modulate the magnitude of the action potential (Ishi et al., supra). The large conductance (BK) channel has been purified from brain and its subunit composition determined. The α subunit of the BK channel has seven rather than six transmembrane domains in contrast to voltage-gated K+ channels. The extra transmembrane domain is located at the subunit N-terminus. A 28-amino-acid stretch in the C-terminal region of the subunit (the "calcium bowl" region) contains many negatively charged residues and is thought to be the region responsible for calcium binding. The β subunit consists of two transmembrane domains connected by a glycosylated extracellular loop, with intracellular N- and C-termini (Kaczorowski, supra; Vergara, C. et al. (1998) Curr. Opin. Neurobiol. 8:321-329).

Cyclic nucleotide-gated (CNG) channels are gated by cytosolic cyclic nucleotides. The best examples of these are the cAMP-gated Na+ channels involved in olfaction and the cGMP-gated cation channels involved in vision. Both systems involve ligand-mediated activation of a G-protein coupled receptor which then alters the level of cyclic nucleotide within the cell. CNG channels also

represent a major pathway for Ca²⁺ entry into neurons, and play roles in neuronal development and plasticity. CNG channels are tetramers containing at least two types of subunits, an α subunit which can form functional homomeric channels, and a β subunit, which modulates the channel properties. All CNG subunits have six transmembrane domains and a pore forming region between the fifth and sixth transmembrane domains, similar to voltage-gated K⁺ channels. A large C-terminal domain contains a cyclic nucleotide binding domain, while the N-terminal domain confers variation among channel subtypes (Zufall, F. et al. (1997) Curr. Opin. Neurobiol. 7:404-412).

The activity of other types of ion channel proteins may also be modulated by a variety of intracellular signaling proteins. Many channels have sites for phosphorylation by one or more protein kinases including protein kinase A, protein kinase C, tyrosine kinase, and casein kinase II, all of which regulate ion channel activity in cells. Kir channels are activated by the binding of the Gβγ subunits of heterotrimeric G-proteins (Reimann, F. and F.M. Ashcroft (1999) Curr. Opin. Cell. Biol. 11:503-508). Other proteins are involved in the localization of ion channels to specific sites in the cell membrane. Such proteins include the PDZ domain proteins known as MAGUKs (membrane-associated guanylate kinases) which regulate the clustering of ion channels at neuronal synapses (Craven, S.E. and D.S. Bredt (1998) Cell 93:495-498).

Disease Correlation

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The etiology of numerous human diseases and disorders can be attributed to defects in the transport of molecules across membranes. Defects in the trafficking of membrane-bound transporters and ion channels are associated with several disorders, e.g., cystic fibrosis, glucose-galactose malabsorption syndrome, hypercholesterolemia, von Gierke disease, and certain forms of diabetes mellitus. Single-gene defect diseases resulting in an inability to transport small molecules across membranes include, e.g., cystinuria, iminoglycinuria, Hartup disease, and Fanconi disease (van't Hoff, W.G. (1996) Exp. Nephrol. 4:253-262; Talente, G.M. et al. (1994) Ann. Intern. Med. 120:218-226; and Chillon, M. et al. (1995) New Engl. J. Med. 332:1475-1480).

Human diseases caused by mutations in ion channel genes include disorders of skeletal muscle, cardiac muscle, and the central nervous system. Mutations in the pore-forming subunits of sodium and chloride channels cause myotonia, a muscle disorder in which relaxation after voluntary contraction is delayed. Sodium channel myotonias have been treated with channel blockers. Mutations in muscle sodium and calcium channels cause forms of periodic paralysis, while mutations in the sarcoplasmic calcium release channel, T-tubule calcium channel, and muscle sodium channel cause malignant hyperthermia. Cardiac arrythmia disorders such as the long QT syndromes and idiopathic ventricular fibrillation are caused by mutations in potassium and sodium channels (Cooper,

E.C. and L.Y. Jan (1998) Proc. Natl. Acad. Sci. USA 96:4759-4766). All four known human idiopathic epilepsy genes code for ion channel proteins (Berkovic, S.F. and I.E. Scheffer (1999) Curr. Opin. Neurology 12:177-182). Other neurological disorders such as ataxias, hemiplegic migraine and hereditary deafness can also result from mutations in ion channel genes (Jen, J. (1999) Curr. Opin. Neurobiol. 9:274-280; Cooper, *supra*).

Ion channels have been the target for many drug therapies. Neurotransmitter-gated channels have been targeted in therapies for treatment of insomnia, anxiety, depression, and schizophrenia. Voltage-gated channels have been targeted in therapies for arrhythmia, ischemic stroke, head trauma, and neurodegenerative disease (Taylor, C.P. and L.S. Narasimhan (1997) Adv. Pharmacol. 39:47-98). Various classes of ion channels also play an important role in the perception of pain, and thus are potential targets for new analgesics. These include the vanilloid-gated ion channels, which are activated by the vanilloid capsaicin, as well as by noxious heat. Local anesthetics such as lidocaine and mexiletine which blockade voltage-gated Na⁺ channels have been useful in the treatment of neuropathic pain (Eglen, *supra*).

Ion channels in the immune system have recently been suggested as targets for immunomodulation. T-cell activation depends upon calcium signaling, and a diverse set of T-cell specific ion channels has been characterized that affect this signaling process. Channel blocking agents can inhibit secretion of lymphokines, cell proliferation, and killing of target cells. A peptide antagonist of the T-cell potassium channel Kv1.3 was found to suppress delayed-type hypersensitivity and allogenic responses in pigs, validating the idea of channel blockers as safe and efficacious immunosuppressants (Cahalan, M.D. and K.G. Chandy (1997) Curr. Opin. Biotechnol. 8:749-756).

Expression profiling

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Microarrays are analytical tools used in bioanalysis. A microarray has a plurality of molecules spatially distributed over, and stably associated with, the surface of a solid support. Microarrays of polypeptides, polynucleotides, and/or antibodies have been developed and find use in a variety of applications, such as gene sequencing, monitoring gene expression, gene mapping, bacterial identification, drug discovery, and combinatorial chemistry.

One area in particular in which microarrays find use is in gene expression analysis. Array technology can provide a simple way to explore the expression of a single polymorphic gene or the expression profile of a large number of related or unrelated genes. When the expression of a single gene is examined, arrays are employed to detect the expression of a specific gene or its variants. When an expression profile is examined, arrays provide a platform for identifying genes that are tissue specific, are affected by a substance being tested in a toxicology assay, are part of a signaling

cascade, carry out housekeeping functions, or are specifically related to a particular genetic predisposition, condition, disease, or disorder.

The potential application of gene expression profiling is relevant to improving the diagnosis, prognosis, and treatment of cancers, such as breast cancer, lung cancer, prostate cancer, ovarian cancer, and bone cancer, as well as the treatment of vascular inflammation and immune responses, liver toxicity, and neurological disorders.

Breast cancer

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More than 180,000 new cases of breast cancer are diagnosed each year, and the mortality rate for breast cancer approaches 10% of all deaths in females between the ages of 45-54 (Gish, K. (1999) AWIS Magazine 28:7-10). However the survival rate based on early diagnosis of localized breast cancer is extremely high (97%), compared with the advanced stage of the disease in which the tumor has spread beyond the breast (22%). Current procedures for clinical breast examination are lacking in sensitivity and specificity, and efforts are underway to develop comprehensive gene expression profiles for breast cancer that may be used in conjunction with conventional screening methods to improve diagnosis and prognosis of this disease (Perou, C.M. et al. (2000) Nature 406:747-752).

Mutations in two genes, BRCA1 and BRCA2, are known to greatly predispose a woman to breast cancer and may be passed on from parents to children (Gish, *supra*). However, this type of hereditary breast cancer accounts for only about 5% to 9% of breast cancers, while the vast majority of breast cancer is due to non-inherited mutations that occur in breast epithelial cells.

The relationship between expression of epidermal growth factor (EGF) and its receptor, EGFR, to human mammary carcinoma has been particularly well studied. (See Khazaie, K. et al. (1993) Cancer and Metastasis Rev. 12:255-274, and references cited therein for a review of this area.) Overexpression of EGFR, particularly coupled with down-regulation of the estrogen receptor, is a marker of poor prognosis in breast cancer patients. In addition, EGFR expression in breast tumor metastases is frequently elevated relative to the primary tumor, suggesting that EGFR is involved in tumor progression and metastasis. This is supported by accumulating evidence that EGF has effects on cell functions related to metastatic potential, such as cell motility, chemotaxis, secretion and differentiation. Changes in expression of other members of the erbB receptor family, of which EGFR is one, have also been implicated in breast cancer. The abundance of erbB receptors, such as HER-2/neu, HER-3, and HER-4, and their ligands in breast cancer points to their functional importance in the pathogenesis of the disease, and may therefore provide targets for therapy of the disease (Bacus, S.S. et al. (1994) Am. J. Clin. Pathol. 102:S13-S24). Other known markers of breast cancer include a

human secreted frizzled protein mRNA that is downregulated in breast tumors; the matrix G1a protein which is overexpressed in human breast carcinoma cells; Drg1 or RTP, a gene whose expression is diminished in colon, breast, and prostate tumors; maspin, a tumor suppressor gene downregulated in invasive breast carcinomas; and CaN19, a member of the S100 protein family, all of which are downregulated in mammary carcinoma cells relative to normal mammary epithelial cells (Zhou, Z. et al. (1998) Int. J. Cancer 78:95-99; Chen, L. et al. (1990) Oncogene 5:1391-1395; Ulrix, W. et al (1999) FEBS Lett 455:23-26; Sager, R. et al. (1996) Curr. Top. Microbiol. Immunol. 213:51-64; and Lee, S.W. et al. (1992) Proc. Natl. Acad. Sci. USA 89:2504-2508).

Cell lines derived from human mammary epithelial cells at various stages of breast cancer provide a useful model to study the process of malignant transformation and tumor progression as it has been shown that these cell lines retain many of the properties of their parental tumors for lengthy culture periods (Wistuba, I.I. et al. (1998) Clin. Cancer Res. 4:2931-2938). Such a model is particularly useful for comparing phenotypic and molecular characteristics of human mammary epithelial cells at various stages of malignant transformation.

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Lung cancer

Lung cancer is the leading cause of cancer death in the United States, affecting more than 100,000 men and 50,000 women each year. Nearly 90% of the patients diagnosed with lung cancer are cigarette smokers. Tobacco smoke contains thousands of noxious substances that induce carcinogen metabolizing enzymes and covalent DNA adduct formation in the exposed bronchial epithelium. In nearly 80% of patients diagnosed with lung cancer, metastasis has already occurred. Most commonly lung cancers metastasize to pleura, brain, bone, pericardium, and liver. The decision to treat with surgery, radiation therapy, or chemotherapy is made on the basis of tumor histology, response to growth factors or hormones, and sensitivity to inhibitors or drugs. With current treatments, most patients die within one year of diagnosis. Earlier diagnosis and a systematic approach to identification, staging, and treatment of lung cancer could positively affect patient outcome.

Lung cancers progress through a series of morphologically distinct stages from hyperplasia to invasive carcinoma. Malignant lung cancers are divided into two groups comprising four histopathological classes. The Non Small Cell Lung Carcinoma (NSCLC) group includes squamous cell carcinomas, adenocarcinomas, and large cell carcinomas and accounts for about 70% of all lung cancer cases. Adenocarcinomas typically arise in the peripheral airways and often form mucin secreting glands. Squamous cell carcinomas typically arise in proximal airways. The histogenesis of

squamous cell carcinomas may be related to chronic inflammation and injury to the bronchial epithelium, leading to squamous metaplasia. The Small Cell Lung Carcinoma (SCLC) group accounts for about 20% of lung cancer cases. SCLCs typically arise in proximal airways and exhibit a number of paraneoplastic syndromes including inappropriate production of adrenocorticotropin and anti-diuretic hormone.

Lung cancer cells accumulate numerous genetic lesions, many of which are associated with cytologically visible chromosomal aberrations. The high frequency of chromosomal deletions associated with lung cancer may reflect the role of multiple tumor suppressor loci in the etiology of this disease. Deletion of the short arm of chromosome 3 is found in over 90% of cases and represents one of the earliest genetic lesions leading to lung cancer. Deletions at chromosome arms 9p and 17p are also common. Other frequently observed genetic lesions include overexpression of telomerase, activation of oncogenes such as K-ras and c-myc, and inactivation of tumor suppressor genes such as RB, p53 and CDKN2.

Genes differentially regulated in lung cancer have been identified by a variety of methods. Using mRNA differential display technology, Manda et al. (1999; Genomics 51:5-14) identified five genes differentially expressed in lung cancer cell lines compared to normal bronchial epithelial cells. Among the known genes, pulmonary surfactant apoprotein A and alpha 2 macroglobulin were down regulated whereas nm23H1 was upregulated. Petersen et al.. (2000; Int J. Cancer, 86:512-517) used suppression subtractive hybridization to identify 552 clones differentially expressed in lung tumor derived cell lines, 205 of which represented known genes. Among the known genes, thrombospondin-1, fibronectin, intercellular adhesion molecule 1, and cytokeratins 6 and 18 were previously observed to be differentially expressed in lung cancers. Wang et al. (2000; Oncogene 19:1519-1528) used a combination of microarray analysis and subtractive hybridization to identify 17 genes differentially overexpressed in squamous cell carcinoma compared with normal lung epithelium. Among the known genes they identified were keratin isoform 6, KOC, SPRC, IGFb2, connexin 26, plakofillin 1 and cytokeratin 13.

Prostate Cancer

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Prostate cancer is a common malignancy in men over the age of 50, and the incidence increases with age. In the US, there are approximately 132,000 newly diagnosed cases of prostate cancer and more than 33,000 deaths from the disorder each year.

Once cancer cells arise in the prostate, they are stimulated by testosterone to a more rapid growth. Thus, removal of the testes can indirectly reduce both rapid growth and metastasis of the cancer. Over 95 percent of prostatic cancers are adenocarcinomas which originate in the prostatic

acini. The remaining 5 percent are divided between squamous cell and transitional cell carcinomas, both of which arise in the prostatic ducts or other parts of the prostate gland.

As with most tumors, prostate cancer develops through a multistage progression ultimately resulting in an aggressive tumor phenotype. The initial step in tumor progression involves the hyperproliferation of normal luminal and/or basal epithelial cells. Androgen responsive cells become hyperplastic and evolve into early-stage tumors. Although early-stage tumors are often androgen sensitive and respond to androgen ablation, a population of androgen independent cells evolve from the hyperplastic population. These cells represent a more advanced form of prostate tumor that may become invasive and potentially become metastatic to the bone, brain, or lung. A variety of genes may be differentially expressed during tumor progression. For example, loss of heterozygosity (LOH) is frequently observed on chromosome 8p in prostate cancer. Fluorescence *in situ* hybridization (FISH) revealed a deletion for at least 1 locus on 8p in 29 (69%) tumors, with a significantly higher frequency of the deletion on 8p21.2-p21.1 in advanced prostate cancer than in localized prostate cancer, implying that deletions on 8p22-p21.3 play an important role in tumor differentiation, while 8p21.2-p21.1 deletion plays a role in progression of prostate cancer (Oba, K. et al. (2001) Cancer Genet. Cytogenet. 124: 20-26).

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A primary diagnostic marker for prostate cancer is prostate specific antigen (PSA). PSA is a tissue-specific serine protease almost exclusively produced by prostatic epithelial cells. The quantity of PSA correlates with the number and volume of the prostatic epithelial cells, and consequently, the levels of PSA are an excellent indicator of abnormal prostate growth. Men with prostate cancer exhibit an early linear increase in PSA levels followed by an exponential increase prior to diagnosis. However, since PSA levels are also influenced by factors such as inflammation, androgen and other growth factors, some scientists maintain that changes in PSA levels are not useful in detecting individual cases of prostate cancer.

Current areas of cancer research provide additional prospects for markers as well as potential therapeutic targets for prostate cancer. Several growth factors have been shown to play a critical role in tumor development, growth, and progression. The growth factors Epidermal Growth Factor (EGF), Fibroblast Growth Factor (FGF), and Tumor Growth Factor alpha (TGFα) are important in the growth of normal as well as hyperproliferative prostate epithelial cells, particularly at early stages of tumor development and progression, and affect signaling pathways in these cells in various ways (Lin, J. et al. (1999) Cancer Res. 59:2891-2897; Putz, T. et al. (1999) Cancer Res. 59:227-233). The TGF-β family of growth factors are generally expressed at increased levels in human cancers and the high expression levels in many cases correlates with advanced stages of malignancy and poor survival

(Gold, L.I. (1999) Crit. Rev. Oncog. 10:303-360). Finally, there are human cell lines representing both the androgen-dependent stage of prostate cancer (LNCap) as well as the androgen-independent, hormone refractory stage of the disease (PC3 and DU-145) that have proved useful in studying gene expression patterns associated with the progression of prostate cancer, and the effects of cell treatments on these expressed genes (Chung, T.D. (1999) Prostate 15:199-207).

Ovarian cancer

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Ovarian cancer is the leading cause of death from a gynecologic cancer. The majority of ovarian cancers are derived from epithelial cells, and 70% of patients with epithelial ovarian cancers present with late-stage disease. As a result, the long-term survival rate for this disease is very low. Identification of early-stage markers for ovarian cancer would significantly increase the survival rate. Genetic variations involved in ovarian cancer development include mutation of p53 and microsatellite instability. Gene expression patterns likely vary when normal ovary is compared to ovarian tumors. Bone cancer

Osteosarcoma is the most common malignant bone tumor in children. Approximately 80% of patients present with non-metastatic disease. After the diagnosis is made by an initial biopsy, treatment involves the use of 3-4 courses of neoadjuvant chemotherapy before definitive surgery, followed by post-operative chemotherapy. With currently available treatment regimens, approximately 30-40% of patients with non-metastatic disease relapse after therapy. Currently, there is no prognostic factor that can be used at the time of initial diagnosis to predict which patients will have a high risk of relapse. The only significant prognostic factor predicting the outcome in a patient with non-metastatic osteosarcoma is the histopathologic response of the primary tumor resected at the time of definitive surgery. The degree of necrosis in the primary tumor is a reflection of the tumor response to neoadjuvant chemotherapy. A higher degree of necrosis (good or favorable response) is associated with a lower risk of relapse and a better outcome. Patients with a lower degree of necrosis (poor or unfavorable response) have a much higher risk of relapse and poor outcome even after complete resection of the primary tumor. Unfortunately, poor outcome cannot be altered despite modification of post-operative chemotherapy to account for the resistance of the primary tumor to neoadjuvant chemotherapy. Thus, there is an urgent need to identify prognostic factors that can be used at the time of diagnosis to recognize the subtypes of osteosarcomas that have various risks of relapse, so that more appropriate chemotherapy can be used at the outset to improve the outcome. Inflammation and immune responses

Atherosclerosis is a pathological condition characterized by a chronic local inflammatory response within the vessel wall of major arteries. Disease progression results in the formation of

atherosclerotic lesions, unstable plaques which occasionally rupture, precipitating a catastrophic thrombotic occlusion of the vessel lumen. Atherosclerosis and the associated coronary artery disease and cerebral stroke represent the most common causes of death in industrialized nations. Although certain key risk factors have been identified, a full molecular characterization that elucidates the causes and identifies all potential therapeutic targets for this complex disease has not been achieved. Molecular characterization of atherosclerosis requires identification of the genes that contribute to lesion growth, stability, dissolution, rupture and induction of occlusive vessel thrombi.

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Blood vessel walls are composed of two tissue layers: an endothelial cell (EC) layer which comprises the lumenal surface of the vessel, and an underlying vascular smooth muscle cell (VSMC) layer. Through dynamic interactions with each other and with surrounding tissues, the vascular endothelium and smooth muscle tissues maintain vascular tone, control selective permeability of the vascular wall, direct vessel remodeling and angiogenesis, and modulate inflammatory and immune responses.

The inflammatory response is a complex vascular reaction mediated by numerous cytokines, chemokines, growth factors, and other signaling molecules expressed by activated ECs, VSMCs and leukocytes. Inflammation protects the organism during trauma and infection, but can also lead to pathological conditions such as atherosclerosis. Activation of vascular endothelium is a central event in a wide range of physiological and disease processes such as vascular tone regulation, coagulation and thrombosis, atherosclerosis, inflammation and some infectious diseases.

The pro-inflammatory cytokines, interleukin (IL)-1 and tumor necrosis factor (TNF), are secreted by a small number of activated macrophages or other cells and can set off a cascade of vascular changes, largely through their ability to alter gene expression patterns in ECs and VSMCs. These vascular changes include vasodilation and increased permeability of microvasculature, edema, and leukocyte extravasation and transmigration across the vessel wall. Ultimately, leukocytes, particularly neutrophils and monocytes/macrophages, accumulate in the extravascular space, where they remove injurious agents by phagocytosis and oxidative killing, a process accompanied by release of toxic factors, such as proteases and reactive oxygen species.

IL-1 and TNF induce pro-inflammatory, thrombotic, and anti-apoptotic changes in gene expression by signaling through receptors on the surface of ECs and VSMCs; these receptors activate transcription factors such as NFkB as well as AP-1, IRF-1, and NF-GMa, leading to alterations in gene expression. Genes known to be differentially regulated in EC by IL-1 and TNF include E selectin, VCAM-1, ICAM-1, PAF, IkBα, IAP-1, MCP-1, eotaxin, ENA-78, G-CSF, A20, ICE, and complement C3 component. A key event in inflammation, adhesion and transmigration of blood

leukocytes across the vascular endothelium, for example, is mediated by increased expression of E selectin, P selectin, ICAM-1, and VCAM-1 on activated endothelium.

Several investigators have examined changes in vascular cell gene expression associated with various inflammatory diseases or model systems. Examining human umbilical vein endothelial cells (HUVEC) activated by recombinant TNF- α or conditioned medium from activated human primary monocytes, Horrevoets et al. (1999; Blood 93:3418-3431) identified 106 differentially regulated genes. In a similar approach, deVries et al. (2000; JBC 275:23939-23947) identified 40 differentially regulated genes in umbilical cord artery-derived smooth muscle cells activated by conditioned media from cultured macrophages after stimulation with oxidized LDL particles. In both studies, many of the identified genes were already known to be involved in inflammation. Comparing expression profiles from inflammatory diseased tissues, cultured macrophages, chondrocyte cell lines, primary chondrocytes, and synoviocytes, Heller et al. (1997; Proc Natl Acad Sci USA 94:2150-2155) identifed candidate genes involved in inflammatory responses, including TNF, IL-1 IL-6, IL-8 G-CSF, RANTES, and V-CAM. From this candidate gene set, tissue inhibitor of metalloproteinase 1, ferritin light chain, and manganese superoxide dismutase were found to be differentially expressed in rheumatoid arthritis (RA) relative to inflammatory bowel disease (IBD). Further, IL-3, chemokine Groa, and metalloproteinase matrix metallo-elastase were expressed in both RA and IBD. Most recently, in an analysis of cultured aortic smooth muscle cells treated with TNF- α , Haley et al. (2000; Circulation 102:2185-2189) found a 20-fold increase in eotaxin, an eosinophil chemotactic factor. The overexpression of eotaxin and its receptor CCR3 in atherosclerotic lesions was confirmed by northern 20 analysis.

Human coronary artery endothelial cells (HCAECs) are primary cells derived from the endothelium of a human coronary artery. HCAECs are used as an experimental model for investigating the role of the endothelium in human vascular biology in vitro. Human umbilical artery endothelial cells (HUAECs) are primary cells derived from the endothelium of an umbilical artery. Human uterine myometrium microvascular endothelial cells (UtMVECs) are primary cells derived from the uterine myometrium microvasculature. Human Iliac Artery Endothelial Cells (HIAECs) are primary cells derived from the endothelium of an iliac artery. Human umbilical vein endothelial cells (HUVECs) are a primary cell line derived from the endothelium of the human umbilical vein. ECV304 is a human endothelial line.

Neurological disorders

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Characterization of region-specific gene expression in the human brain provides a context and background for molecular neurobiology on a variety of neurological disorders. For example,

Alzheimer's disease (AD) is a progressive, neurodestructive process of the human neocortex, characterized by the deterioration of memory and higher cognitive function. A progressive and irreversible brain disorder, AD is characterized by three major pathogenic episodes involving (a) an aberrant processing and deposition of beta-amyloid precursor protein (betaAPP) to form neurotoxic beta-amyloid (betaA) peptides and an aggregated insoluble polymer of betaA that forms the senile plaque, (b) the establishment of intraneuronal neuritic tau pathology yielding widespread deposits of agyrophilic neurofibrillary tangles (NFT) and (c) the initiation and proliferation of a brain-specific inflammatory response. These three seemingly disperse attributes of AD etiopathogenesis are linked by the fact that proinflammatory microglia, reactive astrocytes and their associated cytokines and chemokines are associated with the biology of the microtubule associated protein tau, betaA speciation and aggregation. Missense mutations in the presentlin genes PS1 and PS2, implicated in early onset familial AD, cause abnormal betaAPP processing with resultant overproduction of betaA42 and related neurotoxic peptides. Specific betaA fragments such as betaA42 can further potentiate proinflammatory mechanisms. Expression of the inducible oxidoreductase cyclooxygenase-2 and cytosolic phospholipase A2 (cPLA2) is strongly activated during cerebral ischemia and trauma, epilepsy and AD, indicating the induction of proinflammatory gene pathways as a response to brain injury. Neurotoxic metals such as aluminum and zinc, both implicated in AD etiopathogenesis, and arachidonic acid, a major metabolite of brain cPLA2 activity, each polymerize hyperphosphorylated tau to form NFT-like bundles. Studies have identified a reduced risk for AD in patients aged over 70 years who were previously treated with non-steroidal anti-inflammatory drugs for non-CNS afflictions that include arthritis. (For a review of the interrelationships between the mechanisms of PS1, PS2 and betaAPP gene expression, tau and betaA deposition and the induction, regulation and proliferation in AD of the neuroinflammatory response, see Lukiw, W.J, and Bazan, N.G. (2000) Neurochem. Res. 2000 25:1173-1184).

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Tumor necrosis factor-alpha (TNF- α) is a pleiotropic cytokine that plays a central role in mediation of the inflammatory response through activation of multiple signal transduction pathways. TNF- α is produced by activated lymphocytes, macrophages, and other white blood cells, and activates endothelial cells. Interferon-gamma (IFN γ), also known as Type II interferon or immune interferon, is a cytokine produced primarily by T-lymphocytes and natural killer cells. Mature IFN γ exists as noncovalently-linked homodimers. IFN γ displays antiviral, antiproliferative, immunoregulatory, and proinflammatory activities and is important in host defense mechanisms. IFN- γ induces the production of cytokines; upregulates the expression of class I and II MHC antigens, Fc receptor, and leukocyte adhesion molecule; modulates macrophage effector functions; influences isotype switching; potentiates

the secretion of immunoglobulins by B cells; augments TH1 cell expansion; and may be required for TH1 cell differentiation. IFNγ exerts its biological activities by binding to specific cell surface receptors, which display high affinity binding sites. The IFNγ receptor is present on almost all cell types except mature erythrocytes. Upon binding to its receptor, IFNγ triggers the activation of JAK-1 and JAK-2 kinases resulting in the phosphorylation of STAT1. Both IFNγ and TNF-α are considered proinflammatory cytokines. Cross-talk can exist between the signal transduction pathways of two cytokines; for example, signal transduction cascades initiated by two different cytokines lead to the activation of NfkB.

Liver toxicity

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The human C3A cell line is a clonal derivative of HepG2/C3 (hepatoma cell line, isolated from a 15-year-old male with liver tumor), which was selected for strong contact inhibition of growth. The use of a clonal population enhances the reproducibility of the cells. C3A cells have many characteristics of primary human hepatocytes in culture: i) expression of insulin receptor and insulin-like growth factor II receptor; ii) secretion of a high ratio of serum albumin compared with α-fetoprotein; iii) conversion of ammonia to urea and glutamine; iv) metabolism of aromatic amino acids; and v) proliferation in glucose-free and insulin-free medium. The C3A cell line is now well established as an *in vitro* model of the mature human liver (Mickelson et al. (1995) Hepatology 22:866-875; Nagendra et al. (1997) Am. J. Physiol. 272:G408-G416).

The potential application of gene expression profiling is relevant to measuring the toxic response to potential therapeutic compounds and of the metabolic response to therapeutic agents. For instance, diseases treated with steroids and disorders caused by the metabolic response to treatment with steroids include adenomatosis, cholestasis, cirrhosis, hemangioma, Henoch-Schonlein purpura, hepatitis, hepatocellular and metastatic carcinomas, idiopathic thrombocytopenic purpura, porphyria, sarcoidosis, and Wilson disease. It is desirable to measure the toxic response to potential therapeutic compounds and of the metabolic response to therapeutic agents.

Steroids are a class of lipid-soluble molecules, including cholesterol, bile acids, vitamin D, and hormones, that share a common four-ring structure based on cyclopentanoperhydrophenanthrene and that carrry out a wide variety of functions. Steroid hormones, produced by the adrenal cortex, ovaries, and testes, include glucocorticoids, mineralocorticoids, androgens, and estrogens. Steroid hormones are widely used for fertility control and in anti-inflammatory treatments for physical injuries and diseases such as arthritis, asthma, and auto-immune disorders. Progesterone, a naturally occurring progestin, is primarily used to treat amenorrhea, abnormal uterine bleeding, or as a contraceptive.

Medroxyprogesterone (MAH), also known as 6 α -methyl-17-hydroxyprogesterone, is a synthetic progestin with a pharmacological activity about 15 times greater than progesterone. MAH is usually used for the treatment of renal and endometrial carcinomas, amenorrhea, abnormal uterine bleeding, and endometriosis associated with hormonal imbalance. The primary contraceptive effect of exogenous progestins involves the suppression of the midcycle surge of LH. The exact mechanism of action, however, is unknown. At the cellular level, progestins diffuse freely into target cells and bind to the progesterone receptor. Target cells include the female reproductive tract, mammary gland, hypothalamus, and pituitary. Once bound to the receptor, progestins slow the frequency of release of gonadotropin releasing hormone (GnRH) from the hypothalamus and blunt the pre-ovulatory LH surge, thereby preventing follicular maturation and ovulation. Interestingly, the MAH stimulatory effect on the respiratory centers has been used clinically to treat low blood oxygenation due to sleep apnea, chronic obstructive pulmonary disease, or hypercapnia (excess of CO 2 in blood). Beclomethasone is a synthetic glucocorticoid that is used for treating steroid-dependent asthma, relieving symptoms associated with allergic or nonallergic (vasomotor) rhinitis, or for preventing recurrent nasal polyps following surgical removal. The anti-inflammatory and vasoconstrictive effects of intranasal beclomethasone are 5,000 times greater than those produced by hydrocortisone. Budesonide is a corticosteroid used to control symptoms associated with allergic rhinitis or asthma. Dexamethasone is a synthetic glucocorticoid used in anti-inflammatory or immunosuppressive compositions. Prednisone is metabolized in the liver to its active form, prednisolone, a glucocorticoid with anti-inflammatory properties. Betamethasone is a synthetic glucocorticoid with antiinflammatory and immunosuppressive activity and is used to treat psoriasis and fungal infections, such as athlete's foot and ringworm. By comparing both the levels and sequences expressed in tissues from subjects exposed to or treated with steroid compounds with the levels and sequences expressed in normal untreated tissue it is possible to determine tissue responses to steroids. Budesonide (Bude) is a corticosteroid used to control symptoms associated with allergic rhinitis or asthma. Budesonide has high topical anti-inflammatory activity but low systemic activity. Prednisone is a corticosteroid that is metabolized in the liver to its active form, prednisolone. Prednisone is roughly four times more potent as a glucocorticoid than hydrocortisone. Prednisone is intermediate between hydrocortisone and dexamethasone in duration of action. Prednisone is used in conditions such as allograft rejection, asthma, systemic lupus erythematosus, and many other inflammatory states.

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Glucocorticoids are naturally occurring hormones that prevent or suppress inflammation and immune responses when administered at pharmacological doses. At the molecular level, unbound

glucocorticoids readily cross cell membranes and bind with high affinity to specific cytoplasmic receptors. Subsequent to binding, transcription and, ultimately, protein synthesis are affected. The result can include inhibition of leukocyte infiltration at the site of inflammation, interference in the function of mediators of inflammatory response, and suppression of humoral immune responses. The anti-inflammatory actions of corticosteroids are thought to involve phospholipase A2 inhibitory proteins, collectively called lipocortins. Lipocortins, in turn, control the biosynthesis of potent mediators of inflammation such as prostaglandins and leukotrienes by inhibiting the release of the precursor molecule arachidonic.

There is a need in the art for new compositions, including nucleic acids and proteins, for the diagnosis, prevention, and treatment of transport, neurological, muscle, immunological and cell proliferative disorders.

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SUMMARY OF THE INVENTION

Various embodiments of the invention provide purified polypeptides, transporters and ion channels, referred to collectively as 'TRICH' and individually as 'TRICH-1,' 'TRICH-2,' 'TRICH-3,' 'TRICH-4,' 'TRICH-5,' 'TRICH-6,' 'TRICH-7,' 'TRICH-8,' 'TRICH-9,' 'TRICH-10,' 'TRICH-11,' 'TRICH-13,' 'TRICH-13,' 'TRICH-14,' 'TRICH-15,' 'TRICH-16,' 'TRICH-17,' 'TRICH-18,' 'TRICH-19,' 'TRICH-20,' 'TRICH-21,' 'TRICH-22,' 'TRICH-23,' 'TRICH-24,' 'TRICH-25,' and 'TRICH-26' and methods for using these proteins and their encoding polynucleotides for the detection, diagnosis, and treatment of diseases and medical conditions. Embodiments also provide methods for utilizing the purified transporters and ion channels and/or their encoding polynucleotides for facilitating the drug discovery process, including determination of efficacy, dosage, toxicity, and pharmacology. Related embodiments provide methods for utilizing the purified transporters and ion channels and/or their encoding polynucleotides for investigating the pathogenesis of diseases and medical conditions.

An embodiment provides an isolated polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26. Another embodiment provides an isolated polypeptide comprising an amino acid sequence of SEQ ID NO:1-26.

Still another embodiment provides an isolated polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26. In another embodiment, the polynucleotide encodes a polypeptide selected from the group consisting of SEQ ID NO:1-26. In an alternative embodiment, the polynucleotide is selected from the group consisting of SEQ ID NO:1-52.

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Still another embodiment provides a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26. Another embodiment provides a cell transformed with the recombinant polynucleotide. Yet another embodiment provides a transgenic organism comprising the recombinant polynucleotide.

Another embodiment provides a method for producing a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26. The method comprises a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding the polypeptide, and b) recovering the polypeptide so expressed.

Yet another embodiment provides an isolated antibody which specifically binds to a

polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26.

Still yet another embodiment provides an isolated polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical or at least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). In other embodiments, the polynucleotide can comprise at least about 20, 30, 40, 60, 80, or 100 contiguous nucleotides.

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Yet another embodiment provides a method for detecting a target polynucleotide in a sample, said target polynucleotide being selected from the group consisting of a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical or at least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide or fragments thereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and b) detecting the presence or absence of said hybridization complex. In a related embodiment, the method can include detecting the amount of the hybridization complex. In still other embodiments, the probe can comprise at least about 20, 30, 40, 60, 80, or 100 contiguous nucleotides.

Still yet another embodiment provides a method for detecting a target polynucleotide in a sample, said target polynucleotide being selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, b) a

polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical or at least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof. In a related embodiment, the method can include detecting the amount of the amplified target polynucleotide or fragment thereof.

Another embodiment provides a composition comprising an effective amount of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and a pharmaceutically acceptable excipient. In one embodiment, the composition can comprise an amino acid sequence selected from the group consisting of SEQ ID NO:1-26. Other embodiments provide a method of treating a disease or condition associated with decreased or abnormal expression of functional TRICH, comprising administering to a patient in need of such treatment the composition.

Yet another embodiment provides a method for screening a compound for effectiveness as an agonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting agonist activity in the sample. Another embodiment provides a composition comprising an agonist compound identified by the method and a pharmaceutically acceptable excipient. Yet another embodiment provides a method of treating a disease or condition associated with decreased expression of functional TRICH, comprising

administering to a patient in need of such treatment the composition.

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Still yet another embodiment provides a method for screening a compound for effectiveness as an antagonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting antagonist activity in the sample. Another embodiment provides a composition comprising an antagonist compound identified by the method and a pharmaceutically acceptable excipient. Yet another embodiment provides a method of treating a disease or condition associated with overexpression of functional TRICH, comprising administering to a patient in need of such treatment the composition.

Another embodiment provides a method of screening for a compound that specifically binds to a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26. The method comprises a) combining the polypeptide with at least one test compound under suitable conditions, and b) detecting binding of the polypeptide to the test compound, thereby identifying a compound that specifically binds to the polypeptide.

Yet another embodiment provides a method of screening for a compound that modulates the activity of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence

selected from the group consisting of SEQ ID NO:1-26. The method comprises a) combining the polypeptide with at least one test compound under conditions permissive for the activity of the polypeptide, b) assessing the activity of the polypeptide in the presence of the test compound, and c) comparing the activity of the polypeptide in the presence of the test compound with the activity of the polypeptide in the absence of the test compound, wherein a change in the activity of the polypeptide in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide.

Still yet another embodiment provides a method for screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, the method comprising a) exposing a sample comprising the target polynucleotide to a compound, b) detecting altered expression of the target polynucleotide, and c) comparing the expression of the target polynucleotide in the presence of varying amounts of the compound and in the absence of the compound.

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Another embodiment provides a method for assessing toxicity of a test compound, said method comprising a) treating a biological sample containing nucleic acids with the test compound; b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, ii) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical or at least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, iii) a polynucleotide having a sequence complementary to i), iv) a polynucleotide complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Hybridization occurs under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, ii) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical or at least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, iii) a polynucleotide complementary to the polynucleotide of i), iv) a polynucleotide complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Alternatively, the target polynucleotide can comprise a fragment of a polynucleotide selected from the group consisting of i)-v) above; c) quantifying the amount of hybridization complex; and d) comparing

the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

BRIEF DESCRIPTION OF THE TABLES

Table 1 summarizes the nomenclature for full length polynucleotide and polypeptide embodiments of the invention.

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Table 2 shows the GenBank identification number and annotation of the nearest GenBank homolog, and the PROTEOME database identification numbers and annotations of PROTEOME database homologs, for polypeptide embodiments of the invention. The probability scores for the matches between each polypeptide and its homolog(s) are also shown.

Table 3 shows structural features of polypeptide embodiments, including predicted motifs and domains, along with the methods, algorithms, and searchable databases used for analysis of the polypeptides.

Table 4 lists the cDNA and/or genomic DNA fragments which were used to assemble polynucleotide embodiments, along with selected fragments of the polynucleotides.

Table 5 shows representative cDNA libraries for polynucleotide embodiments.

Table 6 provides an appendix which describes the tissues and vectors used for construction of the cDNA libraries shown in Table 5.

Table 7 shows the tools, programs, and algorithms used to analyze polynucleotides and polypeptides, along with applicable descriptions, references, and threshold parameters.

Table 8 shows single nucleotide polymorphisms found in polynucleotide sequences of the invention, along with allele frequencies in different human populations.

DESCRIPTION OF THE INVENTION

Before the present proteins, nucleic acids, and methods are described, it is understood that embodiments of the invention are not limited to the particular machines, instruments, materials, and methods described, as these may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the invention.

As used herein and in the appended claims, the singular forms "a," "an," and "the" include plural reference unless the context clearly dictates otherwise. Thus, for example, a reference to "a

host cell" includes a plurality of such host cells, and a reference to "an antibody" is a reference to one or more antibodies and equivalents thereof known to those skilled in the art, and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meanings as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any machines, materials, and methods similar or equivalent to those described herein can be used to practice or test the present invention, the preferred machines, materials and methods are now described. All publications mentioned herein are cited for the purpose of describing and disclosing the cell lines, protocols, reagents and vectors which are reported in the publications and which might be used in connection with various embodiments of the invention. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue of prior invention.

DEFINITIONS

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"TRICH" refers to the amino acid sequences of substantially purified TRICH obtained from any species, particularly a mammalian species, including bovine, ovine, porcine, murine, equine, and human, and from any source, whether natural, synthetic, semi-synthetic, or recombinant.

The term "agonist" refers to a molecule which intensifies or mimics the biological activity of TRICH. Agonists may include proteins, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of TRICH either by directly interacting with TRICH or by acting on components of the biological pathway in which TRICH participates.

An "allelic variant" is an alternative form of the gene encoding TRICH. Allelic variants may result from at least one mutation in the nucleic acid sequence and may result in altered mRNAs or in polypeptides whose structure or function may or may not be altered. A gene may have none, one, or many allelic variants of its naturally occurring form. Common mutational changes which give rise to allelic variants are generally ascribed to natural deletions, additions, or substitutions of nucleotides. Each of these types of changes may occur alone, or in combination with the others, one or more times in a given sequence.

"Altered" nucleic acid sequences encoding TRICH include those sequences with deletions, insertions, or substitutions of different nucleotides, resulting in a polypeptide the same as TRICH or a polypeptide with at least one functional characteristic of TRICH. Included within this definition are polymorphisms which may or may not be readily detectable using a particular oligonucleotide probe of the polynucleotide encoding TRICH, and improper or unexpected hybridization to allelic variants, with a locus other than the normal chromosomal locus for the polynucleotide encoding TRICH. The encoded protein may also be "altered," and may contain deletions, insertions, or substitutions of amino

acid residues which produce a silent change and result in a functionally equivalent TRICH. Deliberate amino acid substitutions may be made on the basis of one or more similarities in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues, as long as the biological or immunological activity of TRICH is retained. For example, negatively charged amino acids may include aspartic acid and glutamic acid, and positively charged amino acids may include lysine and arginine. Amino acids with uncharged polar side chains having similar hydrophilicity values may include: asparagine and glutamine; and serine and threonine. Amino acids with uncharged side chains having similar hydrophilicity values may include: leucine, isoleucine, and valine; glycine and alanine; and phenylalanine and tyrosine.

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The terms "amino acid" and "amino acid sequence" can refer to an oligopeptide, a peptide, a polypeptide, or a protein sequence, or a fragment of any of these, and to naturally occurring or synthetic molecules. Where "amino acid sequence" is recited to refer to a sequence of a naturally occurring protein molecule, "amino acid sequence" and like terms are not meant to limit the amino acid sequence to the complete native amino acid sequence associated with the recited protein molecule.

"Amplification" relates to the production of additional copies of a nucleic acid. Amplification may be carried out using polymerase chain reaction (PCR) technologies or other nucleic acid amplification technologies well known in the art.

The term "antagonist" refers to a molecule which inhibits or attenuates the biological activity of TRICH. Antagonists may include proteins such as antibodies, anticalins, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of TRICH either by directly interacting with TRICH or by acting on components of the biological pathway in which TRICH participates.

The term "antibody" refers to intact immunoglobulin molecules as well as to fragments thereof, such as Fab, F(ab')₂, and Fv fragments, which are capable of binding an epitopic determinant. Antibodies that bind TRICH polypeptides can be prepared using intact polypeptides or using fragments containing small peptides of interest as the immunizing antigen. The polypeptide or oligopeptide used to immunize an animal (e.g., a mouse, a rat, or a rabbit) can be derived from the translation of RNA, or synthesized chemically, and can be conjugated to a carrier protein if desired. Commonly used carriers that are chemically coupled to peptides include bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin (KLH). The coupled peptide is then used to immunize the animal.

The term "antigenic determinant" refers to that region of a molecule (i.e., an epitope) that makes contact with a particular antibody. When a protein or a fragment of a protein is used to

immunize a host animal, numerous regions of the protein may induce the production of antibodies which bind specifically to antigenic determinants (particular regions or three-dimensional structures on the protein). An antigenic determinant may compete with the intact antigen (i.e., the immunogen used to elicit the immune response) for binding to an antibody.

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The term "aptamer" refers to a nucleic acid or oligonucleotide molecule that binds to a specific molecular target. Aptamers are derived from an *in vitro* evolutionary process (e.g., SELEX (Systematic Evolution of Ligands by EXponential Enrichment), described in U.S. Patent No. 5,270,163), which selects for target-specific aptamer sequences from large combinatorial libraries. Aptamer compositions may be double-stranded or single-stranded, and may include deoxyribonucleotides, ribonucleotides, nucleotide derivatives, or other nucleotide-like molecules. The nucleotide components of an aptamer may have modified sugar groups (e.g., the 2'-OH group of a ribonucleotide may be replaced by 2'-F or 2'-NH₂), which may improve a desired property, e.g., resistance to nucleases or longer lifetime in blood. Aptamers may be conjugated to other molecules, e.g., a high molecular weight carrier to slow clearance of the aptamer from the circulatory system. Aptamers may be specifically cross-linked to their cognate ligands, e.g., by photo-activation of a cross-linker (Brody, E.N. and L. Gold (2000) J. Biotechnol. 74:5-13).

The term "intramer" refers to an aptamer which is expressed *in vivo*. For example, a vaccinia virus-based RNA expression system has been used to express specific RNA aptamers at high levels in the cytoplasm of leukocytes (Blind, M. et al. (1999) Proc. Natl. Acad. Sci. USA 96:3606-3610).

The term "spiegelmer" refers to an aptamer which includes L-DNA, L-RNA, or other left-handed nucleotide derivatives or nucleotide-like molecules. Aptamers containing left-handed nucleotides are resistant to degradation by naturally occurring enzymes, which normally act on substrates containing right-handed nucleotides.

The term "antisense" refers to any composition capable of base-pairing with the "sense" (coding) strand of a polynucleotide having a specific nucleic acid sequence. Antisense compositions may include DNA; RNA; peptide nucleic acid (PNA); oligonucleotides having modified backbone linkages such as phosphorothioates, methylphosphonates, or benzylphosphonates; oligonucleotides having modified sugar groups such as 2'-methoxyethyl sugars or 2'-methoxyethoxy sugars; or oligonucleotides having modified bases such as 5-methyl cytosine, 2'-deoxyuracil, or 7-deaza-2'-deoxyguanosine. Antisense molecules may be produced by any method including chemical synthesis or transcription. Once introduced into a cell, the complementary antisense molecule base-pairs with a

naturally occurring nucleic acid sequence produced by the cell to form duplexes which block either transcription or translation. The designation "negative" or "minus" can refer to the antisense strand, and the designation "positive" or "plus" can refer to the sense strand of a reference DNA molecule.

The term "biologically active" refers to a protein having structural, regulatory, or biochemical functions of a naturally occurring molecule. Likewise, "immunologically active" or "immunogenic" refers to the capability of the natural, recombinant, or synthetic TRICH, or of any oligopeptide thereof, to induce a specific immune response in appropriate animals or cells and to bind with specific antibodies.

"Complementary" describes the relationship between two single-stranded nucleic acid sequences that anneal by base-pairing. For example, 5'-AGT-3' pairs with its complement, 3'-TCA-5'.

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A "composition comprising a given polynucleotide" and a "composition comprising a given polypeptide" can refer to any composition containing the given polynucleotide or polypeptide. The composition may comprise a dry formulation or an aqueous solution. Compositions comprising polynucleotides encoding TRICH or fragments of TRICH may be employed as hybridization probes. The probes may be stored in freeze-dried form and may be associated with a stabilizing agent such as a carbohydrate. In hybridizations, the probe may be deployed in an aqueous solution containing salts (e.g., NaCl), detergents (e.g., sodium dodecyl sulfate; SDS), and other components (e.g., Denhardt's solution, dry milk, salmon sperm DNA, etc.).

"Consensus sequence" refers to a nucleic acid sequence which has been subjected to repeated DNA sequence analysis to resolve uncalled bases, extended using the XL-PCR kit (Applied Biosystems, Foster City CA) in the 5' and/or the 3' direction, and resequenced, or which has been assembled from one or more overlapping cDNA, EST, or genomic DNA fragments using a computer program for fragment assembly, such as the GELVIEW fragment assembly system (GCG, Madison WI) or Phrap (University of Washington, Seattle WA). Some sequences have been both extended and assembled to produce the consensus sequence.

"Conservative amino acid substitutions" are those substitutions that are predicted to least interfere with the properties of the original protein, i.e., the structure and especially the function of the protein is conserved and not significantly changed by such substitutions. The table below shows amino acids which may be substituted for an original amino acid in a protein and which are regarded as conservative amino acid substitutions.

	Original Residue	Conservative Substitution
	Ala	Gly, Ser
	Arg	His, Lys
	Asn	Asp, Gln, His
5	Asp	Asn, Glu
	Cys	Ala, Ser
	Gln	Asn, Glu, His
	Glu	Asp, Gln, His
	Gly	Ala
10	His	Asn, Arg, Gln, Glu
	Ile	Leu, Val
	Leu	Ile, Val
	Lys	Arg, Gln, Glu
	Met	Leu, Ile
15	Phe	His, Met, Leu, Trp, Tyr
	Ser	Cys, Thr
	Thr	Ser, Val
	Trp	Phe, Tyr
	Tyr	His, Phe, Trp
20	Val	Ile, Leu, Thr

Conservative amino acid substitutions generally maintain (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a beta sheet or alpha helical conformation, (b) the charge or hydrophobicity of the molecule at the site of the substitution, and/or (c) the bulk of the side chain.

A "deletion" refers to a change in the amino acid or nucleotide sequence that results in the absence of one or more amino acid residues or nucleotides.

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The term "derivative" refers to a chemically modified polynucleotide or polypeptide. Chemical modifications of a polynucleotide can include, for example, replacement of hydrogen by an alkyl, acyl, hydroxyl, or amino group. A derivative polynucleotide encodes a polypeptide which retains at least one biological or immunological function of the natural molecule. A derivative polypeptide is one modified by glycosylation, pegylation, or any similar process that retains at least one biological or immunological function of the polypeptide from which it was derived.

A "detectable label" refers to a reporter molecule or enzyme that is capable of generating a measurable signal and is covalently or noncovalently joined to a polynucleotide or polypeptide.

"Differential expression" refers to increased or upregulated; or decreased, downregulated, or absent gene or protein expression, determined by comparing at least two different samples. Such comparisons may be carried out between, for example, a treated and an untreated sample, or a diseased and a normal sample.

"Exon shuffling" refers to the recombination of different coding regions (exons). Since an exon may represent a structural or functional domain of the encoded protein, new proteins may be assembled through the novel reassortment of stable substructures, thus allowing acceleration of the evolution of new protein functions.

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A "fragment" is a unique portion of TRICH or a polynucleotide encoding TRICH which can be identical in sequence to, but shorter in length than, the parent sequence. A fragment may comprise up to the entire length of the defined sequence, minus one nucleotide/amino acid residue. For example, a fragment may comprise from about 5 to about 1000 contiguous nucleotides or amino acid residues. A fragment used as a probe, primer, antigen, therapeutic molecule, or for other purposes, may be at least 5, 10, 15, 16, 20, 25, 30, 40, 50, 60, 75, 100, 150, 250 or at least 500 contiguous nucleotides or amino acid residues in length. Fragments may be preferentially selected from certain regions of a molecule. For example, a polypeptide fragment may comprise a certain length of contiguous amino acids selected from the first 250 or 500 amino acids (or first 25% or 50%) of a polypeptide as shown in a certain defined sequence. Clearly these lengths are exemplary, and any length that is supported by the specification, including the Sequence Listing, tables, and figures, may be encompassed by the present embodiments.

A fragment of SEQ ID NO:27-52 can comprise a region of unique polynucleotide sequence that specifically identifies SEQ ID NO:27-52, for example, as distinct from any other sequence in the genome from which the fragment was obtained. A fragment of SEQ ID NO:27-52 can be employed in one or more embodiments of methods of the invention, for example, in hybridization and amplification technologies and in analogous methods that distinguish SEQ ID NO:27-52 from related polynucleotides. The precise length of a fragment of SEQ ID NO:27-52 and the region of SEQ ID NO:27-52 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A fragment of SEQ ID NO:1-26 is encoded by a fragment of SEQ ID NO:27-52. A fragment of SEQ ID NO:1-26 can comprise a region of unique amino acid sequence that specifically identifies SEQ ID NO:1-26. For example, a fragment of SEQ ID NO:1-26 can be used as an immunogenic peptide for the development of antibodies that specifically recognize SEQ ID NO:1-26. The precise length of a fragment of SEQ ID NO:1-26 and the region of SEQ ID NO:1-26 to which the fragment corresponds can be determined based on the intended purpose for the fragment using one or more analytical methods described herein or otherwise known in the art.

A "full length" polynucleotide is one containing at least a translation initiation codon (e.g.,

methionine) followed by an open reading frame and a translation termination codon. A "full length" polynucleotide sequence encodes a "full length" polypeptide sequence.

"Homology" refers to sequence similarity or, interchangeably, sequence identity, between two or more polynucleotide sequences or two or more polypeptide sequences.

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The terms "percent identity" and "% identity," as applied to polynucleotide sequences, refer to the percentage of residue matches between at least two polynucleotide sequences aligned using a standardized algorithm. Such an algorithm may insert, in a standardized and reproducible way, gaps in the sequences being compared in order to optimize alignment between two sequences, and therefore achieve a more meaningful comparison of the two sequences.

Percent identity between polynucleotide sequences may be determined using one or more computer algorithms or programs known in the art or described herein. For example, percent identity can be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program. This program is part of the LASERGENE software package, a suite of molecular biological analysis programs (DNASTAR, Madison WI). CLUSTAL V is described in Higgins, D.G. and P.M. Sharp (1989; CABIOS 5:151-153) and in Higgins, D.G. et al. (1992; CABIOS 8:189-191). For pairwise alignments of polynucleotide sequences, the default parameters are set as follows: Ktuple=2, gap penalty=5, window=4, and "diagonals saved"=4. The "weighted" residue weight table is selected as the default. Percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polynucleotide sequences.

Alternatively, a suite of commonly used and freely available sequence comparison algorithms which can be used is provided by the National Center for Biotechnology Information (NCBI) Basic Local Alignment Search Tool (BLAST) (Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410), which is available from several sources, including the NCBI, Bethesda, MD, and on the Internet at http://www.ncbi.nlm.nih.gov/BLAST/. The BLAST software suite includes various sequence analysis programs including "blastn," that is used to align a known polynucleotide sequence with other polynucleotide sequences from a variety of databases. Also available is a tool called "BLAST 2 Sequences" that is used for direct pairwise comparison of two nucleotide sequences. "BLAST 2 Sequences" can be accessed and used interactively at http://www.ncbi.nlm.nih.gov/gorf/bl2.html. The "BLAST 2 Sequences" tool can be used for both blastn and blastp (discussed below). BLAST programs are commonly used with gap and other parameters set to default settings. For example, to compare two nucleotide sequences, one may use blastn with the "BLAST 2 Sequences" tool Version

2.0.12 (April-21-2000) set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Reward for match: 1

Penalty for mismatch: -2

Open Gap: 5 and Extension Gap: 2 penalties

Gap x drop-off: 50

Expect: 10

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Word Size: 11

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Percent identity may be measured over the length of an entire defined sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined sequence, for instance, a fragment of at least 20, at least 30, at least 40, at least 50, at least 70, at least 100, or at least 200 contiguous nucleotides. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures, or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

Nucleic acid sequences that do not show a high degree of identity may nevertheless encode similar amino acid sequences due to the degeneracy of the genetic code. It is understood that changes in a nucleic acid sequence can be made using this degeneracy to produce multiple nucleic acid sequences that all encode substantially the same protein.

The phrases "percent identity" and "% identity," as applied to polypeptide sequences, refer to the percentage of residue matches between at least two polypeptide sequences aligned using a standardized algorithm. Methods of polypeptide sequence alignment are well-known. Some alignment methods take into account conservative amino acid substitutions. Such conservative substitutions, explained in more detail above, generally preserve the charge and hydrophobicity at the site of substitution, thus preserving the structure (and therefore function) of the polypeptide.

Percent identity between polypeptide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program (described and referenced above). For pairwise alignments of polypeptide sequences using CLUSTAL V, the default parameters are set as follows: Ktuple=1, gap penalty=3, window=5, and "diagonals saved"=5. The PAM250 matrix is selected as the default residue weight table. As with polynucleotide alignments, the percent identity is reported by

CLUSTAL V as the "percent similarity" between aligned polypeptide sequence pairs.

Alternatively the NCBI BLAST software suite may be used. For example, for a pairwise comparison of two polypeptide sequences, one may use the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) with blastp set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Open Gap: 11 and Extension Gap: 1 penalties

Gap x drop-off: 50

Expect: 10

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Word Size: 3

Filter: on

Percent identity may be measured over the length of an entire defined polypeptide sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined polypeptide sequence, for instance, a fragment of at least 15, at least 20, at least 30, at least 40, at least 50, at least 70 or at least 150 contiguous residues. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

"Human artificial chromosomes" (HACs) are linear microchromosomes which may contain DNA sequences of about 6 kb to 10 Mb in size and which contain all of the elements required for chromosome replication, segregation and maintenance.

The term "humanized antibody" refers to an antibody molecule in which the amino acid sequence in the non-antigen binding regions has been altered so that the antibody more closely resembles a human antibody, and still retains its original binding ability.

"Hybridization" refers to the process by which a polynucleotide strand anneals with a complementary strand through base pairing under defined hybridization conditions. Specific hybridization is an indication that two nucleic acid sequences share a high degree of complementarity. Specific hybridization complexes form under permissive annealing conditions and remain hybridized after the "washing" step(s). The washing step(s) is particularly important in determining the stringency of the hybridization process, with more stringent conditions allowing less non-specific binding, i.e., binding between pairs of nucleic acid strands that are not perfectly matched. Permissive conditions for annealing of nucleic acid sequences are routinely determinable by one of ordinary skill in

the art and may be consistent among hybridization experiments, whereas wash conditions may be varied among experiments to achieve the desired stringency, and therefore hybridization specificity. Permissive annealing conditions occur, for example, at 68°C in the presence of about 6 x SSC, about 1% (w/v) SDS, and about $100 \mu g/ml$ sheared, denatured salmon sperm DNA.

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Generally, stringency of hybridization is expressed, in part, with reference to the temperature under which the wash step is carried out. Such wash temperatures are typically selected to be about 5°C to 20°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. An equation for calculating T_m and conditions for nucleic acid hybridization are well known and can be found in Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual, 2^{nd} ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; specifically see volume 2, chapter 9.

High stringency conditions for hybridization between polynucleotides of the present invention include wash conditions of 68°C in the presence of about 0.2 x SSC and about 0.1% SDS, for 1 hour. Alternatively, temperatures of about 65°C, 60°C, 55°C, or 42°C may be used. SSC concentration may be varied from about 0.1 to 2 x SSC, with SDS being present at about 0.1%. Typically, blocking reagents are used to block non-specific hybridization. Such blocking reagents include, for instance, sheared and denatured salmon sperm DNA at about 100-200 μ g/ml. Organic solvent, such as formamide at a concentration of about 35-50% v/v, may also be used under particular circumstances, such as for RNA:DNA hybridizations. Useful variations on these wash conditions will be readily apparent to those of ordinary skill in the art. Hybridization, particularly under high stringency conditions, may be suggestive of evolutionary similarity between the nucleotides. Such similarity is strongly indicative of a similar role for the nucleotides and their encoded polypeptides.

The term "hybridization complex" refers to a complex formed between two nucleic acids by virtue of the formation of hydrogen bonds between complementary bases. A hybridization complex may be formed in solution (e.g., C₀t or R₀t analysis) or formed between one nucleic acid present in solution and another nucleic acid immobilized on a solid support (e.g., paper, membranes, filters, chips, pins or glass slides, or any other appropriate substrate to which cells or their nucleic acids have been fixed).

The words "insertion" and "addition" refer to changes in an amino acid or polynucleotide sequence resulting in the addition of one or more amino acid residues or nucleotides, respectively.

"Immune response" can refer to conditions associated with inflammation, trauma, immune

disorders, or infectious or genetic disease, etc. These conditions can be characterized by expression of various factors, e.g., cytokines, chemokines, and other signaling molecules, which may affect cellular and systemic defense systems.

An "immunogenic fragment" is a polypeptide or oligopeptide fragment of TRICH which is capable of eliciting an immune response when introduced into a living organism, for example, a mammal. The term "immunogenic fragment" also includes any polypeptide or oligopeptide fragment of TRICH which is useful in any of the antibody production methods disclosed herein or known in the art.

The term "microarray" refers to an arrangement of a plurality of polynucleotides, polypeptides, antibodies, or other chemical compounds on a substrate.

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The terms "element" and "array element" refer to a polynucleotide, polypeptide, antibody, or other chemical compound having a unique and defined position on a microarray.

The term "modulate" refers to a change in the activity of TRICH. For example, modulation may cause an increase or a decrease in protein activity, binding characteristics, or any other biological, functional, or immunological properties of TRICH.

The phrases "nucleic acid" and "nucleic acid sequence" refer to a nucleotide, oligonucleotide, polynucleotide, or any fragment thereof. These phrases also refer to DNA or RNA of genomic or synthetic origin which may be single-stranded or double-stranded and may represent the sense or the antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material.

"Operably linked" refers to the situation in which a first nucleic acid sequence is placed in a functional relationship with a second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding sequence. Operably linked DNA sequences may be in close proximity or contiguous and, where necessary to join two protein coding regions, in the same reading frame.

"Peptide nucleic acid" (PNA) refers to an antisense molecule or anti-gene agent which comprises an oligonucleotide of at least about 5 nucleotides in length linked to a peptide backbone of amino acid residues ending in lysine. The terminal lysine confers solubility to the composition. PNAs preferentially bind complementary single stranded DNA or RNA and stop transcript elongation, and may be pegylated to extend their lifespan in the cell.

"Post-translational modification" of an TRICH may involve lipidation, glycosylation, phosphorylation, acetylation, racemization, proteolytic cleavage, and other modifications known in the art. These processes may occur synthetically or biochemically. Biochemical modifications will vary

by cell type depending on the enzymatic milieu of TRICH.

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"Probe" refers to nucleic acids encoding TRICH, their complements, or fragments thereof, which are used to detect identical, allelic or related nucleic acids. Probes are isolated oligonucleotides or polynucleotides attached to a detectable label or reporter molecule. Typical labels include radioactive isotopes, ligands, chemiluminescent agents, and enzymes. "Primers" are short nucleic acids, usually DNA oligonucleotides, which may be annealed to a target polynucleotide by complementary base-pairing. The primer may then be extended along the target DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification (and identification) of a nucleic acid, e.g., by the polymerase chain reaction (PCR).

Probes and primers as used in the present invention typically comprise at least 15 contiguous nucleotides of a known sequence. In order to enhance specificity, longer probes and primers may also be employed, such as probes and primers that comprise at least 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, or at least 150 consecutive nucleotides of the disclosed nucleic acid sequences. Probes and primers may be considerably longer than these examples, and it is understood that any length supported by the specification, including the tables, figures, and Sequence Listing, may be used.

Methods for preparing and using probes and primers are described in the references, for example Sambrook, J. et al. (1989; Molecular Cloning: A Laboratory Manual, 2nd ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY), Ausubel, F.M. et al. (1999; Short Protocols in Molecular Biology, 4th ed., John Wiley & Sons, New York NY), and Innis, M. et al. (1990; PCR Protocols, A Guide to Methods and Applications, Academic Press, San Diego CA). PCR primer pairs can be derived from a known sequence, for example, by using computer programs intended for that purpose such as Primer (Version 0.5, 1991, Whitehead Institute for Biomedical Research, Cambridge MA).

Oligonucleotides for use as primers are selected using software known in the art for such purpose. For example, OLIGO 4.06 software is useful for the selection of PCR primer pairs of up to 100 nucleotides each, and for the analysis of oligonucleotides and larger polynucleotides of up to 5,000 nucleotides from an input polynucleotide sequence of up to 32 kilobases. Similar primer selection programs have incorporated additional features for expanded capabilities. For example, the PrimOU primer selection program (available to the public from the Genome Center at University of Texas South West Medical Center, Dallas TX) is capable of choosing specific primers from megabase sequences and is thus useful for designing primers on a genome-wide scope. The Primer3 primer selection program (available to the public from the Whitehead Institute/MIT Center for Genome Research, Cambridge MA) allows the user to input a "mispriming library," in which sequences to

avoid as primer binding sites are user-specified. Primer3 is useful, in particular, for the selection of oligonucleotides for microarrays. (The source code for the latter two primer selection programs may also be obtained from their respective sources and modified to meet the user's specific needs.) The PrimeGen program (available to the public from the UK Human Genome Mapping Project Resource Centre, Cambridge UK) designs primers based on multiple sequence alignments, thereby allowing selection of primers that hybridize to either the most conserved or least conserved regions of aligned nucleic acid sequences. Hence, this program is useful for identification of both unique and conserved oligonucleotides and polynucleotide fragments. The oligonucleotides and polynucleotide fragments identified by any of the above selection methods are useful in hybridization technologies, for example, as PCR or sequencing primers, microarray elements, or specific probes to identify fully or partially complementary polynucleotides in a sample of nucleic acids. Methods of oligonucleotide selection are not limited to those described above.

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A "recombinant nucleic acid" is a nucleic acid that is not naturally occurring or has a sequence that is made by an artificial combination of two or more otherwise separated segments of sequence. This artificial combination is often accomplished by chemical synthesis or, more commonly, by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques such as those described in Sambrook, *supra*. The term recombinant includes nucleic acids that have been altered solely by addition, substitution, or deletion of a portion of the nucleic acid. Frequently, a recombinant nucleic acid may include a nucleic acid sequence operably linked to a promoter sequence. Such a recombinant nucleic acid may be part of a vector that is used, for example, to transform a cell.

Alternatively, such recombinant nucleic acids may be part of a viral vector, e.g., based on a vaccinia virus, that could be use to vaccinate a mammal wherein the recombinant nucleic acid is expressed, inducing a protective immunological response in the mammal.

A "regulatory element" refers to a nucleic acid sequence usually derived from untranslated regions of a gene and includes enhancers, promoters, introns, and 5' and 3' untranslated regions (UTRs). Regulatory elements interact with host or viral proteins which control transcription, translation, or RNA stability.

"Reporter molecules" are chemical or biochemical moieties used for labeling a nucleic acid, amino acid, or antibody. Reporter molecules include radionuclides; enzymes; fluorescent, chemiluminescent, or chromogenic agents; substrates; cofactors; inhibitors; magnetic particles; and other moieties known in the art.

An "RNA equivalent," in reference to a DNA molecule, is composed of the same linear sequence of nucleotides as the reference DNA molecule with the exception that all occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

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The term "sample" is used in its broadest sense. A sample suspected of containing TRICH, nucleic acids encoding TRICH, or fragments thereof may comprise a bodily fluid; an extract from a cell, chromosome, organelle, or membrane isolated from a cell; a cell; genomic DNA, RNA, or cDNA, in solution or bound to a substrate; a tissue; a tissue print; etc.

The terms "specific binding" and "specifically binding" refer to that interaction between a protein or peptide and an agonist, an antibody, an antagonist, a small molecule, or any natural or synthetic binding composition. The interaction is dependent upon the presence of a particular structure of the protein, e.g., the antigenic determinant or epitope, recognized by the binding molecule. For example, if an antibody is specific for epitope "A," the presence of a polypeptide comprising the epitope A, or the presence of free unlabeled A, in a reaction containing free labeled A and the antibody will reduce the amount of labeled A that binds to the antibody.

The term "substantially purified" refers to nucleic acid or amino acid sequences that are removed from their natural environment and are isolated or separated, and are at least about 60% free, preferably at least about 75% free, and most preferably at least about 90% free from other components with which they are naturally associated.

A "substitution" refers to the replacement of one or more amino acid residues or nucleotides by different amino acid residues or nucleotides, respectively.

"Substrate" refers to any suitable rigid or semi-rigid support including membranes, filters, chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, tubing, plates, polymers, microparticles and capillaries. The substrate can have a variety of surface forms, such as wells, trenches, pins, channels and pores, to which polynucleotides or polypeptides are bound.

A "transcript image" or "expression profile" refers to the collective pattern of gene expression by a particular cell type or tissue under given conditions at a given time.

"Transformation" describes a process by which exogenous DNA is introduced into a recipient cell. Transformation may occur under natural or artificial conditions according to various methods well known in the art, and may rely on any known method for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell. The method for transformation is selected based on the type of host cell being transformed and may include, but is not limited to, bacteriophage or viral

infection, electroporation, heat shock, lipofection, and particle bombardment. The term "transformed cells" includes stably transformed cells in which the inserted DNA is capable of replication either as an autonomously replicating plasmid or as part of the host chromosome, as well as transiently transformed cells which express the inserted DNA or RNA for limited periods of time.

A "transgenic organism," as used herein, is any organism, including but not limited to animals and plants, in which one or more of the cells of the organism contains heterologous nucleic acid introduced by way of human intervention, such as by transgenic techniques well known in the art. The nucleic acid is introduced into the cell, directly or indirectly by introduction into a precursor of the cell, by way of deliberate genetic manipulation, such as by microinjection or by infection with a recombinant virus. In another embodiment, the nucleic acid can be introduced by infection with a recombinant viral vector, such as a lentiviral vector (Lois, C. et al. (2002) Science 295:868-872). The term genetic manipulation does not include classical cross-breeding, or *in vitro* fertilization, but rather is directed to the introduction of a recombinant DNA molecule. The transgenic organisms contemplated in accordance with the present invention include bacteria, cyanobacteria, fungi, plants and animals. The isolated DNA of the present invention can be introduced into the host by methods known in the art, for example infection, transfection, transformation or transconjugation. Techniques for transferring the DNA of the present invention into such organisms are widely known and provided in references such as Sambrook et al. (1989), *supra*.

A "variant" of a particular nucleic acid sequence is defined as a nucleic acid sequence having at least 40% sequence identity to the particular nucleic acid sequence over a certain length of one of the nucleic acid sequences using blastn with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of nucleic acids may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity over a certain defined length. A variant may be described as, for example, an "allelic" (as defined above), "splice," "species," or "polymorphic" variant. A splice variant may have significant identity to a reference molecule, but will generally have a greater or lesser number of polynucleotides due to alternate splicing of exons during mRNA processing. The corresponding polypeptide may possess additional functional domains or lack domains that are present in the reference molecule. Species variants are polynucleotides that vary from one species to another. The resulting polypeptides will generally have significant amino acid identity relative to each other. A polymorphic variant is a variation in the polynucleotide sequence of a particular gene between

individuals of a given species. Polymorphic variants also may encompass "single nucleotide polymorphisms" (SNPs) in which the polynucleotide sequence varies by one nucleotide base. The presence of SNPs may be indicative of, for example, a certain population, a disease state, or a propensity for a disease state.

A "variant" of a particular polypeptide sequence is defined as a polypeptide sequence having at least 40% sequence identity to the particular polypeptide sequence over a certain length of one of the polypeptide sequences using blastp with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of polypeptides may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity over a certain defined length of one of the polypeptides.

THE INVENTION

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Various embodiments of the invention include new human transporters and ion channels (TRICH), the polynucleotides encoding TRICH, and the use of these compositions for the diagnosis, treatment, or prevention of transport, neurological, muscle, immunological and cell proliferative disorders.

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide embodiments of the invention. Each polynucleotide and its corresponding polypeptide are correlated to a single Incyte project identification number (Incyte Project ID). Each polypeptide sequence is denoted by both a polypeptide sequence identification number (Polypeptide SEQ ID NO:) and an Incyte polypeptide sequence number (Incyte Polypeptide ID) as shown. Each polynucleotide sequence is denoted by both a polynucleotide sequence identification number (Polynucleotide SEQ ID NO:) and an Incyte polynucleotide consensus sequence number (Incyte Polynucleotide ID) as shown. Column 6 shows the Incyte ID numbers of physical, full length clones corresponding to the polypeptide and polynucleotide sequences of the invention. The full length clones encode polypeptides which have at least 95% sequence identity to the polypeptide sequences shown in column 3.

Table 2 shows sequences with homology to the polypeptides of the invention as identified by BLAST analysis against the GenBank protein (genpept) database and the PROTEOME database. Columns 1 and 2 show the polypeptide sequence identification number (Polypeptide SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for polypeptides of the invention. Column 3 shows the GenBank identification number (GenBank ID NO:) of the nearest

GenBank homolog and the PROTEOME database identification numbers (PROTEOME ID NO:) of the nearest PROTEOME database homologs. Column 4 shows the probability scores for the matches between each polypeptide and its homolog(s). Column 5 shows the annotation of the GenBank and PROTEOME database homolog(s) along with relevant citations where applicable, all of which are expressly incorporated by reference herein.

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Table 3 shows various structural features of the polypeptides of the invention. Columns 1 and 2 show the polypeptide sequence identification number (SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for each polypeptide of the invention. Column 3 shows the number of amino acid residues in each polypeptide. Column 4 shows potential phosphorylation sites, and column 5 shows potential glycosylation sites, as determined by the MOTIFS program of the GCG sequence analysis software package (Genetics Computer Group, Madison WI). Column 6 shows amino acid residues comprising signature sequences, domains, and motifs. Column 7 shows analytical methods for protein structure/function analysis and in some cases, searchable databases to which the analytical methods were applied.

Together, Tables 2 and 3 summarize the properties of polypeptides of the invention, and these properties establish that the claimed polypeptides are transporters and ion channels. For example, SEQ ID NO:1 is 49% identical, from residue S11 to residue K626, to human CTL1 protein (GenBank ID g6996442) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 9.0e-168, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:1 also contains an eight transmembrane helices regions as determined by using a hidden Markov model for the prediction of transmembrane helices. (See Table 3.) In an alternative example, SEQ ID NO:3 is 57% identical, from residue E10 to residue V115, to human SLC11A3 iron transporter (GenBank ID g8895485) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 4.7e-25, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. In an alternative example, SEQ ID NO:6 is 88% identical, from residue M1 to residue S944, to rat potassium channel (GenBank ID g2745729) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 0.0, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:6 also contains a PAC motif, a PAS domain, a cyclic nucleotide-binding domain, and an ion transport protein domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)based PFAM database of conserved protein family domains. (See Table 3.) Data from

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BLAST PRODOM and BLAST DOMO analyses provide further corroborative evidence that SEQ ID NO:6 is a potassium channel. In an alternative example, SEQ ID NO:10 is 99% identical, from residue M1 to residue I418, 95% identical, from residue S420 to residue S680, and 94% identical, from residue P665 to residue H894, to human Eag-related gene member 2 (GenBank ID g11878259) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 0.0, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:10 is localized to the plasma membrane, has transporter and channel activity and is a voltage-gated potassium channel, as determined by BLAST analysis using the PROTEOME database. SEQ ID NO:10 also contains a PAC domain, cyclic nucleotide binding domain and ion transport domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) SEQ ID NO:10 contains five transmembrane-spanning regions as determined by TMHMMR analysis. Data from further BLAST analyses of the PRODOM and DOMO databases provide additional corroborative evidence that SEQ ID NO:10 is a potassium channel. In an alternative example, SEQ ID NO:11 is 86% identical, from residue A94 to residue S785, to rat potassium channel (GenBank ID g2745729) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 0.0, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. As determined by BLAST analysis using the PROTEOME database, SEQ ID NO:11 is localized to the plasma membrane, is homologous to rat ether-a-go-go related 2, which is a slowly activating delayed rectifier potassium channel, and may facilitate the differentiation of pre-vertebral neurons (PROTEOME ID 331276|Rn.10875); SEQ ID NO:11 is also homologous to rat ether-a-go-go-related gene 3 which is an inward rectifier potassium channel that functions in potassium transport specifically in the nervous system (PROTEOME ID 331274|Rn.10874). SEQ ID NO:11 also contains a cyclic nucleotide-binding domain and an ion transport protein domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from additional BLAST analyses provide further corroborative evidence that SEQ ID NO:11 is a potassium channel. In an alternative example, SEQ ID NO:14 is 38% identical, from residue Q13 to residue S1049, to Schizosaccharomyces pombe membrane ATPase (GenBank ID g3451312) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 1.5e-189, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:14 is

localized to the membrane, and is a member of the P-type, Ca2+-type, ATPase subfamily, as determined by BLAST analysis using the PROTEOME database. SEQ ID NO:14 also contains an E1-E2 ATPase domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:14 is a membrane ATPase. In an alternative example, SEQ ID NO:19 is 98% identical, from residue M1 to residue L602, to human sodium-dependent high-affinity dicarboxylate transporter (GenBank ID g8132324) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 0.0, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:19 also has homology to proteins that have transporter gene function and are sodium-dependent dicarboxylate transporters, as determined by BLAST analysis using the PROTEOME database. SEQ ID NO:19 also contains a sodium-dependent dicarboxylate transporter domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS and BLAST analyses provide further corroborative evidence that SEQ ID NO:19 is a sodium-dependent dicarboxylate transporter. SEQ ID NO:2, SEQ ID NO:4,-5, SEQ ID NO:7-9, SEQ ID NO:12-13, SEQ ID NO:15-18, and SEQ ID NO:20-26 were analyzed and annotated in a similar manner. The algorithms and parameters for the analysis of SEO ID NO:1-26 are described in Table 7.

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As shown in Table 4, the full length polynucleotide embodiments were assembled using cDNA sequences or coding (exon) sequences derived from genomic DNA, or any combination of these two types of sequences. Column 1 lists the polynucleotide sequence identification number (Polynucleotide SEQ ID NO:), the corresponding Incyte polynucleotide consensus sequence number (Incyte ID) for each polynucleotide of the invention, and the length of each polynucleotide sequence in basepairs. Column 2 shows the nucleotide start (5') and stop (3') positions of the cDNA and/or genomic sequences used to assemble the full length polynucleotide embodiments, and of fragments of the polynucleotides which are useful, for example, in hybridization or amplification technologies that identify SEQ ID NO:8-14 or that distinguish between SEQ ID NO:8-14 and related polynucleotides.

The polynucleotide fragments described in Column 2 of Table 4 may refer specifically, for example, to Incyte cDNAs derived from tissue-specific cDNA libraries or from pooled cDNA libraries. Alternatively, the polynucleotide fragments described in column 2 may refer to GenBank cDNAs or ESTs which contributed to the assembly of the full length polynucleotides. In addition, the

polynucleotide fragments described in column 2 may identify sequences derived from the ENSEMBL (The Sanger Centre, Cambridge, UK) database (i.e., those sequences including the designation "ENST"). Alternatively, the polynucleotide fragments described in column 2 may be derived from the NCBI RefSeq Nucleotide Sequence Records Database (i.e., those sequences including the designation "NM" or "NT") or the NCBI RefSeq Protein Sequence Records (i.e., those sequences including the designation "NP"). Alternatively, the polynucleotide fragments described in column 2 may refer to assemblages of both cDNA and Genscan-predicted exons brought together by an "exon stitching" algorithm. For example, a polynucleotide sequence identified as $FL_XXXXXX_N_1_N_2_YYYYY_N_3_N_4$ represents a "stitched" sequence in which XXXXXX is the identification number of the cluster of sequences to which the algorithm was applied, and YYYYY is the 10 number of the prediction generated by the algorithm, and $N_{1,2,3...}$, if present, represent specific exons that may have been manually edited during analysis (See Example V). Alternatively, the polynucleotide fragments in column 2 may refer to assemblages of exons brought together by an "exon-stretching" algorithm. For example, a polynucleotide sequence identified as FLXXXXXX_gAAAAA_gBBBBB_1_N is a "stretched" sequence, with XXXXXX being the Incyte 15 project identification number, gAAAAA being the GenBank identification number of the human genomic sequence to which the "exon-stretching" algorithm was applied, gBBBBB being the GenBank identification number or NCBI RefSeq identification number of the nearest GenBank protein homolog, and N referring to specific exons (See Example V). In instances where a RefSeq sequence was used as a protein homolog for the "exon-stretching" algorithm, a RefSeq identifier (denoted by "NM," 20

Alternatively, a prefix identifies component sequences that were hand-edited, predicted from genomic DNA sequences, or derived from a combination of sequence analysis methods. The following Table lists examples of component sequence prefixes and corresponding sequence analysis methods associated with the prefixes (see Example IV and Example V).

"NP," or "NT") may be used in place of the GenBank identifier (i.e., gBBBBB).

Prefix	Type of analysis and/or examples of programs	
GNN, GFG,	Exon prediction from genomic sequences using, for example,	
ENST	GENSCAN (Stanford University, CA, USA) or FGENES	
	(Computer Genomics Group, The Sanger Centre, Cambridge, UK).	
GBI	Hand-edited analysis of genomic sequences.	
FL	Stitched or stretched genomic sequences (see Example V).	

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INCY	Full length transcript and exon prediction from mapping of EST	
	sequences to the genome. Genomic location and EST composition	
	data are combined to predict the exons and resulting transcript.	

In some cases, Incyte cDNA coverage redundant with the sequence coverage shown in Table 4 was obtained to confirm the final consensus polynucleotide sequence, but the relevant Incyte cDNA identification numbers are not shown.

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Table 5 shows the representative cDNA libraries for those full length polynucleotides which were assembled using Incyte cDNA sequences. The representative cDNA library is the Incyte cDNA library which is most frequently represented by the Incyte cDNA sequences which were used to assemble and confirm the above polynucleotides. The tissues and vectors which were used to construct the cDNA libraries shown in Table 5 are described in Table 6.

Table 8 shows single nucleotide polymorphisms (SNPs) found in polynucleotide sequences of the invention, along with allele frequencies in different human populations. Columns 1 and 2 show the polynucleotide sequence identification number (SEQ ID NO:) and the corresponding Incyte project identification number (PID) for polynucleotides of the invention. Column 3 shows the Incyte identification number for the EST in which the SNP was detected (EST ID), and column 4 shows the identification number for the SNP (SNP ID). Column 5 shows the position within the EST sequence at which the SNP is located (EST SNP), and column 6 shows the position of the SNP within the full-length polynucleotide sequence (CB1 SNP). Column 7 shows the allele found in the EST sequence. Columns 8 and 9 show the two alleles found at the SNP site. Column 10 shows the amino acid encoded by the codon including the SNP site, based upon the allele found in the EST. Columns 11-14 show the frequency of allele 1 in four different human populations. An entry of n/d (not detected) indicates that the frequency of allele 1 in the population was too low to be detected, while n/a (not available) indicates that the allele frequency was not determined for the population.

The invention also encompasses TRICH variants. A preferred TRICH variant is one which has at least about 80%, or alternatively at least about 90%, or even at least about 95% amino acid sequence identity to the TRICH amino acid sequence, and which contains at least one functional or structural characteristic of TRICH.

Various embodiments also encompass polynucleotides which encode TRICH. In a particular embodiment, the invention encompasses a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:27-52, which encodes TRICH. The polynucleotide

sequences of SEQ ID NO:27-52, as presented in the Sequence Listing, embrace the equivalent RNA sequences, wherein occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

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The invention also encompasses variants of a polynucleotide encoding TRICH. In particular, such a variant polynucleotide will have at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to a polynucleotide encoding TRICH. A particular aspect of the invention encompasses a variant of a polynucleotide comprising a sequence selected from the group consisting of SEQ ID NO:27-52 which has at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to a nucleic acid sequence selected from the group consisting of SEQ ID NO:27-52. Any one of the polynucleotide variants described above can encode a polypeptide which contains at least one functional or structural characteristic of TRICH.

In addition, or in the alternative, a polynucleotide variant of the invention is a splice variant of a polynucleotide encoding TRICH. A splice variant may have portions which have significant sequence identity to a polynucleotide encoding TRICH, but will generally have a greater or lesser number of polynucleotides due to additions or deletions of blocks of sequence arising from alternate splicing of exons during mRNA processing. A splice variant may have less than about 70%, or alternatively less than about 60%, or alternatively less than about 50% polynucleotide sequence identity to a polynucleotide encoding TRICH over its entire length; however, portions of the splice variant will have at least about 70%, or alternatively at least about 85%, or alternatively at least about 95%, or alternatively 100% polynucleotide sequence identity to portions of the polynucleotide encoding TRICH. For example, a polynucleotide comprising a sequence of SEQ ID NO:34 and a polynucleotide comprising a sequence of SEQ ID NO:43 are splice variants of each other; a polynucleotide comprising a sequence of SEQ ID NO:46 and a polynucleotide comprising a sequence of SEQ ID NO:52 are splice variants of each other; a polynucleotide comprising a sequence of SEQ ID NO:39 and a polynucleotide comprising a sequence of SEQ ID NO:50 are splice variants of each other; and a polynucleotide comprising a sequence of SEQ ID NO:32, a polynucleotide comprising a sequence of SEQ ID NO:36, and a polynucleotide comprising a sequence of SEQ ID NO:37 are splice variants of each other. Any one of the splice variants described above can encode a polypeptide which contains at least one functional or structural characteristic of TRICH.

It will be appreciated by those skilled in the art that as a result of the degeneracy of the genetic code, a multitude of polynucleotide sequences encoding TRICH, some bearing minimal

similarity to the polynucleotide sequences of any known and naturally occurring gene, may be produced. Thus, the invention contemplates each and every possible variation of polynucleotide sequence that could be made by selecting combinations based on possible codon choices. These combinations are made in accordance with the standard triplet genetic code as applied to the polynucleotide sequence of naturally occurring TRICH, and all such variations are to be considered as being specifically disclosed.

Although polynucleotides which encode TRICH and its variants are generally capable of hybridizing to polynucleotides encoding naturally occurring TRICH under appropriately selected conditions of stringency, it may be advantageous to produce polynucleotides encoding TRICH or its derivatives possessing a substantially different codon usage, e.g., inclusion of non-naturally occurring codons. Codons may be selected to increase the rate at which expression of the peptide occurs in a particular prokaryotic or eukaryotic host in accordance with the frequency with which particular codons are utilized by the host. Other reasons for substantially altering the nucleotide sequence encoding TRICH and its derivatives without altering the encoded amino acid sequences include the production of RNA transcripts having more desirable properties, such as a greater half-life, than transcripts produced from the naturally occurring sequence.

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The invention also encompasses production of polynucleotides which encode TRICH and TRICH derivatives, or fragments thereof, entirely by synthetic chemistry. After production, the synthetic polynucleotide may be inserted into any of the many available expression vectors and cell systems using reagents well known in the art. Moreover, synthetic chemistry may be used to introduce mutations into a polynucleotide encoding TRICH or any fragment thereof.

Embodiments of the invention can also include polynucleotides that are capable of hybridizing to the claimed polynucleotides, and, in particular, to those having the sequences shown in SEQ ID NO:27-52 and fragments thereof, under various conditions of stringency (Wahl, G.M. and S.L. Berger (1987) Methods Enzymol. 152:399-407; Kimmel, A.R. (1987) Methods Enzymol. 152:507-511). Hybridization conditions, including annealing and wash conditions, are described in "Definitions."

Methods for DNA sequencing are well known in the art and may be used to practice any of the embodiments of the invention. The methods may employ such enzymes as the Klenow fragment of DNA polymerase I, SEQUENASE (US Biochemical, Cleveland OH), Taq polymerase (Applied Biosystems), thermostable T7 polymerase (Amersham Biosciences, Piscataway NJ), or combinations of polymerases and proofreading exonucleases such as those found in the ELONGASE amplification system (Invitrogen, Carlsbad CA). Preferably, sequence preparation is automated with machines

such as the MICROLAB 2200 liquid transfer system (Hamilton, Reno NV), PTC200 thermal cycler (MJ Research, Watertown MA) and ABI CATALYST 800 thermal cycler (Applied Biosystems). Sequencing is then carried out using either the ABI 373 or 377 DNA sequencing system (Applied Biosystems), the MEGABACE 1000 DNA sequencing system (Amersham Biosciences), or other systems known in the art. The resulting sequences are analyzed using a variety of algorithms which are well known in the art (Ausubel et al., *supra*, ch. 7; Meyers, R.A. (1995) Molecular Biology and Biotechnology, Wiley VCH, New York NY, pp. 856-853).

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The nucleic acids encoding TRICH may be extended utilizing a partial nucleotide sequence and employing various PCR-based methods known in the art to detect upstream sequences, such as promoters and regulatory elements. For example, one method which may be employed, restriction-site PCR, uses universal and nested primers to amplify unknown sequence from genomic DNA within a cloning vector (Sarkar, G. (1993) PCR Methods Applic. 2:318-322). Another method, inverse PCR, uses primers that extend in divergent directions to amplify unknown sequence from a circularized template. The template is derived from restriction fragments comprising a known genomic locus and surrounding sequences (Triglia, T. et al. (1988) Nucleic Acids Res. 16:8186). A third method, capture PCR, involves PCR amplification of DNA fragments adjacent to known sequences in human and yeast artificial chromosome DNA (Lagerstrom, M. et al. (1991) PCR Methods Applic. 1:111-119). In this method, multiple restriction enzyme digestions and ligations may be used to insert an engineered double-stranded sequence into a region of unknown sequence before performing PCR. Other methods which may be used to retrieve unknown sequences are known in the art (Parker, J.D. et al. (1991) Nucleic Acids Res. 19:3055-3060). Additionally, one may use PCR, nested primers, and PROMOTERFINDER libraries (Clontech, Palo Alto CA) to walk genomic DNA. This procedure avoids the need to screen libraries and is useful in finding intron/exon junctions. For all PCR-based methods, primers may be designed using commercially available software, such as OLIGO 4.06 primer analysis software (National Biosciences, Plymouth MN) or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the template at temperatures of about 68°C to 72°C.

When screening for full length cDNAs, it is preferable to use libraries that have been size-selected to include larger cDNAs. In addition, random-primed libraries, which often include sequences containing the 5' regions of genes, are preferable for situations in which an oligo d(T) library does not yield a full-length cDNA. Genomic libraries may be useful for extension of sequence into 5' non-transcribed regulatory regions.

Capillary electrophoresis systems which are commercially available may be used to analyze the size or confirm the nucleotide sequence of sequencing or PCR products. In particular, capillary sequencing may employ flowable polymers for electrophoretic separation, four different nucleotide-specific, laser-stimulated fluorescent dyes, and a charge coupled device camera for detection of the emitted wavelengths. Output/light intensity may be converted to electrical signal using appropriate software (e.g., GENOTYPER and SEQUENCE NAVIGATOR, Applied Biosystems), and the entire process from loading of samples to computer analysis and electronic data display may be computer controlled. Capillary electrophoresis is especially preferable for sequencing small DNA fragments which may be present in limited amounts in a particular sample.

In another embodiment of the invention, polynucleotides or fragments thereof which encode TRICH may be cloned in recombinant DNA molecules that direct expression of TRICH, or fragments or functional equivalents thereof, in appropriate host cells. Due to the inherent degeneracy of the genetic code, other polynucleotides which encode substantially the same or a functionally equivalent polypeptides may be produced and used to express TRICH.

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The polynucleotides of the invention can be engineered using methods generally known in the art in order to alter TRICH-encoding sequences for a variety of purposes including, but not limited to, modification of the cloning, processing, and/or expression of the gene product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. For example, oligonucleotide-mediated site-directed mutagenesis may be used to introduce mutations that create new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, and so forth.

The nucleotides of the present invention may be subjected to DNA shuffling techniques such as MOLECULARBREEDING (Maxygen Inc., Santa Clara CA; described in U.S. Patent No. 5,837,458; Chang, C.-C. et al. (1999) Nat. Biotechnol. 17:793-797; Christians, F.C. et al. (1999) Nat. Biotechnol. 17:259-264; and Crameri, A. et al. (1996) Nat. Biotechnol. 14:315-319) to alter or improve the biological properties of TRICH, such as its biological or enzymatic activity or its ability to bind to other molecules or compounds. DNA shuffling is a process by which a library of gene variants is produced using PCR-mediated recombination of gene fragments. The library is then subjected to selection or screening procedures that identify those gene variants with the desired properties. These preferred variants may then be pooled and further subjected to recursive rounds of DNA shuffling and selection/screening. Thus, genetic diversity is created through "artificial" breeding and rapid molecular evolution. For example, fragments of a single gene containing random point mutations may be

recombined, screened, and then reshuffled until the desired properties are optimized. Alternatively, fragments of a given gene may be recombined with fragments of homologous genes in the same gene family, either from the same or different species, thereby maximizing the genetic diversity of multiple naturally occurring genes in a directed and controllable manner.

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In another embodiment, polynucleotides encoding TRICH may be synthesized, in whole or in part, using one or more chemical methods well known in the art (Caruthers, M.H. et al. (1980) Nucleic Acids Symp. Ser. 7:215-223; Horn, T. et al. (1980) Nucleic Acids Symp. Ser. 7:225-232). Alternatively, TRICH itself or a fragment thereof may be synthesized using chemical methods known in the art. For example, peptide synthesis can be performed using various solution-phase or solid-phase techniques (Creighton, T. (1984) Proteins, Structures and Molecular Properties, WH Freeman, New York NY, pp. 55-60; Roberge, J.Y. et al. (1995) Science 269:202-204). Automated synthesis may be achieved using the ABI 431A peptide synthesizer (Applied Biosystems). Additionally, the amino acid sequence of TRICH, or any part thereof, may be altered during direct synthesis and/or combined with sequences from other proteins, or any part thereof, to produce a variant polypeptide or a polypeptide having a sequence of a naturally occurring polypeptide.

The peptide may be substantially purified by preparative high performance liquid chromatography (Chiez, R.M. and F.Z. Regnier (1990) Methods Enzymol. 182:392-421). The composition of the synthetic peptides may be confirmed by amino acid analysis or by sequencing (Creighton, *supra*, pp. 28-53).

In order to express a biologically active TRICH, the polynucleotides encoding TRICH or derivatives thereof may be inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for transcriptional and translational control of the inserted coding sequence in a suitable host. These elements include regulatory sequences, such as enhancers, constitutive and inducible promoters, and 5' and 3' untranslated regions in the vector and in polynucleotides encoding TRICH. Such elements may vary in their strength and specificity. Specific initiation signals may also be used to achieve more efficient translation of polynucleotides encoding TRICH. Such signals include the ATG initiation codon and adjacent sequences, e.g. the Kozak sequence. In cases where a polynucleotide sequence encoding TRICH and its initiation codon and upstream regulatory sequences are inserted into the appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a fragment thereof, is inserted, exogenous translational control signals including an in-frame ATG initiation codon should be provided by the vector. Exogenous translational elements and initiation codons may be of various

origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers appropriate for the particular host cell system used (Scharf, D. et al. (1994) Results Probl. Cell Differ. 20:125-162).

Methods which are well known to those skilled in the art may be used to construct expression vectors containing polynucleotides encoding TRICH and appropriate transcriptional and translational control elements. These methods include *in vitro* recombinant DNA techniques, synthetic techniques, and *in vivo* genetic recombination (Sambrook, J. et al. (1989) Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Press, Plainview NY, ch. 4, 8, and 16-17; Ausubel et al., *supra*, ch. 1, 3, and 15).

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A variety of expression vector/host systems may be utilized to contain and express polynucleotides encoding TRICH. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with viral expression vectors (e.g., baculovirus); plant cell systems transformed with viral expression vectors (e.g., cauliflower mosaic virus, CaMV, or tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or animal cell systems (Sambrook, supra; Ausubel et al., supra; Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509; Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945; Takamatsu, N. (1987) EMBO J. 6:307-311; The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196; Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659; Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355). Expression vectors derived from retroviruses, adenoviruses, or herpes or vaccinia viruses, or from various bacterial plasmids, may be used for delivery of polynucleotides to the targeted organ, tissue, or cell population (Di Nicola, M. et al. (1998) Cancer Gen. Ther. 5:350-356; Yu, M. et al. (1993) Proc. Natl. Acad. Sci. USA 90:6340-6344; Buller, R.M. et al. (1985) Nature 317:813-815; McGregor, D.P. et al. (1994) Mol. Immunol. 31:219-226; Verma, I.M. and N. Somia (1997) Nature 389:239-242). The invention is not limited by the host cell employed.

In bacterial systems, a number of cloning and expression vectors may be selected depending upon the use intended for polynucleotides encoding TRICH. For example, routine cloning, subcloning, and propagation of polynucleotides encoding TRICH can be achieved using a multifunctional *E. coli* vector such as PBLUESCRIPT (Stratagene, La Jolla CA) or PSPORT1 plasmid (Invitrogen). Ligation of polynucleotides encoding TRICH into the vector's multiple cloning site disrupts the *lacZ*.

gene, allowing a colorimetric screening procedure for identification of transformed bacteria containing recombinant molecules. In addition, these vectors may be useful for *in vitro* transcription, dideoxy sequencing, single strand rescue with helper phage, and creation of nested deletions in the cloned sequence (Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509). When large quantities of TRICH are needed, e.g. for the production of antibodies, vectors which direct high level expression of TRICH may be used. For example, vectors containing the strong, inducible SP6 or T7 bacteriophage promoter may be used.

Yeast expression systems may be used for production of TRICH. A number of vectors containing constitutive or inducible promoters, such as alpha factor, alcohol oxidase, and PGH promoters, may be used in the yeast *Saccharomyces cerevisiae* or *Pichia pastoris*. In addition, such vectors direct either the secretion or intracellular retention of expressed proteins and enable integration of foreign polynucleotide sequences into the host genome for stable propagation (Ausubel et al., *supra*; Bitter, G.A. et al. (1987) Methods Enzymol. 153:516-544; Scorer, C.A. et al. (1994) Bio/Technology 12:181-184).

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Plant systems may also be used for expression of TRICH. Transcription of polynucleotides encoding TRICH may be driven by viral promoters, e.g., the 35S and 19S promoters of CaMV used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. (1987) EMBO J. 6:307-311). Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used (Coruzzi, G. et al. (1984) EMBO J. 3:1671-1680; Broglie, R. et al. (1984) Science 224:838-843; Winter, J. et al. (1991) Results Probl. Cell Differ. 17:85-105). These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection (The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196).

In mammalian cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, polynucleotides encoding TRICH may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain infective virus which expresses TRICH in host cells (Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659). In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells. SV40 or EBV-based vectors may also be used for high-level protein expression.

Human artificial chromosomes (HACs) may also be employed to deliver larger fragments of

DNA than can be contained in and expressed from a plasmid. HACs of about 6 kb to 10 Mb are constructed and delivered via conventional delivery methods (liposomes, polycationic amino polymers, or vesicles) for therapeutic purposes (Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355).

For long term production of recombinant proteins in mammalian systems, stable expression of TRICH in cell lines is preferred. For example, polynucleotides encoding TRICH can be transformed into cell lines using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for about 1 to 2 days in enriched media before being switched to selective media. The purpose of the selectable marker is to confer resistance to a selective agent, and its presence allows growth and recovery of cells which successfully express the introduced sequences. Resistant clones of stably transformed cells may be propagated using tissue culture techniques appropriate to the cell type.

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Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase and adenine phosphoribosyltransferase genes, for use in *tk* and *apr* cells, respectively (Wigler, M. et al. (1977) Cell 11:223-232; Lowy, I. et al. (1980) Cell 22:817-823). Also, antimetabolite, antibiotic, or herbicide resistance can be used as the basis for selection. For example, *dhfr* confers resistance to methotrexate; *neo* confers resistance to the aminoglycosides neomycin and G-418; and *als* and *pat* confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively (Wigler, M. et al. (1980) Proc. Natl. Acad. Sci. USA 77:3567-3570; Colbere-Garapin, F. et al. (1981) J. Mol. Biol. 150:1-14). Additional selectable genes have been described, e.g., *trpB* and *hisD*, which alter cellular requirements for metabolites (Hartman, S.C. and R.C. Mulligan (1988) Proc. Natl. Acad. Sci. USA 85:8047-8051). Visible markers, e.g., anthocyanins, green fluorescent proteins (GFP; Clontech), β-glucuronidase and its substrate β-glucuronide, or luciferase and its substrate luciferin may be used. These markers can be used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system (Rhodes, C.A. (1995) Methods Mol. Biol. 55:121-131).

Although the presence/absence of marker gene expression suggests that the gene of interest is also present, the presence and expression of the gene may need to be confirmed. For example, if the sequence encoding TRICH is inserted within a marker gene sequence, transformed cells containing polynucleotides encoding TRICH can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a sequence encoding TRICH under the

control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

In general, host cells that contain the polynucleotide encoding TRICH and that express TRICH may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations, PCR amplification, and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein sequences.

Immunological methods for detecting and measuring the expression of TRICH using either specific polyclonal or monoclonal antibodies are known in the art. Examples of such techniques include enzyme-linked immunosorbent assays (ELISAs), radioimmunoassays (RIAs), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on TRICH is preferred, but a competitive binding assay may be employed. These and other assays are well known in the art (Hampton, R. et al. (1990) Serological Methods, a Laboratory Manual, APS Press, St. Paul MN, Sect. IV; Coligan, J.E. et al. (1997) Current Protocols in Immunology, Greene Pub. Associates and Wiley-Interscience, New York NY; Pound, J.D. (1998) Immunochemical Protocols, Humana Press, Totowa NJ).

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A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides encoding TRICH include oligolabeling, nick translation, end-labeling, or PCR amplification using a labeled nucleotide. Alternatively, polynucleotides encoding TRICH, or any fragments thereof, may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes *in vitro* by addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits, such as those provided by Amersham Biosciences, Promega (Madison WI), and US Biochemical. Suitable reporter molecules or labels which may be used for ease of detection include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents, as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

Host cells transformed with polynucleotides encoding TRICH may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a transformed cell may be secreted or retained intracellularly depending on the sequence

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and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides which encode TRICH may be designed to contain signal sequences which direct secretion of TRICH through a prokaryotic or eukaryotic cell membrane.

In addition, a host cell strain may be chosen for its ability to modulate expression of the inserted polynucleotides or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" or "pro" form of the protein may also be used to specify protein targeting, folding, and/or activity. Different host cells which have specific cellular machinery and characteristic mechanisms for post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and WI38) are available from the American Type Culture Collection (ATCC, Manassas VA) and may be chosen to ensure the correct modification and processing of the foreign protein.

In another embodiment of the invention, natural, modified, or recombinant polynucleotides encoding TRICH may be ligated to a heterologous sequence resulting in translation of a fusion protein in any of the aforementioned host systems. For example, a chimeric TRICH protein containing a heterologous moiety that can be recognized by a commercially available antibody may facilitate the screening of peptide libraries for inhibitors of TRICH activity. Heterologous protein and peptide moieties may also facilitate purification of fusion proteins using commercially available affinity matrices. Such moieties include, but are not limited to, glutathione S-transferase (GST), maltose binding protein (MBP), thioredoxin (Trx), calmodulin binding peptide (CBP), 6-His, FLAG, c-myc, and hemagglutinin (HA). GST, MBP, Trx, CBP, and 6-His enable purification of their cognate fusion proteins on immobilized glutathione, maltose, phenylarsine oxide, calmodulin, and metal-chelate resins, respectively. FLAG, c-myc, and hemagglutinin (HA) enable immunoaffinity purification of fusion proteins using commercially available monoclonal and polyclonal antibodies that specifically recognize these epitope tags. A fusion protein may also be engineered to contain a proteolytic cleavage site located between the TRICH encoding sequence and the heterologous protein sequence, so that TRICH may be cleaved away from the heterologous moiety following purification. Methods for fusion protein expression and purification are discussed in Ausubel et al. (supra, ch. 10 and 16). A variety of commercially available kits may also be used to facilitate expression and purification of fusion proteins.

In another embodiment, synthesis of radiolabeled TRICH may be achieved *in vitro* using the TNT rabbit reticulocyte lysate or wheat germ extract system (Promega). These systems couple

transcription and translation of protein-coding sequences operably associated with the T7, T3, or SP6 promoters. Translation takes place in the presence of a radiolabeled amino acid precursor, for example, ³⁵S-methionine.

TRICH, fragments of TRICH, or variants of TRICH may be used to screen for compounds that specifically bind to TRICH. One or more test compounds may be screened for specific binding to TRICH. In various embodiments, 1, 2, 3, 4, 5, 10, 20, 50, 100, or 200 test compounds can be screened for specific binding to TRICH. Examples of test compounds can include antibodies, anticalins, oligonucleotides, proteins (e.g., ligands or receptors), or small molecules.

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In related embodiments, variants of TRICH can be used to screen for binding of test compounds, such as antibodies, to TRICH, a variant of TRICH, or a combination of TRICH and/or one or more variants TRICH. In an embodiment, a variant of TRICH can be used to screen for compounds that bind to a variant of TRICH, but not to TRICH having the exact sequence of a sequence of SEQ ID NO:1-26. TRICH variants used to perform such screening can have a range of about 50% to about 99% sequence identity to TRICH, with various embodiments having 60%, 70%, 75%, 80%, 85%, 90%, and 95% sequence identity.

In an embodiment, a compound identified in a screen for specific binding to TRICH can be closely related to the natural ligand of TRICH, e.g., a ligand or fragment thereof, a natural substrate, a structural or functional mimetic, or a natural binding partner (Coligan, J.E. et al. (1991) <u>Current Protocols in Immunology</u> 1(2):Chapter 5). In another embodiment, the compound thus identified can be a natural ligand of a receptor TRICH (Howard, A.D. et al. (2001) Trends Pharmacol. Sci.22:132-140; Wise, A. et al. (2002) Drug Discovery Today 7:235-246).

In other embodiments, a compound identified in a screen for specific binding to TRICH can be closely related to the natural receptor to which TRICH binds, at least a fragment of the receptor, or a fragment of the receptor including all or a portion of the ligand binding site or binding pocket. For example, the compound may be a receptor for TRICH which is capable of propagating a signal, or a decoy receptor for TRICH which is not capable of propagating a signal (Ashkenazi, A. and V.M. Divit (1999) Curr. Opin. Cell Biol. 11:255-260; Mantovani, A. et al. (2001) Trends Immunol. 22:328-336). The compound can be rationally designed using known techniques. Examples of such techniques include those used to construct the compound etanercept (ENBREL; Amgen Inc., Thousand Oaks CA), which is efficacious for treating rheumatoid arthritis in humans. Etanercept is an engineered p75 tumor necrosis factor (TNF) receptor dimer linked to the Fc portion of human IgG₁ (Taylor, P.C. et al. (2001) Curr. Opin. Immunol. 13:611-616).

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In one embodiment, two or more antibodies having similar or, alternatively, different specificities can be screened for specific binding to TRICH, fragments of TRICH, or variants of TRICH. The binding specificity of the antibodies thus screened can thereby be selected to identify particular fragments or variants of TRICH. In one embodiment, an antibody can be selected such that its binding specificity allows for preferential identification of specific fragments or variants of TRICH. In another embodiment, an antibody can be selected such that its binding specificity allows for preferential diagnosis of a specific disease or condition having increased, decreased, or otherwise abnormal production of TRICH.

In an embodiment, anticalins can be screened for specific binding to TRICH, fragments of TRICH, or variants of TRICH. Anticalins are ligand-binding proteins that have been constructed based on a lipocalin scaffold (Weiss, G.A. and H.B. Lowman (2000) Chem. Biol. 7:R177-R184; Skerra, A. (2001) J. Biotechnol. 74:257-275). The protein architecture of lipocalins can include a beta-barrel having eight antiparallel beta-strands, which supports four loops at its open end. These loops form the natural ligand-binding site of the lipocalins, a site which can be re-engineered *in vitro* by amino acid substitutions to impart novel binding specificities. The amino acid substitutions can be made using methods known in the art or described herein, and can include conservative substitutions (e.g., substitutions that do not alter binding specificity) or substitutions that modestly, moderately, or significantly alter binding specificity.

In one embodiment, screening for compounds which specifically bind to, stimulate, or inhibit TRICH involves producing appropriate cells which express TRICH, either as a secreted protein or on the cell membrane. Preferred cells include cells from mammals, yeast, *Drosophila*, or *E. coli*. Cells expressing TRICH or cell membrane fractions which contain TRICH are then contacted with a test compound and binding, stimulation, or inhibition of activity of either TRICH or the compound is analyzed.

An assay may simply test binding of a test compound to the polypeptide, wherein binding is detected by a fluorophore, radioisotope, enzyme conjugate, or other detectable label. For example, the assay may comprise the steps of combining at least one test compound with TRICH, either in solution or affixed to a solid support, and detecting the binding of TRICH to the compound. Alternatively, the assay may detect or measure binding of a test compound in the presence of a labeled competitor. Additionally, the assay may be carried out using cell-free preparations, chemical libraries, or natural product mixtures, and the test compound(s) may be free in solution or affixed to a solid support.

An assay can be used to assess the ability of a compound to bind to its natural ligand and/or to

inhibit the binding of its natural ligand to its natural receptors. Examples of such assays include radio-labeling assays such as those described in U.S. Patent No. 5,914,236 and U.S. Patent No. 6,372,724. In a related embodiment, one or more amino acid substitutions can be introduced into a polypeptide compound (such as a receptor) to improve or alter its ability to bind to its natural ligands (Matthews, D.J. and J.A. Wells. (1994) Chem. Biol. 1:25-30). In another related embodiment, one or more amino acid substitutions can be introduced into a polypeptide compound (such as a ligand) to improve or alter its ability to bind to its natural receptors (Cunningham, B.C. and J.A. Wells (1991) Proc. Natl. Acad. Sci. USA 88:3407-3411; Lowman, H.B. et al. (1991) J. Biol. Chem. 266:10982-10988).

TRICH, fragments of TRICH, or variants of TRICH may be used to screen for compounds that modulate the activity of TRICH. Such compounds may include agonists, antagonists, or partial or inverse agonists. In one embodiment, an assay is performed under conditions permissive for TRICH activity, wherein TRICH is combined with at least one test compound, and the activity of TRICH in the presence of a test compound is compared with the activity of TRICH in the absence of the test compound. A change in the activity of TRICH in the presence of the test compound is indicative of a compound that modulates the activity of TRICH. Alternatively, a test compound is combined with an *in vitro* or cell-free system comprising TRICH under conditions suitable for TRICH activity, and the assay is performed. In either of these assays, a test compound which modulates the activity of TRICH may do so indirectly and need not come in direct contact with the test compound. At least one and up to a plurality of test compounds may be screened.

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In another embodiment, polynucleotides encoding TRICH or their mammalian homologs may be "knocked out" in an animal model system using homologous recombination in embryonic stem (ES) cells. Such techniques are well known in the art and are useful for the generation of animal models of human disease (see, e.g., U.S. Patent No. 5,175,383 and U.S. Patent No. 5,767,337). For example, mouse ES cells, such as the mouse 129/SvJ cell line, are derived from the early mouse embryo and grown in culture. The ES cells are transformed with a vector containing the gene of interest disrupted by a marker gene, e.g., the neomycin phosphotransferase gene (*neo*; Capecchi, M.R. (1989) Science 244:1288-1292). The vector integrates into the corresponding region of the host genome by homologous recombination. Alternatively, homologous recombination takes place using the Cre-loxP system to knockout a gene of interest in a tissue- or developmental stage-specific manner (Marth, J.D. (1996) Clin. Invest. 97:1999-2002; Wagner, K.U. et al. (1997) Nucleic Acids Res. 25:4323-4330). Transformed ES cells are identified and microinjected into mouse cell blastocysts such as those from the C57BL/6 mouse strain. The blastocysts are surgically transferred to pseudopregnant dams, and

the resulting chimeric progeny are genotyped and bred to produce heterozygous or homozygous strains. Transgenic animals thus generated may be tested with potential therapeutic or toxic agents.

Polynucleotides encoding TRICH may also be manipulated *in vitro* in ES cells derived from human blastocysts. Human ES cells have the potential to differentiate into at least eight separate cell lineages including endoderm, mesoderm, and ectodermal cell types. These cell lineages differentiate into, for example, neural cells, hematopoietic lineages, and cardiomyocytes (Thomson, J.A. et al. (1998) Science 282:1145-1147).

Polynucleotides encoding TRICH can also be used to create "knockin" humanized animals (pigs) or transgenic animals (mice or rats) to model human disease. With knockin technology, a region of a polynucleotide encoding TRICH is injected into animal ES cells, and the injected sequence integrates into the animal cell genome. Transformed cells are injected into blastulae, and the blastulae are implanted as described above. Transgenic progeny or inbred lines are studied and treated with potential pharmaceutical agents to obtain information on treatment of a human disease. Alternatively, a mammal inbred to overexpress TRICH, e.g., by secreting TRICH in its milk, may also serve as a convenient source of that protein (Janne, J. et al. (1998) Biotechnol. Annu. Rev. 4:55-74).

THERAPEUTICS

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Chemical and structural similarity, e.g., in the context of sequences and motifs, exists between regions of TRICH and transporters and ion channels. The expression of TRICH is closely associated with normal heart tissue, liver tumor tissue and diseased corpus callosum tissue. In addition, examples of tissues expressing TRICH can be found in Table 6 and can also be found in Example XI.

Therefore, TRICH appears to play a role in transport, neurological, muscle, immunological and cell proliferative disorders. In the treatment of disorders associated with increased TRICH expression or activity, it is desirable to decrease the expression or activity of TRICH. In the treatment of disorders associated with decreased TRICH expression or activity of TRICH.

Therefore, in one embodiment, TRICH or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH. Examples of such disorders include, but are not limited to, a transport disorder such as akinesia, amyotrophic lateral sclerosis, ataxia telangiectasia, cystic fibrosis, Becker's muscular dystrophy, Bell's palsy, Charcot-Marie Tooth disease, diabetes mellitus, diabetes insipidus, diabetic neuropathy, Duchenne muscular dystrophy, hyperkalemic periodic paralysis, normokalemic periodic paralysis, Parkinson's disease, malignant hyperthermia, multidrug resistance, myasthenia gravis,

myotonic dystrophy, catatonia, tardive dyskinesia, dystonias, peripheral neuropathy, cerebral neoplasms, prostate cancer, cardiac disorders associated with transport, e.g., angina, bradyarrythmia, tachyarrythmia, hypertension, Long QT syndrome, myocarditis, cardiomyopathy, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, thyrotoxic myopathy, ethanol myopathy, dermatomyositis, inclusion body myositis, infectious myositis, polymyositis, neurological disorders associated with transport, e.g., Alzheimer's disease, amnesia, bipolar disorder, dementia, depression, epilepsy, Tourette's disorder, paranoid psychoses, and schizophrenia, and other disorders associated with transport, e.g., neurofibromatosis, postherpetic neuralgia, trigeminal neuropathy, sarcoidosis, sickle cell anemia, Wilson's disease, cataracts, infertility, pulmonary artery stenosis, sensorineural autosomal deafness, hyperglycemia, hypoglycemia, Grave's disease, goiter, Cushing's disease, Addison's disease, glucose-galactose malabsorption syndrome, glycogen storage disease, hypercholesterolemia, adrenoleukodystrophy, Zellweger syndrome, Menkes disease, occipital horn syndrome, von Gierke disease, pseudohypoaldosteronism type 1, Liddle's syndrome, cystinuria, iminoglycinuria, Hartup disease, Fanconi disease, and Bartter syndrome; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, hemiplegic migraine, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, Tourette's disorder, progressive supranuclear palsy, corticobasal degeneration, and familial frontotemporal dementia; a muscle disorder such as cardiomyopathy, myocarditis, Duchenne's muscular dystrophy, Becker's

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muscular dystrophy, myotonic dystrophy, central core disease, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, infectious myositis, polymyositis, dermatomyositis, inclusion body myositis, thyrotoxic myopathy, ethanol myopathy, angina, anaphylactic shock, arrhythmias, asthma, cardiovascular shock, Cushing's syndrome, hypertension, hypoglycemia, myocardial infarction, migraine, pheochromocytoma, and myopathies including encephalopathy, epilepsy, Kearns-Sayre syndrome, lactic acidosis, myoclonic disorder, ophthalmoplegia, acid maltase deficiency (AMD, also known as Pompe's disease), generalized myotonia, and myotonia congenita; an immunological disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; and a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus.

In another embodiment, a vector capable of expressing TRICH or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH including, but not limited to, those described above.

In a further embodiment, a composition comprising a substantially purified TRICH in conjunction with a suitable pharmaceutical carrier may be administered to a subject to treat or prevent

a disorder associated with decreased expression or activity of TRICH including, but not limited to, those provided above.

In still another embodiment, an agonist which modulates the activity of TRICH may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH including, but not limited to, those listed above.

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In a further embodiment, an antagonist of TRICH may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of TRICH. Examples of such disorders include, but are not limited to, those transport, neurological, muscle, immunological and cell proliferative disorders described above. In one aspect, an antibody which specifically binds TRICH may be used directly as an antagonist or indirectly as a targeting or delivery mechanism for bringing a pharmaceutical agent to cells or tissues which express TRICH.

In an additional embodiment, a vector expressing the complement of the polynucleotide encoding TRICH may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of TRICH including, but not limited to, those described above.

In other embodiments, any protein, agonist, antagonist, antibody, complementary sequence, or vector embodiments may be administered in combination with other appropriate therapeutic agents. Selection of the appropriate agents for use in combination therapy may be made by one of ordinary skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic agents may act synergistically to effect the treatment or prevention of the various disorders described above. Using this approach, one may be able to achieve therapeutic efficacy with lower dosages of each agent, thus reducing the potential for adverse side effects.

An antagonist of TRICH may be produced using methods which are generally known in the art. In particular, purified TRICH may be used to produce antibodies or to screen libraries of pharmaceutical agents to identify those which specifically bind TRICH. Antibodies to TRICH may also be generated using methods that are well known in the art. Such antibodies may include, but are not limited to, polyclonal, monoclonal, chimeric, and single chain antibodies, Fab fragments, and fragments produced by a Fab expression library. Neutralizing antibodies (i.e., those which inhibit dimer formation) are generally preferred for therapeutic use. Single chain antibodies (e.g., from camels or llamas) may be potent enzyme inhibitors and may have advantages in the design of peptide mimetics, and in the development of immuno-adsorbents and biosensors (Muyldermans, S. (2001) J. Biotechnol. 74:277-302).

For the production of antibodies, various hosts including goats, rabbits, rats, mice, camels,

dromedaries, llamas, humans, and others may be immunized by injection with TRICH or with any fragment or oligopeptide thereof which has immunogenic properties. Depending on the host species, various adjuvants may be used to increase immunological response. Such adjuvants include, but are not limited to, Freund's, mineral gels such as aluminum hydroxide, and surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, KLH, and dinitrophenol. Among adjuvants used in humans, BCG (bacilli Calmette-Guerin) and *Corynebacterium parvum* are especially preferable.

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It is preferred that the oligopeptides, peptides, or fragments used to induce antibodies to TRICH have an amino acid sequence consisting of at least about 5 amino acids, and generally will consist of at least about 10 amino acids. It is also preferable that these oligopeptides, peptides, or fragments are identical to a portion of the amino acid sequence of the natural protein. Short stretches of TRICH amino acids may be fused with those of another protein, such as KLH, and antibodies to the chimeric molecule may be produced.

Monoclonal antibodies to TRICH may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-hybridoma technique (Kohler, G. et al. (1975) Nature 256:495-497; Kozbor, D. et al. (1985) J. Immunol. Methods 81:31-42; Cote, R.J. et al. (1983) Proc. Natl. Acad. Sci. USA 80:2026-2030; Cole, S.P. et al. (1984) Mol. Cell Biol. 62:109-120).

In addition, techniques developed for the production of "chimeric antibodies," such as the splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used (Morrison, S.L. et al. (1984) Proc. Natl. Acad. Sci. USA 81:6851-6855; Neuberger, M.S. et al. (1984) Nature 312:604-608; Takeda, S. et al. (1985) Nature 314:452-454). Alternatively, techniques described for the production of single chain antibodies may be adapted, using methods known in the art, to produce TRICH-specific single chain antibodies. Antibodies with related specificity, but of distinct idiotypic composition, may be generated by chain shuffling from random combinatorial immunoglobulin libraries (Burton, D.R. (1991) Proc. Natl. Acad. Sci. USA 88:10134-10137).

Antibodies may also be produced by inducing *in vivo* production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents as disclosed in the literature (Orlandi, R. et al. (1989) Proc. Natl. Acad. Sci. USA 86:3833-3837; Winter, G. et al. (1991) Nature 349:293-299).

Antibody fragments which contain specific binding sites for TRICH may also be generated. For example, such fragments include, but are not limited to, $F(ab^2)_2$ fragments produced by pepsin digestion of the antibody molecule and Fab fragments generated by reducing the disulfide bridges of the $F(ab^2)_2$ fragments. Alternatively, Fab expression libraries may be constructed to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity (Huse, W.D. et al. (1989) Science 246:1275-1281).

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Various immunoassays may be used for screening to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric assays using either polyclonal or monoclonal antibodies with established specificities are well known in the art. Such immunoassays typically involve the measurement of complex formation between TRICH and its specific antibody. A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering TRICH epitopes is generally used, but a competitive binding assay may also be employed (Pound, *supra*).

Various methods such as Scatchard analysis in conjunction with radioimmunoassay techniques may be used to assess the affinity of antibodies for TRICH. Affinity is expressed as an association constant, K_a , which is defined as the molar concentration of TRICH-antibody complex divided by the molar concentrations of free antigen and free antibody under equilibrium conditions. The K_a determined for a preparation of polyclonal antibodies, which are heterogeneous in their affinities for multiple TRICH epitopes, represents the average affinity, or avidity, of the antibodies for TRICH. The K_a determined for a preparation of monoclonal antibodies, which are monospecific for a particular TRICH epitope, represents a true measure of affinity. High-affinity antibody preparations with K_a ranging from about 10^9 to 10^{12} L/mole are preferred for use in immunoassays in which the TRICH-antibody complex must withstand rigorous manipulations. Low-affinity antibody preparations with K_a ranging from about 10^6 to 10^7 L/mole are preferred for use in immunopurification and similar procedures which ultimately require dissociation of TRICH, preferably in active form, from the antibody (Catty, D. (1988) Antibodies, Volume I: A Practical Approach, IRL Press, Washington DC; Liddell, J.E. and A. Cryer (1991) A Practical Guide to Monoclonal Antibodies, John Wiley & Sons, New York NY).

The titer and avidity of polyclonal antibody preparations may be further evaluated to determine the quality and suitability of such preparations for certain downstream applications. For example, a polyclonal antibody preparation containing at least 1-2 mg specific antibody/ml, preferably 5-10 mg specific antibody/ml, is generally employed in procedures requiring precipitation of TRICH-antibody

complexes. Procedures for evaluating antibody specificity, titer, and avidity, and guidelines for antibody quality and usage in various applications, are generally available (Catty, *supra*; Coligan et al., *supra*).

In another embodiment of the invention, polynucleotides encoding TRICH, or any fragment or complement thereof, may be used for therapeutic purposes. In one aspect, modifications of gene expression can be achieved by designing complementary sequences or antisense molecules (DNA, RNA, PNA, or modified oligonucleotides) to the coding or regulatory regions of the gene encoding TRICH. Such technology is well known in the art, and antisense oligonucleotides or larger fragments can be designed from various locations along the coding or control regions of sequences encoding TRICH (Agrawal, S., ed. (1996) Antisense Therapeutics, Humana Press, Totawa NJ).

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In therapeutic use, any gene delivery system suitable for introduction of the antisense sequences into appropriate target cells can be used. Antisense sequences can be delivered intracellularly in the form of an expression plasmid which, upon transcription, produces a sequence complementary to at least a portion of the cellular sequence encoding the target protein (Slater, J.E. et al. (1998) J. Allergy Clin. Immunol. 102:469-475; Scanlon, K.J. et al. (1995) 9:1288-1296). Antisense sequences can also be introduced intracellularly through the use of viral vectors, such as retrovirus and adeno-associated virus vectors (Miller, A.D. (1990) Blood 76:271; Ausubel et al., *supra*; Uckert, W. and W. Walther (1994) Pharmacol. Ther. 63:323-347). Other gene delivery mechanisms include liposome-derived systems, artificial viral envelopes, and other systems known in the art (Rossi, J.J. (1995) Br. Med. Bull. 51:217-225; Boado, R.J. et al. (1998) J. Pharm. Sci. 87:1308-1315; Morris, M.C. et al. (1997) Nucleic Acids Res. 25:2730-2736).

In another embodiment of the invention, polynucleotides encoding TRICH may be used for somatic or germline gene therapy. Gene therapy may be performed to (i) correct a genetic deficiency (e.g., in the cases of severe combined immunodeficiency (SCID)-X1 disease characterized by X-linked inheritance (Cavazzana-Calvo, M. et al. (2000) Science 288:669-672), severe combined immunodeficiency syndrome associated with an inherited adenosine deaminase (ADA) deficiency (Blaese, R.M. et al. (1995) Science 270:475-480; Bordignon, C. et al. (1995) Science 270:470-475), cystic fibrosis (Zabner, J. et al. (1993) Cell 75:207-216; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:643-666; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:667-703), thalassamias, familial hypercholesterolemia, and hemophilia resulting from Factor VIII or Factor IX deficiencies (Crystal, R.G. (1995) Science 270:404-410; Verma, I.M. and N. Somia (1997) Nature 389:239-242)), (ii) express a conditionally lethal gene product (e.g., in the case of cancers which result from unregulated

cell proliferation), or (iii) express a protein which affords protection against intracellular parasites (e.g., against human retroviruses, such as human immunodeficiency virus (HIV) (Baltimore, D. (1988) Nature 335:395-396; Poeschla, E. et al. (1996) Proc. Natl. Acad. Sci. USA 93:11395-11399), hepatitis B or C virus (HBV, HCV); fungal parasites, such as Candida albicans and Paracoccidioides brasiliensis; and protozoan parasites such as Plasmodium falciparum and Trypanosoma cruzi). In the case where a genetic deficiency in TRICH expression or regulation causes disease, the expression of TRICH from an appropriate population of transduced cells may alleviate the clinical manifestations caused by the genetic deficiency.

In a further embodiment of the invention, diseases or disorders caused by deficiencies in TRICH are treated by constructing mammalian expression vectors encoding TRICH and introducing these vectors by mechanical means into TRICH-deficient cells. Mechanical transfer technologies for use with cells *in vivo* or *ex vitro* include (i) direct DNA microinjection into individual cells, (ii) ballistic gold particle delivery, (iii) liposome-mediated transfection, (iv) receptor-mediated gene transfer, and (v) the use of DNA transposons (Morgan, R.A. and W.F. Anderson (1993) Annu. Rev. Biochem. 62:191-217; Ivics, Z. (1997) Cell 91:501-510; Boulay, J.-L. and H. Récipon (1998) Curr. Opin. Biotechnol. 9:445-450).

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Expression vectors that may be effective for the expression of TRICH include, but are not limited to, the PCDNA 3.1, EPITAG, PRCCMV2, PREP, PVAX, PCR2-TOPOTA vectors (Invitrogen, Carlsbad CA), PCMV-SCRIPT, PCMV-TAG, PEGSH/PERV (Stratagene, La Jolla CA), and PTET-OFF, PTET-ON, PTRE2, PTRE2-LUC, PTK-HYG (Clontech, Palo Alto CA). TRICH may be expressed using (i) a constitutively active promoter, (e.g., from cytomegalovirus (CMV), Rous sarcoma virus (RSV), SV40 virus, thymidine kinase (TK), or β-actin genes), (ii) an inducible promoter (e.g., the tetracycline-regulated promoter (Gossen, M. and H. Bujard (1992) Proc. Natl. Acad. Sci. USA 89:5547-5551; Gossen, M. et al. (1995) Science 268:1766-1769; Rossi, F.M.V. and H.M. Blau (1998) Curr. Opin. Biotechnol. 9:451-456), commercially available in the T-REX plasmid (Invitrogen)); the ecdysone-inducible promoter (available in the plasmids PVGRXR and PIND; Invitrogen); the FK506/rapamycin inducible promoter; or the RU486/mifepristone inducible promoter (Rossi, F.M.V. and H.M. Blau, supra)), or (iii) a tissue-specific promoter or the native promoter of the endogenous gene encoding TRICH from a normal individual.

Commercially available liposome transformation kits (e.g., the PERFECT LIPID TRANSFECTION KIT, available from Invitrogen) allow one with ordinary skill in the art to deliver polynucleotides to target cells in culture and require minimal effort to optimize experimental

parameters. In the alternative, transformation is performed using the calcium phosphate method (Graham, F.L. and A.J. Eb (1973) Virology 52:456-467), or by electroporation (Neumann, E. et al. (1982) EMBO J. 1:841-845). The introduction of DNA to primary cells requires modification of these standardized mammalian transfection protocols.

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In another embodiment of the invention, diseases or disorders caused by genetic defects with respect to TRICH expression are treated by constructing a retrovirus vector consisting of (i) the polynucleotide encoding TRICH under the control of an independent promoter or the retrovirus long terminal repeat (LTR) promoter, (ii) appropriate RNA packaging signals, and (iii) a Rev-responsive element (RRE) along with additional retrovirus cis-acting RNA sequences and coding sequences required for efficient vector propagation. Retrovirus vectors (e.g., PFB and PFBNEO) are commercially available (Stratagene) and are based on published data (Riviere, I. et al. (1995) Proc. Natl. Acad. Sci. USA 92:6733-6737), incorporated by reference herein. The vector is propagated in an appropriate vector producing cell line (VPCL) that expresses an envelope gene with a tropism for receptors on the target cells or a promiscuous envelope protein such as VSVg (Armentano, D. et al. (1987) J. Virol. 61:1647-1650; Bender, M.A. et al. (1987) J. Virol. 61:1639-1646; Adam, M.A. and A.D. Miller (1988) J. Virol. 62:3802-3806; Dull, T. et al. (1998) J. Virol. 72:8463-8471; Zufferey, R. et al. (1998) J. Virol. 72:9873-9880). U.S. Patent No. 5,910,434 to Rigg ("Method for obtaining retrovirus packaging cell lines producing high transducing efficiency retroviral supernatant") discloses a method for obtaining retrovirus packaging cell lines and is hereby incorporated by reference. Propagation of retrovirus vectors, transduction of a population of cells (e.g., CD4+ T-cells), and the return of transduced cells to a patient are procedures well known to persons skilled in the art of gene therapy and have been well documented (Ranga, U. et al. (1997) J. Virol. 71:7020-7029; Bauer, G. et al. (1997) Blood 89:2259-2267; Bonyhadi, M.L. (1997) J. Virol. 71:4707-4716; Ranga, U. et al. (1998) Proc. Natl. Acad. Sci. USA 95:1201-1206; Su, L. (1997) Blood 89:2283-2290).

In an embodiment, an adenovirus-based gene therapy delivery system is used to deliver polynucleotides encoding TRICH to cells which have one or more genetic abnormalities with respect to the expression of TRICH. The construction and packaging of adenovirus-based vectors are well known to those with ordinary skill in the art. Replication defective adenovirus vectors have proven to be versatile for importing genes encoding immunoregulatory proteins into intact islets in the pancreas (Csete, M.E. et al. (1995) Transplantation 27:263-268). Potentially useful adenoviral vectors are described in U.S. Patent No. 5,707,618 to Armentano ("Adenovirus vectors for gene therapy"), hereby incorporated by reference. For adenoviral vectors, see also Antinozzi, P.A. et al. (1999; Annu.

Rev. Nutr. 19:511-544) and Verma, I.M. and N. Somia (1997; Nature 18:389:239-242).

In another embodiment, a herpes-based, gene therapy delivery system is used to deliver polynucleotides encoding TRICH to target cells which have one or more genetic abnormalities with respect to the expression of TRICH. The use of herpes simplex virus (HSV)-based vectors may be especially valuable for introducing TRICH to cells of the central nervous system, for which HSV has a tropism. The construction and packaging of herpes-based vectors are well known to those with ordinary skill in the art. A replication-competent herpes simplex virus (HSV) type 1-based vector has been used to deliver a reporter gene to the eyes of primates (Liu, X. et al. (1999) Exp. Eye Res. 169:385-395). The construction of a HSV-1 virus vector has also been disclosed in detail in U.S. Patent No. 5,804,413 to DeLuca ("Herpes simplex virus strains for gene transfer"), which is hereby incorporated by reference. U.S. Patent No. 5,804,413 teaches the use of recombinant HSV d92 which consists of a genome containing at least one exogenous gene to be transferred to a cell under the control of the appropriate promoter for purposes including human gene therapy. Also taught by this patent are the construction and use of recombinant HSV strains deleted for ICP4, ICP27 and ICP22. For HSV vectors, see also Goins, W.F. et al. (1999; J. Virol. 73:519-532) and Xu, H. et al. (1994; Dev. Biol. 163:152-161). The manipulation of cloned herpesvirus sequences, the generation of recombinant virus following the transfection of multiple plasmids containing different segments of the large herpesvirus genomes, the growth and propagation of herpesvirus, and the infection of cells with herpesvirus are techniques well known to those of ordinary skill in the art.

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In another embodiment, an alphavirus (positive, single-stranded RNA virus) vector is used to deliver polynucleotides encoding TRICH to target cells. The biology of the prototypic alphavirus, Semliki Forest Virus (SFV), has been studied extensively and gene transfer vectors have been based on the SFV genome (Garoff, H. and K.-J. Li (1998) Curr. Opin. Biotechnol. 9:464-469). During alphavirus RNA replication, a subgenomic RNA is generated that normally encodes the viral capsid proteins. This subgenomic RNA replicates to higher levels than the full length genomic RNA, resulting in the overproduction of capsid proteins relative to the viral proteins with enzymatic activity (e.g., protease and polymerase). Similarly, inserting the coding sequence for TRICH into the alphavirus genome in place of the capsid-coding region results in the production of a large number of TRICH-coding RNAs and the synthesis of high levels of TRICH in vector transduced cells. While alphavirus infection is typically associated with cell lysis within a few days, the ability to establish a persistent infection in hamster normal kidney cells (BHK-21) with a variant of Sindbis virus (SIN) indicates that the lytic replication of alphaviruses can be altered to suit the needs of the gene therapy

application (Dryga, S.A. et al. (1997) Virology 228:74-83). The wide host range of alphaviruses will allow the introduction of TRICH into a variety of cell types. The specific transduction of a subset of cells in a population may require the sorting of cells prior to transduction. The methods of manipulating infectious cDNA clones of alphaviruses, performing alphavirus cDNA and RNA transfections, and performing alphavirus infections, are well known to those with ordinary skill in the art.

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Oligonucleotides derived from the transcription initiation site, e.g., between about positions -10 and +10 from the start site, may also be employed to inhibit gene expression. Similarly, inhibition can be achieved using triple helix base-pairing methodology. Triple helix pairing is useful because it causes inhibition of the ability of the double helix to open sufficiently for the binding of polymerases, transcription factors, or regulatory molecules. Recent therapeutic advances using triplex DNA have been described in the literature (Gee, J.E. et al. (1994) in Huber, B.E. and B.I. Carr, Molecular and Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp. 163-177). A complementary sequence or antisense molecule may also be designed to block translation of mRNA by preventing the transcript from binding to ribosomes.

Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage. For example, engineered hammerhead motif ribozyme molecules may specifically and efficiently catalyze endonucleolytic cleavage of RNA molecules encoding TRICH.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the target molecule for ribozyme cleavage sites, including the following sequences: GUA, GUU, and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides, corresponding to the region of the target gene containing the cleavage site, may be evaluated for secondary structural features which may render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing accessibility to hybridization with complementary oligonucleotides using ribonuclease protection assays.

Complementary ribonucleic acid molecules and ribozymes may be prepared by any method known in the art for the synthesis of nucleic acid molecules. These include techniques for chemically synthesizing oligonucleotides such as solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by *in vitro* and *in vivo* transcription of DNA molecules encoding TRICH. Such DNA sequences may be incorporated into a wide variety of vectors with suitable RNA

polymerase promoters such as T7 or SP6. Alternatively, these cDNA constructs that synthesize complementary RNA, constitutively or inducibly, can be introduced into cell lines, cells, or tissues.

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RNA molecules may be modified to increase intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends of the molecule, or the use of phosphorothioate or 2'O-methyl rather than phosphodiesterase linkages within the backbone of the molecule. This concept is inherent in the production of PNAs and can be extended in all of these molecules by the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, as well as acetyl-, methyl-, thio-, and similarly modified forms of adenine, cytidine, guanine, thymine, and uridine which are not as easily recognized by endogenous endonucleases.

An additional embodiment of the invention encompasses a method for screening for a compound which is effective in altering expression of a polynucleotide encoding TRICH. Compounds which may be effective in altering expression of a specific polynucleotide may include, but are not limited to, oligonucleotides, antisense oligonucleotides, triple helix-forming oligonucleotides, transcription factors and other polypeptide transcriptional regulators, and non-macromolecular chemical entities which are capable of interacting with specific polynucleotide sequences. Effective compounds may alter polynucleotide expression by acting as either inhibitors or promoters of polynucleotide expression. Thus, in the treatment of disorders associated with increased TRICH expression or activity, a compound which specifically inhibits expression of the polynucleotide encoding TRICH may be therapeutically useful, and in the treatment of disorders associated with decreased TRICH expression or activity, a compound which specifically promotes expression of the polynucleotide encoding TRICH may be therapeutically useful.

At least one, and up to a plurality, of test compounds may be screened for effectiveness in altering expression of a specific polynucleotide. A test compound may be obtained by any method commonly known in the art, including chemical modification of a compound known to be effective in altering polynucleotide expression; selection from an existing, commercially-available or proprietary library of naturally-occurring or non-natural chemical compounds; rational design of a compound based on chemical and/or structural properties of the target polynucleotide; and selection from a library of chemical compounds created combinatorially or randomly. A sample comprising a polynucleotide encoding TRICH is exposed to at least one test compound thus obtained. The sample may comprise, for example, an intact or permeabilized cell, or an *in vitro* cell-free or reconstituted biochemical system. Alterations in the expression of a polynucleotide encoding TRICH are assayed by any method commonly known in the art. Typically, the expression of a specific nucleotide is

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detected by hybridization with a probe having a nucleotide sequence complementary to the sequence of the polynucleotide encoding TRICH. The amount of hybridization may be quantified, thus forming the basis for a comparison of the expression of the polynucleotide both with and without exposure to one or more test compounds. Detection of a change in the expression of a polynucleotide exposed to a test compound indicates that the test compound is effective in altering the expression of the polynucleotide. A screen for a compound effective in altering expression of a specific polynucleotide can be carried out, for example, using a *Schizosaccharomyces pombe* gene expression system (Atkins, D. et al. (1999) U.S. Patent No. 5,932,435; Arndt, G.M. et al. (2000) Nucleic Acids Res. 28:E15) or a human cell line such as HeLa cell (Clarke, M.L. et al. (2000) Biochem. Biophys. Res. Commun. 268:8-13). A particular embodiment of the present invention involves screening a combinatorial library of oligonucleotides (such as deoxyribonucleotides, ribonucleotides, peptide nucleic acids, and modified oligonucleotides) for antisense activity against a specific polynucleotide sequence (Bruice, T.W. et al. (1997) U.S. Patent No. 5,686,242; Bruice, T.W. et al. (2000) U.S. Patent No. 6,022,691).

Many methods for introducing vectors into cells or tissues are available and equally suitable for use *in vivo*, *in vitro*, and *ex vivo*. For *ex vivo* therapy, vectors may be introduced into stem cells taken from the patient and clonally propagated for autologous transplant back into that same patient. Delivery by transfection, by liposome injections, or by polycationic amino polymers may be achieved using methods which are well known in the art (Goldman, C.K. et al. (1997) Nat. Biotechnol. 15:462-466).

Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as humans, dogs, cats, cows, horses, rabbits, and monkeys.

An additional embodiment of the invention relates to the administration of a composition which generally comprises an active ingredient formulated with a pharmaceutically acceptable excipient. Excipients may include, for example, sugars, starches, celluloses, gums, and proteins. Various formulations are commonly known and are thoroughly discussed in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing, Easton PA). Such compositions may consist of TRICH, antibodies to TRICH, and mimetics, agonists, antagonists, or inhibitors of TRICH.

The compositions utilized in this invention may be administered by any number of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, pulmonary, transdermal, subcutaneous, intraperitoneal, intranasal, enteral, topical,

sublingual, or rectal means.

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Compositions for pulmonary administration may be prepared in liquid or dry powder form. These compositions are generally aerosolized immediately prior to inhalation by the patient. In the case of small molecules (e.g. traditional low molecular weight organic drugs), aerosol delivery of fast-acting formulations is well-known in the art. In the case of macromolecules (e.g. larger peptides and proteins), recent developments in the field of pulmonary delivery via the alveolar region of the lung have enabled the practical delivery of drugs such as insulin to blood circulation (see, e.g., Patton, J.S. et al., U.S. Patent No. 5,997,848). Pulmonary delivery has the advantage of administration without needle injection, and obviates the need for potentially toxic penetration enhancers.

Compositions suitable for use in the invention include compositions wherein the active ingredients are contained in an effective amount to achieve the intended purpose. The determination of an effective dose is well within the capability of those skilled in the art.

Specialized forms of compositions may be prepared for direct intracellular delivery of macromolecules comprising TRICH or fragments thereof. For example, liposome preparations containing a cell-impermeable macromolecule may promote cell fusion and intracellular delivery of the macromolecule. Alternatively, TRICH or a fragment thereof may be joined to a short cationic N-terminal portion from the HIV Tat-1 protein. Fusion proteins thus generated have been found to transduce into the cells of all tissues, including the brain, in a mouse model system (Schwarze, S.R. et al. (1999) Science 285:1569-1572).

For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays, e.g., of neoplastic cells, or in animal models such as mice, rats, rabbits, dogs, monkeys, or pigs. An animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of active ingredient, for example TRICH or fragments thereof, antibodies of TRICH, and agonists, antagonists or inhibitors of TRICH, which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity may be determined by standard pharmaceutical procedures in cell cultures or with experimental animals, such as by calculating the ED₅₀ (the dose therapeutically effective in 50% of the population) or LD₅₀ (the dose lethal to 50% of the population) statistics. The dose ratio of toxic to therapeutic effects is the therapeutic index, which can be expressed as the LD₅₀/ED₅₀ ratio. Compositions which exhibit large therapeutic indices are preferred. The data obtained from cell culture assays and animal studies are

used to formulate a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that includes the ED₅₀ with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, the sensitivity of the patient, and the route of administration.

The exact dosage will be determined by the practitioner, in light of factors related to the subject requiring treatment. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Factors which may be taken into account include the severity of the disease state, the general health of the subject, the age, weight, and gender of the subject, time and frequency of administration, drug combination(s), reaction sensitivities, and response to therapy. Long-acting compositions may be administered every 3 to 4 days, every week, or biweekly depending on the half-life and clearance rate of the particular formulation.

Normal dosage amounts may vary from about 0.1 μ g to 100,000 μ g, up to a total dose of about 1 gram, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature and generally available to practitioners in the art. Those skilled in the art will employ different formulations for nucleotides than for proteins or their inhibitors. Similarly, delivery of polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc.

DIAGNOSTICS

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In another embodiment, antibodies which specifically bind TRICH may be used for the diagnosis of disorders characterized by expression of TRICH, or in assays to monitor patients being treated with TRICH or agonists, antagonists, or inhibitors of TRICH. Antibodies useful for diagnostic purposes may be prepared in the same manner as described above for therapeutics. Diagnostic assays for TRICH include methods which utilize the antibody and a label to detect TRICH in human body fluids or in extracts of cells or tissues. The antibodies may be used with or without modification, and may be labeled by covalent or non-covalent attachment of a reporter molecule. A wide variety of reporter molecules, several of which are described above, are known in the art and may be used.

A variety of protocols for measuring TRICH, including ELISAs, RIAs, and FACS, are known in the art and provide a basis for diagnosing altered or abnormal levels of TRICH expression. Normal or standard values for TRICH expression are established by combining body fluids or cell extracts taken from normal mammalian subjects, for example, human subjects, with antibodies to TRICH under conditions suitable for complex formation. The amount of standard complex formation may be quantitated by various methods, such as photometric means. Quantities of TRICH expressed in

subject, control, and disease samples from biopsied tissues are compared with the standard values. Deviation between standard and subject values establishes the parameters for diagnosing disease.

In another embodiment of the invention, polynucleotides encoding TRICH may be used for diagnostic purposes. The polynucleotides which may be used include oligonucleotides, complementary RNA and DNA molecules, and PNAs. The polynucleotides may be used to detect and quantify gene expression in biopsied tissues in which expression of TRICH may be correlated with disease. The diagnostic assay may be used to determine absence, presence, and excess expression of TRICH, and to monitor regulation of TRICH levels during therapeutic intervention.

In one aspect, hybridization with PCR probes which are capable of detecting polynucleotides, including genomic sequences, encoding TRICH or closely related molecules may be used to identify nucleic acid sequences which encode TRICH. The specificity of the probe, whether it is made from a highly specific region, e.g., the 5' regulatory region, or from a less specific region, e.g., a conserved motif, and the stringency of the hybridization or amplification will determine whether the probe identifies only naturally occurring sequences encoding TRICH, allelic variants, or related sequences.

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Probes may also be used for the detection of related sequences, and may have at least 50% sequence identity to any of the TRICH encoding sequences. The hybridization probes of the subject invention may be DNA or RNA and may be derived from the sequence of SEQ ID NO:27-52 or from genomic sequences including promoters, enhancers, and introns of the TRICH gene.

Means for producing specific hybridization probes for polynucleotides encoding TRICH include the cloning of polynucleotides encoding TRICH or TRICH derivatives into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes *in vitro* by means of the addition of the appropriate RNA polymerases and the appropriate labeled nucleotides. Hybridization probes may be labeled by a variety of reporter groups, for example, by radionuclides such as ³²P or ³⁵S, or by enzymatic labels, such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems, and the like.

Polynucleotides encoding TRICH may be used for the diagnosis of disorders associated with expression of TRICH. Examples of such disorders include, but are not limited to, a transport disorder such as akinesia, amyotrophic lateral sclerosis, ataxia telangiectasia, cystic fibrosis, Becker's muscular dystrophy, Bell's palsy, Charcot-Marie Tooth disease, diabetes mellitus, diabetes insipidus, diabetic neuropathy, Duchenne muscular dystrophy, hyperkalemic periodic paralysis, normokalemic periodic paralysis, Parkinson's disease, malignant hyperthermia, multidrug resistance, myasthenia gravis, myotonic dystrophy, catatonia, tardive dyskinesia, dystonias, peripheral neuropathy, cerebral

neoplasms, prostate cancer, cardiac disorders associated with transport, e.g., angina, bradyarrythmia, tachyarrythmia, hypertension, Long QT syndrome, myocarditis, cardiomyopathy, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, thyrotoxic myopathy, ethanol myopathy, dermatomyositis, inclusion body myositis, infectious myositis, polymyositis, neurological disorders associated with transport, e.g., Alzheimer's disease, amnesia, bipolar disorder, dementia, depression, epilepsy, Tourette's disorder, paranoid psychoses, and schizophrenia, and other disorders associated with transport, e.g., neurofibromatosis, postherpetic neuralgia, trigeminal neuropathy, sarcoidosis, sickle cell anemia, Wilson's disease, cataracts, infertility, pulmonary artery stenosis, sensorineural autosomal deafness, hyperglycemia, hypoglycemia, Grave's disease, goiter, Cushing's disease, Addison's disease, glucose-galactose malabsorption syndrome, glycogen storage disease, hypercholesterolemia, adrenoleukodystrophy, Zellweger syndrome, Menkes disease, occipital horn syndrome, von Gierke disease, pseudohypoaldosteronism type 1, Liddle's syndrome, cystinuria, iminoglycinuria, Hartup disease, Fanconi disease, and Bartter syndrome; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, hemiplegic migraine, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, Tourette's disorder, progressive supranuclear palsy, corticobasal degeneration, and familial frontotemporal dementia; a muscle disorder such as cardiomyopathy, myocarditis, Duchenne's muscular dystrophy, Becker's muscular dystrophy, myotonic dystrophy, central core disease, nemaline myopathy, centronuclear

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myopathy, lipid myopathy, mitochondrial myopathy, infectious myositis, polymyositis, dermatomyositis, inclusion body myositis, thyrotoxic myopathy, ethanol myopathy, angina, anaphylactic shock, arrhythmias, asthma, cardiovascular shock, Cushing's syndrome, hypertension, hypoglycemia, myocardial infarction, migraine, pheochromocytoma, and myopathies including encephalopathy, epilepsy, Kearns-Sayre syndrome, lactic acidosis, myoclonic disorder, ophthalmoplegia, acid maltase deficiency (AMD, also known as Pompe's disease), generalized myotonia, and myotonia congenita; an immunological disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; and a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus. Polynucleotides encoding TRICH may be used in Southern or northern analysis, dot blot, or other membrane-based technologies; in PCR technologies; in dipstick, pin, and multiformat ELISA-like assays; and in microarrays utilizing fluids or tissues from patients to detect altered TRICH expression. Such qualitative or quantitative methods are well known in the art.

In a particular aspect, polynucleotides encoding TRICH may be used in assays that detect the presence of associated disorders, particularly those mentioned above. Polynucleotides complementary to sequences encoding TRICH may be labeled by standard methods and added to a fluid or tissue

sample from a patient under conditions suitable for the formation of hybridization complexes. After a suitable incubation period, the sample is washed and the signal is quantified and compared with a standard value. If the amount of signal in the patient sample is significantly altered in comparison to a control sample then the presence of altered levels of polynucleotides encoding TRICH in the sample indicates the presence of the associated disorder. Such assays may also be used to evaluate the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials, or to monitor the treatment of an individual patient.

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In order to provide a basis for the diagnosis of a disorder associated with expression of TRICH, a normal or standard profile for expression is established. This may be accomplished by combining body fluids or cell extracts taken from normal subjects, either animal or human, with a sequence, or a fragment thereof, encoding TRICH, under conditions suitable for hybridization or amplification. Standard hybridization may be quantified by comparing the values obtained from normal subjects with values from an experiment in which a known amount of a substantially purified polynucleotide is used. Standard values obtained in this manner may be compared with values obtained from samples from patients who are symptomatic for a disorder. Deviation from standard values is used to establish the presence of a disorder.

Once the presence of a disorder is established and a treatment protocol is initiated, hybridization assays may be repeated on a regular basis to determine if the level of expression in the patient begins to approximate that which is observed in the normal subject. The results obtained from successive assays may be used to show the efficacy of treatment over a period ranging from several days to months.

With respect to cancer, the presence of an abnormal amount of transcript (either under- or overexpressed) in biopsied tissue from an individual may indicate a predisposition for the development of the disease, or may provide a means for detecting the disease prior to the appearance of actual clinical symptoms. A more definitive diagnosis of this type may allow health professionals to employ preventative measures or aggressive treatment earlier, thereby preventing the development or further progression of the cancer.

Additional diagnostic uses for oligonucleotides designed from the sequences encoding TRICH may involve the use of PCR. These oligomers may be chemically synthesized, generated enzymatically, or produced *in vitro*. Oligomers will preferably contain a fragment of a polynucleotide encoding TRICH, or a fragment of a polynucleotide complementary to the polynucleotide encoding TRICH, and will be employed under optimized conditions for identification of a specific gene or

condition. Oligomers may also be employed under less stringent conditions for detection or quantification of closely related DNA or RNA sequences.

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In a particular aspect, oligonucleotide primers derived from polynucleotides encoding TRICH may be used to detect single nucleotide polymorphisms (SNPs). SNPs are substitutions, insertions and deletions that are a frequent cause of inherited or acquired genetic disease in humans. Methods of SNP detection include, but are not limited to, single-stranded conformation polymorphism (SSCP) and fluorescent SSCP (fSSCP) methods. In SSCP, oligonucleotide primers derived from polynucleotides encoding TRICH are used to amplify DNA using the polymerase chain reaction (PCR). The DNA may be derived, for example, from diseased or normal tissue, biopsy samples, bodily fluids, and the like. SNPs in the DNA cause differences in the secondary and tertiary structures of PCR products in single-stranded form, and these differences are detectable using gel electrophoresis in non-denaturing gels. In fSCCP, the oligonucleotide primers are fluorescently labeled, which allows detection of the amplimers in high-throughput equipment such as DNA sequencing machines. Additionally, sequence database analysis methods, termed in silico SNP (isSNP), are capable of identifying polymorphisms by comparing the sequence of individual overlapping DNA fragments which assemble into a common consensus sequence. These computer-based methods filter out sequence variations due to laboratory preparation of DNA and sequencing errors using statistical models and automated analyses of DNA sequence chromatograms. In the alternative, SNPs may be detected and characterized by mass spectrometry using, for example, the high throughput MASSARRAY system (Sequenom, Inc., San Diego CA).

SNPs may be used to study the genetic basis of human disease. For example, at least 16 common SNPs have been associated with non-insulin-dependent diabetes mellitus. SNPs are also useful for examining differences in disease outcomes in monogenic disorders, such as cystic fibrosis, sickle cell anemia, or chronic granulomatous disease. For example, variants in the mannose-binding lectin, MBL2, have been shown to be correlated with deleterious pulmonary outcomes in cystic fibrosis. SNPs also have utility in pharmacogenomics, the identification of genetic variants that influence a patient's response to a drug, such as life-threatening toxicity. For example, a variation in N-acetyl transferase is associated with a high incidence of peripheral neuropathy in response to the anti-tuberculosis drug isoniazid, while a variation in the core promoter of the ALOX5 gene results in diminished clinical response to treatment with an anti-asthma drug that targets the 5-lipoxygenase pathway. Analysis of the distribution of SNPs in different populations is useful for investigating genetic drift, mutation, recombination, and selection, as well as for tracing the origins of populations

and their migrations (Taylor, J.G. et al. (2001) Trends Mol. Med. 7:507-512; Kwok, P.-Y. and Z. Gu (1999) Mol. Med. Today 5:538-543; Nowotny, P. et al. (2001) Curr. Opin. Neurobiol. 11:637-641).

Methods which may also be used to quantify the expression of TRICH include radiolabeling or biotinylating nucleotides, coamplification of a control nucleic acid, and interpolating results from standard curves (Melby, P.C. et al. (1993) J. Immunol. Methods 159:235-244; Duplaa, C. et al. (1993) Anal. Biochem. 212:229-236). The speed of quantitation of multiple samples may be accelerated by running the assay in a high-throughput format where the oligomer or polynucleotide of interest is presented in various dilutions and a spectrophotometric or colorimetric response gives rapid quantitation.

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In further embodiments, oligonucleotides or longer fragments derived from any of the polynucleotides described herein may be used as elements on a microarray. The microarray can be used in transcript imaging techniques which monitor the relative expression levels of large numbers of genes simultaneously as described below. The microarray may also be used to identify genetic variants, mutations, and polymorphisms. This information may be used to determine gene function, to understand the genetic basis of a disorder, to diagnose a disorder, to monitor progression/regression of disease as a function of gene expression, and to develop and monitor the activities of therapeutic agents in the treatment of disease. In particular, this information may be used to develop a pharmacogenomic profile of a patient in order to select the most appropriate and effective treatment regimen for that patient. For example, therapeutic agents which are highly effective and display the fewest side effects may be selected for a patient based on his/her pharmacogenomic profile.

In another embodiment, TRICH, fragments of TRICH, or antibodies specific for TRICH may be used as elements on a microarray. The microarray may be used to monitor or measure protein-protein interactions, drug-target interactions, and gene expression profiles, as described above.

A particular embodiment relates to the use of the polynucleotides of the present invention to generate a transcript image of a tissue or cell type. A transcript image represents the global pattern of gene expression by a particular tissue or cell type. Global gene expression patterns are analyzed by quantifying the number of expressed genes and their relative abundance under given conditions and at a given time (Seilhamer et al., "Comparative Gene Transcript Analysis," U.S. Patent No. 5,840,484; hereby expressly incorporated by reference herein). Thus a transcript image may be generated by hybridizing the polynucleotides of the present invention or their complements to the totality of transcripts or reverse transcripts of a particular tissue or cell type. In one embodiment, the hybridization takes place in high-throughput format, wherein the polynucleotides of the present

invention or their complements comprise a subset of a plurality of elements on a microarray. The resultant transcript image would provide a profile of gene activity.

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Transcript images may be generated using transcripts isolated from tissues, cell lines, biopsies, or other biological samples. The transcript image may thus reflect gene expression *in vivo*, as in the case of a tissue or biopsy sample, or *in vitro*, as in the case of a cell line.

Transcript images which profile the expression of the polynucleotides of the present invention may also be used in conjunction with in vitro model systems and preclinical evaluation of pharmaceuticals, as well as toxicological testing of industrial and naturally-occurring environmental compounds. All compounds induce characteristic gene expression patterns, frequently termed molecular fingerprints or toxicant signatures, which are indicative of mechanisms of action and toxicity (Nuwaysir, E.F. et al. (1999) Mol. Carcinog. 24:153-159; Steiner, S. and N.L. Anderson (2000) Toxicol. Lett. 112-113:467-471). If a test compound has a signature similar to that of a compound with known toxicity, it is likely to share those toxic properties. These fingerprints or signatures are most useful and refined when they contain expression information from a large number of genes and gene families. Ideally, a genome-wide measurement of expression provides the highest quality signature. Even genes whose expression is not altered by any tested compounds are important as well, as the levels of expression of these genes are used to normalize the rest of the expression data. The normalization procedure is useful for comparison of expression data after treatment with different compounds. While the assignment of gene function to elements of a toxicant signature aids in interpretation of toxicity mechanisms, knowledge of gene function is not necessary for the statistical matching of signatures which leads to prediction of toxicity (see, for example, Press Release 00-02 from the National Institute of Environmental Health Sciences, released February 29, 2000, available at http://www.niehs.nih.gov/oc/news/toxchip.htm). Therefore, it is important and desirable in toxicological screening using toxicant signatures to include all expressed gene sequences.

In an embodiment, the toxicity of a test compound can be assessed by treating a biological sample containing nucleic acids with the test compound. Nucleic acids that are expressed in the treated biological sample are hybridized with one or more probes specific to the polynucleotides of the present invention, so that transcript levels corresponding to the polynucleotides of the present invention may be quantified. The transcript levels in the treated biological sample are compared with levels in an untreated biological sample. Differences in the transcript levels between the two samples are indicative of a toxic response caused by the test compound in the treated sample.

Another embodiment relates to the use of the polypeptides disclosed herein to analyze the

proteome of a tissue or cell type. The term proteome refers to the global pattern of protein expression in a particular tissue or cell type. Each protein component of a proteome can be subjected individually to further analysis. Proteome expression patterns, or profiles, are analyzed by quantifying the number of expressed proteins and their relative abundance under given conditions and at a given time. A profile of a cell's proteome may thus be generated by separating and analyzing the polypeptides of a particular tissue or cell type. In one embodiment, the separation is achieved using two-dimensional gel electrophoresis, in which proteins from a sample are separated by isoelectric focusing in the first dimension, and then according to molecular weight by sodium dodecyl sulfate slab gel electrophoresis in the second dimension (Steiner and Anderson, supra). The proteins are visualized in the gel as discrete and uniquely positioned spots, typically by staining the gel with an agent such as Coomassie Blue or silver or fluorescent stains. The optical density of each protein spot is generally proportional to the level of the protein in the sample. The optical densities of equivalently positioned protein spots from different samples, for example, from biological samples either treated or untreated with a test compound or therapeutic agent, are compared to identify any changes in protein spot density related to the treatment. The proteins in the spots are partially sequenced using, for example, standard methods employing chemical or enzymatic cleavage followed by mass spectrometry. The identity of the protein in a spot may be determined by comparing its partial sequence, preferably of at least 5 contiguous amino acid residues, to the polypeptide sequences of interest. In some cases, further sequence data may be obtained for definitive protein identification.

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A proteomic profile may also be generated using antibodies specific for TRICH to quantify the levels of TRICH expression. In one embodiment, the antibodies are used as elements on a microarray, and protein expression levels are quantified by exposing the microarray to the sample and detecting the levels of protein bound to each array element (Lucking, A. et al. (1999) Anal. Biochem. 270:103-111; Mendoze, L.G. et al. (1999) Biotechniques 27:778-788). Detection may be performed by a variety of methods known in the art, for example, by reacting the proteins in the sample with a thiolor amino-reactive fluorescent compound and detecting the amount of fluorescence bound at each array element.

Toxicant signatures at the proteome level are also useful for toxicological screening, and should be analyzed in parallel with toxicant signatures at the transcript level. There is a poor correlation between transcript and protein abundances for some proteins in some tissues (Anderson, N.L. and J. Seilhamer (1997) Electrophoresis 18:533-537), so proteome toxicant signatures may be useful in the analysis of compounds which do not significantly affect the transcript image, but which

alter the proteomic profile. In addition, the analysis of transcripts in body fluids is difficult, due to rapid degradation of mRNA, so proteomic profiling may be more reliable and informative in such cases.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins that are expressed in the treated biological sample are separated so that the amount of each protein can be quantified. The amount of each protein is compared to the amount of the corresponding protein in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample. Individual proteins are identified by sequencing the amino acid residues of the individual proteins and comparing these partial sequences to the polypeptides of the present invention.

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In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins from the biological sample are incubated with antibodies specific to the polypeptides of the present invention. The amount of protein recognized by the antibodies is quantified. The amount of protein in the treated biological sample is compared with the amount in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample.

Microarrays may be prepared, used, and analyzed using methods known in the art (Brennan, T.M. et al. (1995) U.S. Patent No. 5,474,796; Schena, M. et al. (1996) Proc. Natl. Acad. Sci. USA 93:10614-10619; Baldeschweiler et al. (1995) PCT application WO95/251116; Shalon, D. et al. (1995) PCT application WO95/35505; Heller, R.A. et al. (1997) Proc. Natl. Acad. Sci. USA 94:2150-2155; Heller, M.J. et al. (1997) U.S. Patent No. 5,605,662). Various types of microarrays are well known and thoroughly described in Schena, M., ed. (1999; <u>DNA Microarrays: A Practical Approach</u>, Oxford University Press, London).

In another embodiment of the invention, nucleic acid sequences encoding TRICH may be used to generate hybridization probes useful in mapping the naturally occurring genomic sequence. Either coding or noncoding sequences may be used, and in some instances, noncoding sequences may be preferable over coding sequences. For example, conservation of a coding sequence among members of a multi-gene family may potentially cause undesired cross hybridization during chromosomal mapping. The sequences may be mapped to a particular chromosome, to a specific region of a chromosome, or to artificial chromosome constructions, e.g., human artificial chromosomes (HACs), yeast artificial chromosomes (YACs), bacterial artificial chromosomes (BACs), bacterial P1 constructions, or single chromosome cDNA libraries (Harrington, J.J. et al. (1997) Nat. Genet. 15:345-

355; Price, C.M. (1993) Blood Rev. 7:127-134; Trask, B.J. (1991) Trends Genet. 7:149-154). Once mapped, the nucleic acid sequences may be used to develop genetic linkage maps, for example, which correlate the inheritance of a disease state with the inheritance of a particular chromosome region or restriction fragment length polymorphism (RFLP) (Lander, E.S. and D. Botstein (1986) Proc. Natl. Acad. Sci. USA 83:7353-7357).

Fluorescent in situ hybridization (FISH) may be correlated with other physical and genetic map data (Heinz-Ulrich, et al. (1995) in Meyers, supra, pp. 965-968). Examples of genetic map data can be found in various scientific journals or at the Online Mendelian Inheritance in Man (OMIM) World Wide Web site. Correlation between the location of the gene encoding TRICH on a physical map and a specific disorder, or a predisposition to a specific disorder, may help define the region of DNA associated with that disorder and thus may further positional cloning efforts.

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In situ hybridization of chromosomal preparations and physical mapping techniques, such as linkage analysis using established chromosomal markers, may be used for extending genetic maps. Often the placement of a gene on the chromosome of another mammalian species, such as mouse, may reveal associated markers even if the exact chromosomal locus is not known. This information is valuable to investigators searching for disease genes using positional cloning or other gene discovery techniques. Once the gene or genes responsible for a disease or syndrome have been crudely localized by genetic linkage to a particular genomic region, e.g., ataxia-telangiectasia to 11q22-23, any sequences mapping to that area may represent associated or regulatory genes for further investigation (Gatti, R.A. et al. (1988) Nature 336:577-580). The nucleotide sequence of the instant invention may also be used to detect differences in the chromosomal location due to translocation, inversion, etc., among normal, carrier, or affected individuals.

In another embodiment of the invention, TRICH, its catalytic or immunogenic fragments, or oligopeptides thereof can be used for screening libraries of compounds in any of a variety of drug screening techniques. The fragment employed in such screening may be free in solution, affixed to a solid support, borne on a cell surface, or located intracellularly. The formation of binding complexes between TRICH and the agent being tested may be measured.

Another technique for drug screening provides for high throughput screening of compounds having suitable binding affinity to the protein of interest (Geysen, et al. (1984) PCT application WO84/03564). In this method, large numbers of different small test compounds are synthesized on a solid substrate. The test compounds are reacted with TRICH, or fragments thereof, and washed. Bound TRICH is then detected by methods well known in the art. Purified TRICH can also be

coated directly onto plates for use in the aforementioned drug screening techniques. Alternatively, non-neutralizing antibodies can be used to capture the peptide and immobilize it on a solid support.

In another embodiment, one may use competitive drug screening assays in which neutralizing antibodies capable of binding TRICH specifically compete with a test compound for binding TRICH. In this manner, antibodies can be used to detect the presence of any peptide which shares one or more antigenic determinants with TRICH.

In additional embodiments, the nucleotide sequences which encode TRICH may be used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleotide sequences that are currently known, including, but not limited to, such properties as the triplet genetic code and specific base pair interactions.

Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following embodiments are, therefore, to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever.

Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following preferred specific embodiments are, therefore, to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever.

The disclosures of all patents, applications, and publications mentioned above and below, including U.S. Ser. No. 60/313,242, U.S. Ser. No. 60/324,782, U.S. Ser. No. 60/328,184, U.S. Ser. No. 60/345,937, U.S. Ser. No. 60/335,698, U.S. Ser. No. 60/332,804, U.S. Ser. No. 60/333,922, U.S. Ser. No. 60/388,180, U.S. Ser. No. 60/375,637, and U.S. Ser. No. 60/377,444, are hereby expressly incorporated by reference.

25 EXAMPLES

I. Construction of cDNA Libraries

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Incyte cDNAs were derived from cDNA libraries described in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA). Some tissues were homogenized and lysed in guanidinium isothiocyanate, while others were homogenized and lysed in phenol or in a suitable mixture of denaturants, such as TRIZOL (Invitrogen), a monophasic solution of phenol and guanidine isothiocyanate. The resulting lysates were centrifuged over CsCl cushions or extracted with chloroform. RNA was precipitated from the lysates with either isopropanol or sodium acetate and

ethanol, or by other routine methods.

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Phenol extraction and precipitation of RNA were repeated as necessary to increase RNA purity. In some cases, RNA was treated with DNase. For most libraries, poly(A)+ RNA was isolated using oligo d(T)-coupled paramagnetic particles (Promega), OLIGOTEX latex particles (QIAGEN, Chatsworth CA), or an OLIGOTEX mRNA purification kit (QIAGEN). Alternatively, RNA was isolated directly from tissue lysates using other RNA isolation kits, e.g., the POLY(A)PURE mRNA purification kit (Ambion, Austin TX).

In some cases, Stratagene was provided with RNA and constructed the corresponding cDNA libraries. Otherwise, cDNA was synthesized and cDNA libraries were constructed with the UNIZAP vector system (Stratagene) or SUPERSCRIPT plasmid system (Invitrogen), using the recommended procedures or similar methods known in the art (Ausubel et al., supra, ch. 5). Reverse transcription was initiated using oligo d(T) or random primers. Synthetic oligonucleotide adapters were ligated to double stranded cDNA, and the cDNA was digested with the appropriate restriction enzyme or enzymes. For most libraries, the cDNA was size-selected (300-1000 bp) using SEPHACRYL S1000, SEPHAROSE CL2B, or SEPHAROSE CL4B column chromatography (Amersham Biosciences) or preparative agarose gel electrophoresis. cDNAs were ligated into compatible restriction enzyme sites of the polylinker of a suitable plasmid, e.g., PBLUESCRIPT plasmid (Stratagene), PSPORT1 plasmid (Invitrogen), PCDNA2.1 plasmid (Invitrogen, Carlsbad CA), PBK-CMV plasmid (Stratagene), PCR2-TOPOTA plasmid (Invitrogen), PCMV-ICIS plasmid (Stratagene), pIGEN (Incyte Genomics, Palo Alto CA), pRARE (Incyte Genomics), or pINCY (Incyte Genomics), or derivatives thereof. Recombinant plasmids were transformed into competent E. coli cells including XL1-Blue, XL1-BlueMRF, or SOLR from Stratagene or DH5α, DH10B, or ElectroMAX DH10B from Invitrogen.

II. Isolation of cDNA Clones

Plasmids obtained as described in Example I were recovered from host cells by *in vivo* excision using the UNIZAP vector system (Stratagene) or by cell lysis. Plasmids were purified using at least one of the following: a Magic or WIZARD Minipreps DNA purification system (Promega); an AGTC Miniprep purification kit (Edge Biosystems, Gaithersburg MD); and QIAWELL 8 Plasmid, QIAWELL 8 Plasmid, QIAWELL 8 Ultra Plasmid purification systems or the R.E.A.L. PREP 96 plasmid purification kit from QIAGEN. Following precipitation, plasmids were resuspended in 0.1 ml of distilled water and stored, with or without lyophilization, at 4°C.

Alternatively, plasmid DNA was amplified from host cell lysates using direct link PCR in a

high-throughput format (Rao, V.B. (1994) Anal. Biochem. 216:1-14). Host cell lysis and thermal cycling steps were carried out in a single reaction mixture. Samples were processed and stored in 384-well plates, and the concentration of amplified plasmid DNA was quantified fluorometrically using PICOGREEN dye (Molecular Probes, Eugene OR) and a FLUOROSKAN II fluorescence scanner (Labsystems Oy, Helsinki, Finland).

III. Sequencing and Analysis

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Incyte cDNA recovered in plasmids as described in Example II were sequenced as follows. Sequencing reactions were processed using standard methods or high-throughput instrumentation such as the ABI CATALYST 800 (Applied Biosystems) thermal cycler or the PTC-200 thermal cycler (MJ Research) in conjunction with the HYDRA microdispenser (Robbins Scientific) or the MICROLAB 2200 (Hamilton) liquid transfer system. cDNA sequencing reactions were prepared using reagents provided by Amersham Biosciences or supplied in ABI sequencing kits such as the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems). Electrophoretic separation of cDNA sequencing reactions and detection of labeled polynucleotides were carried out using the MEGABACE 1000 DNA sequencing system (Amersham Biosciences); the ABI PRISM 373 or 377 sequencing system (Applied Biosystems) in conjunction with standard ABI protocols and base calling software; or other sequence analysis systems known in the art. Reading frames within the cDNA sequences were identified using standard methods (Ausubel et al., supra, ch. 7). Some of the cDNA sequences were selected for extension using the techniques disclosed in Example VIII.

The polynucleotide sequences derived from Incyte cDNAs were validated by removing vector, linker, and poly(A) sequences and by masking ambiguous bases, using algorithms and programs based on BLAST, dynamic programming, and dinucleotide nearest neighbor analysis. The Incyte cDNA sequences or translations thereof were then queried against a selection of public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases, and BLOCKS, PRINTS, DOMO, PRODOM; PROTEOME databases with sequences from *Homo sapiens*, *Rattus norvegicus*, *Mus musculus*, *Caenorhabditis elegans*, *Saccharomyces cerevisiae*, *Schizosaccharomyces pombe*, and *Candida albicans* (Incyte Genomics, Palo Alto CA); hidden Markov model (HMM)-based protein family databases such as PFAM, INCY, and TIGRFAM (Haft, D.H. et al. (2001) Nucleic Acids Res. 29:41-43); and HMM-based protein domain databases such as SMART (Schultz, J. et al. (1998) Proc. Natl. Acad. Sci. USA 95:5857-5864; Letunic, I. et al. (2002) Nucleic Acids Res. 30:242-244). (HMM is a probabilistic approach which analyzes consensus

primary structures of gene families; see, for example, Eddy, S.R. (1996) Curr. Opin. Struct. Biol. 6:361-365.) The queries were performed using programs based on BLAST, FASTA, BLIMPS, and HMMER. The Incyte cDNA sequences were assembled to produce full length polynucleotide sequences. Alternatively, GenBank cDNAs, GenBank ESTs, stitched sequences, stretched sequences, or Genscan-predicted coding sequences (see Examples IV and V) were used to extend Incyte cDNA assemblages to full length. Assembly was performed using programs based on Phred, Phrap, and Consed, and cDNA assemblages were screened for open reading frames using programs based on GeneMark, BLAST, and FASTA. The full length polynucleotide sequences were translated to derive the corresponding full length polypeptide sequences. Alternatively, a polypeptide may begin at any of the methionine residues of the full length translated polypeptide. Full length polypeptide sequences were subsequently analyzed by querying against databases such as the GenBank protein databases (genpept), SwissProt, the PROTEOME databases, BLOCKS, PRINTS, DOMO, PRODOM, Prosite, hidden Markov model (HMM)-based protein family databases such as PFAM, INCY, and TIGRFAM; and HMM-based protein domain databases such as SMART. Full length polynucleotide sequences are also analyzed using MACDNASIS PRO software (MiraiBio, Alameda CA) and LASERGENE software (DNASTAR). Polynucleotide and polypeptide sequence alignments are generated using default parameters specified by the CLUSTAL algorithm as incorporated into the MEGALIGN multisequence alignment program (DNASTAR), which also calculates the percent identity between aligned sequences.

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Table 7 summarizes the tools, programs, and algorithms used for the analysis and assembly of Incyte cDNA and full length sequences and provides applicable descriptions, references, and threshold parameters. The first column of Table 7 shows the tools, programs, and algorithms used, the second column provides brief descriptions thereof, the third column presents appropriate references, all of which are incorporated by reference herein in their entirety, and the fourth column presents, where applicable, the scores, probability values, and other parameters used to evaluate the strength of a match between two sequences (the higher the score or the lower the probability value, the greater the identity between two sequences).

The programs described above for the assembly and analysis of full length polynucleotide and polypeptide sequences were also used to identify polynucleotide sequence fragments from SEQ ID NO:27-52. Fragments from about 20 to about 4000 nucleotides which are useful in hybridization and amplification technologies are described in Table 4, column 2.

IV. Identification and Editing of Coding Sequences from Genomic DNA

Putative transporters and ion channels were initially identified by running the Genscan gene identification program against public genomic sequence databases (e.g., gbpri and gbhtg). Genscan is a general-purpose gene identification program which analyzes genomic DNA sequences from a variety of organisms (Burge, C. and S. Karlin (1997) J. Mol. Biol. 268:78-94; Burge, C. and S. Karlin (1998) Curr. Opin. Struct. Biol. 8:346-354). The program concatenates predicted exons to form an assembled cDNA sequence extending from a methionine to a stop codon. The output of Genscan is a FASTA database of polynucleotide and polypeptide sequences. The maximum range of sequence for Genscan to analyze at once was set to 30 kb. To determine which of these Genscan predicted cDNA sequences encode transporters and ion channels, the encoded polypeptides were analyzed by querying against PFAM models for transporters and ion channels. Potential transporters and ion channels were also identified by homology to Incyte cDNA sequences that had been annotated as transporters and ion channels. These selected Genscan-predicted sequences were then compared by BLAST analysis to the genpept and gbpri public databases. Where necessary, the Genscan-predicted sequences were then edited by comparison to the top BLAST hit from genpept to correct errors in the sequence predicted by Genscan, such as extra or omitted exons. BLAST analysis was also used to find any Incyte cDNA or public cDNA coverage of the Genscan-predicted sequences, thus providing evidence for transcription. When Incyte cDNA coverage was available, this information was used to correct or confirm the Genscan predicted sequence. Full length polynucleotide sequences were obtained by assembling Genscan-predicted coding sequences with Incyte cDNA sequences and/or public cDNA sequences using the assembly process described in Example III. Alternatively, full length polynucleotide sequences were derived entirely from edited or unedited Genscan-predicted coding sequences.

V. Assembly of Genomic Sequence Data with cDNA Sequence Data "Stitched" Sequences

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Partial cDNA sequences were extended with exons predicted by the Genscan gene identification program described in Example IV. Partial cDNAs assembled as described in Example III were mapped to genomic DNA and parsed into clusters containing related cDNAs and Genscan exon predictions from one or more genomic sequences. Each cluster was analyzed using an algorithm based on graph theory and dynamic programming to integrate cDNA and genomic information, generating possible splice variants that were subsequently confirmed, edited, or extended to create a full length sequence. Sequence intervals in which the entire length of the interval was present on more than one sequence in the cluster were identified, and intervals thus identified were considered to

be equivalent by transitivity. For example, if an interval was present on a cDNA and two genomic sequences, then all three intervals were considered to be equivalent. This process allows unrelated but consecutive genomic sequences to be brought together, bridged by cDNA sequence. Intervals thus identified were then "stitched" together by the stitching algorithm in the order that they appear along their parent sequences to generate the longest possible sequence, as well as sequence variants. Linkages between intervals which proceed along one type of parent sequence (cDNA to cDNA or genomic sequence) were given preference over linkages which change parent type (cDNA to genomic sequence). The resultant stitched sequences were translated and compared by BLAST analysis to the genpept and gbpri public databases. Incorrect exons predicted by Genscan were corrected by comparison to the top BLAST hit from genpept. Sequences were further extended with additional cDNA sequences, or by inspection of genomic DNA, when necessary.

"Stretched" Sequences

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Partial DNA sequences were extended to full length with an algorithm based on BLAST analysis. First, partial cDNAs assembled as described in Example III were queried against public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases using the BLAST program. The nearest GenBank protein homolog was then compared by BLAST analysis to either Incyte cDNA sequences or GenScan exon predicted sequences described in Example IV. A chimeric protein was generated by using the resultant high-scoring segment pairs (HSPs) to map the translated sequences onto the GenBank protein homolog. Insertions or deletions may occur in the chimeric protein with respect to the original GenBank protein homolog. The GenBank protein homolog, the chimeric protein, or both were used as probes to search for homologous genomic sequences from the public human genome databases. Partial DNA sequences were therefore "stretched" or extended by the addition of homologous genomic sequences. The resultant stretched sequences were examined to determine whether it contained a complete gene.

VI. Chromosomal Mapping of TRICH Encoding Polynucleotides

The sequences which were used to assemble SEQ ID NO:27-52 were compared with sequences from the Incyte LIFESEQ database and public domain databases using BLAST and other implementations of the Smith-Waterman algorithm. Sequences from these databases that matched SEQ ID NO:27-52 were assembled into clusters of contiguous and overlapping sequences using assembly algorithms such as Phrap (Table 7). Radiation hybrid and genetic mapping data available from public resources such as the Stanford Human Genome Center (SHGC), Whitehead Institute for Genome Research (WIGR), and Généthon were used to determine if any of the clustered sequences

had been previously mapped. Inclusion of a mapped sequence in a cluster resulted in the assignment of all sequences of that cluster, including its particular SEQ ID NO:, to that map location.

Map locations are represented by ranges, or intervals, of human chromosomes. The map position of an interval, in centiMorgans, is measured relative to the terminus of the chromosome's parm. (The centiMorgan (cM) is a unit of measurement based on recombination frequencies between chromosomal markers. On average, 1 cM is roughly equivalent to 1 megabase (Mb) of DNA in humans, although this can vary widely due to hot and cold spots of recombination.) The cM distances are based on genetic markers mapped by Généthon which provide boundaries for radiation hybrid markers whose sequences were included in each of the clusters. Human genome maps and other resources available to the public, such as the NCBI "GeneMap'99" World Wide Web site (http://www.ncbi.nlm.nih.gov/genemap/), can be employed to determine if previously identified disease genes map within or in proximity to the intervals indicated above.

VII. Analysis of Polynucleotide Expression

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Northern analysis is a laboratory technique used to detect the presence of a transcript of a gene and involves the hybridization of a labeled nucleotide sequence to a membrane on which RNAs from a particular cell type or tissue have been bound (Sambrook, *supra*, ch. 7; Ausubel et al., *supra*, ch. 4).

Analogous computer techniques applying BLAST were used to search for identical or related molecules in databases such as GenBank or LIFESEQ (Incyte Genomics). This analysis is much faster than multiple membrane-based hybridizations. In addition, the sensitivity of the computer search can be modified to determine whether any particular match is categorized as exact or similar. The basis of the search is the product score, which is defined as:

BLAST Score x Percent Identity

5 x minimum {length(Seq. 1), length(Seq. 2)}

The product score takes into account both the degree of similarity between two sequences and the length of the sequence match. The product score is a normalized value between 0 and 100, and is calculated as follows: the BLAST score is multiplied by the percent nucleotide identity and the product is divided by (5 times the length of the shorter of the two sequences). The BLAST score is calculated by assigning a score of +5 for every base that matches in a high-scoring segment pair (HSP), and -4 for every mismatch. Two sequences may share more than one HSP (separated by

gaps). If there is more than one HSP, then the pair with the highest BLAST score is used to calculate the product score. The product score represents a balance between fractional overlap and quality in a BLAST alignment. For example, a product score of 100 is produced only for 100% identity over the entire length of the shorter of the two sequences being compared. A product score of 70 is produced either by 100% identity and 70% overlap at one end, or by 88% identity and 100% overlap at the other. A product score of 50 is produced either by 100% identity and 50% overlap at one end, or 79% identity and 100% overlap.

Alternatively, polynucleotides encoding TRICH are analyzed with respect to the tissue sources from which they were derived. For example, some full length sequences are assembled, at least in part, with overlapping Incyte cDNA sequences (see Example III). Each cDNA sequence is derived from a cDNA library constructed from a human tissue. Each human tissue is classified into one of the following organ/tissue categories: cardiovascular system; connective tissue; digestive system; embryonic structures; endocrine system; exocrine glands; genitalia, female; genitalia, male; germ cells; hemic and immune system; liver; musculoskeletal system; nervous system; pancreas; respiratory system; sense organs; skin; stomatognathic system; unclassified/mixed; or urinary tract. The number of libraries in each category is counted and divided by the total number of libraries across all categories. Similarly, each human tissue is classified into one of the following disease/condition categories: cancer, cell line, developmental, inflammation, neurological, trauma, cardiovascular, pooled, and other, and the number of libraries in each category is counted and divided by the total number of libraries across all categories. The resulting percentages reflect the tissue- and disease-specific expression of cDNA encoding TRICH. cDNA sequences and cDNA library/tissue information are found in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA).

VIII. Extension of TRICH Encoding Polynucleotides

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Full length polynucleotides are produced by extension of an appropriate fragment of the full length molecule using oligonucleotide primers designed from this fragment. One primer was synthesized to initiate 5' extension of the known fragment, and the other primer was synthesized to initiate 3' extension of the known fragment. The initial primers were designed using OLIGO 4.06 software (National Biosciences), or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the target sequence at temperatures of about 68 °C to about 72 °C. Any stretch of nucleotides which would result in hairpin structures and primer-primer dimerizations was avoided.

Selected human cDNA libraries were used to extend the sequence. If more than one

extension was necessary or desired, additional or nested sets of primers were designed.

High fidelity amplification was obtained by PCR using methods well known in the art. PCR was performed in 96-well plates using the PTC-200 thermal cycler (MJ Research, Inc.). The reaction mix contained DNA template, 200 nmol of each primer, reaction buffer containing Mg²⁺, (NH₄)₂SO₄, and 2-mercaptoethanol, Taq DNA polymerase (Amersham Biosciences), ELONGASE enzyme (Invitrogen), and Pfu DNA polymerase (Stratagene), with the following parameters for primer pair PCI A and PCI B: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C. In the alternative, the parameters for primer pair T7 and SK+ were as follows: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 57°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C.

The concentration of DNA in each well was determined by dispensing 100 μ l PICOGREEN quantitation reagent (0.25% (v/v) PICOGREEN; Molecular Probes, Eugene OR) dissolved in 1X TE and 0.5 μ l of undiluted PCR product into each well of an opaque fluorimeter plate (Corning Costar, Acton MA), allowing the DNA to bind to the reagent. The plate was scanned in a Fluoroskan II (Labsystems Oy, Helsinki, Finland) to measure the fluorescence of the sample and to quantify the concentration of DNA. A 5 μ l to 10 μ l aliquot of the reaction mixture was analyzed by electrophoresis on a 1% agarose gel to determine which reactions were successful in extending the sequence.

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The extended nucleotides were desalted and concentrated, transferred to 384-well plates, digested with CviJI cholera virus endonuclease (Molecular Biology Research, Madison WI), and sonicated or sheared prior to religation into pUC 18 vector (Amersham Biosciences). For shotgun sequencing, the digested nucleotides were separated on low concentration (0.6 to 0.8%) agarose gels, fragments were excised, and agar digested with Agar ACE (Promega). Extended clones were religated using T4 ligase (New England Biolabs, Beverly MA) into pUC 18 vector (Amersham Biosciences), treated with Pfu DNA polymerase (Stratagene) to fill-in restriction site overhangs, and transfected into competent *E. coli* cells. Transformed cells were selected on antibiotic-containing media, and individual colonies were picked and cultured overnight at 37°C in 384-well plates in LB/2x carb liquid media.

The cells were lysed, and DNA was amplified by PCR using Taq DNA polymerase (Amersham Biosciences) and Pfu DNA polymerase (Stratagene) with the following parameters: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 72°C, 2 min; Step 5: steps 2, 3, and

4 repeated 29 times; Step 6: 72 °C, 5 min; Step 7: storage at 4 °C. DNA was quantified by PICOGREEN reagent (Molecular Probes) as described above. Samples with low DNA recoveries were reamplified using the same conditions as described above. Samples were diluted with 20% dimethysulfoxide (1:2, v/v), and sequenced using DYENAMIC energy transfer sequencing primers and the DYENAMIC DIRECT kit (Amersham Biosciences) or the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems).

In like manner, full length polynucleotides are verified using the above procedure or are used to obtain 5' regulatory sequences using the above procedure along with oligonucleotides designed for such extension, and an appropriate genomic library.

IX. Identification of Single Nucleotide Polymorphisms in TRICH Encoding Polynucleotides

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Common DNA sequence variants known as single nucleotide polymorphisms (SNPs) were identified in SEQ ID NO:27-52 using the LIFESEQ database (Incyte Genomics). Sequences from the same gene were clustered together and assembled as described in Example III, allowing the identification of all sequence variants in the gene. An algorithm consisting of a series of filters was used to distinguish SNPs from other sequence variants. Preliminary filters removed the majority of basecall errors by requiring a minimum Phred quality score of 15, and removed sequence alignment errors and errors resulting from improper trimming of vector sequences, chimeras, and splice variants. An automated procedure of advanced chromosome analysis analysed the original chromatogram files in the vicinity of the putative SNP. Clone error filters used statistically generated algorithms to identify errors introduced during laboratory processing, such as those caused by reverse transcriptase, polymerase, or somatic mutation. Clustering error filters used statistically generated algorithms to identify errors resulting from clustering of close homologs or pseudogenes, or due to contamination by non-human sequences. A final set of filters removed duplicates and SNPs found in immunoglobulins or T-cell receptors.

Certain SNPs were selected for further characterization by mass spectrometry using the high throughput MASSARRAY system (Sequenom, Inc.) to analyze allele frequencies at the SNP sites in four different human populations. The Caucasian population comprised 92 individuals (46 male, 46 female), including 83 from Utah, four French, three Venezualan, and two Amish individuals. The African population comprised 194 individuals (97 male, 97 female), all African Americans. The Hispanic population comprised 324 individuals (162 male, 162 female), all Mexican Hispanic. The Asian population comprised 126 individuals (64 male, 62 female) with a reported parental breakdown

of 43% Chinese, 31% Japanese, 13% Korean, 5% Vietnamese, and 8% other Asian. Allele frequencies were first analyzed in the Caucasian population; in some cases those SNPs which showed no allelic variance in this population were not further tested in the other three populations.

X. Labeling and Use of Individual Hybridization Probes

Hybridization probes derived from SEQ ID NO:27-52 are employed to screen cDNAs, genomic DNAs, or mRNAs. Although the labeling of oligonucleotides, consisting of about 20 base pairs, is specifically described, essentially the same procedure is used with larger nucleotide fragments. Oligonucleotides are designed using state-of-the-art software such as OLIGO 4.06 software (National Biosciences) and labeled by combining 50 pmol of each oligomer, 250 μ Ci of $[\gamma^{-32}P]$ adenosine triphosphate (Amersham Biosciences), and T4 polynucleotide kinase (DuPont NEN, Boston MA). The labeled oligonucleotides are substantially purified using a SEPHADEX G-25 superfine size exclusion dextran bead column (Amersham Biosciences). An aliquot containing 10^7 counts per minute of the labeled probe is used in a typical membrane-based hybridization analysis of human genomic DNA digested with one of the following endonucleases: Ase I, Bgl II, Eco RI, Pst I, Xba I, or Pvu II (DuPont NEN).

The DNA from each digest is fractionated on a 0.7% agarose gel and transferred to nylon membranes (Nytran Plus, Schleicher & Schuell, Durham NH). Hybridization is carried out for 16 hours at 40°C. To remove nonspecific signals, blots are sequentially washed at room temperature under conditions of up to, for example, 0.1 x saline sodium citrate and 0.5% sodium dodecyl sulfate. Hybridization patterns are visualized using autoradiography or an alternative imaging means and compared.

XI. Microarrays

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The linkage or synthesis of array elements upon a microarray can be achieved utilizing photolithography, piezoelectric printing (ink-jet printing; see, e.g., Baldeschweiler et al., supra), mechanical microspotting technologies, and derivatives thereof. The substrate in each of the aforementioned technologies should be uniform and solid with a non-porous surface (Schena, M., ed. (1999) DNA Microarrays: A Practical Approach, Oxford University Press, London). Suggested substrates include silicon, silica, glass slides, glass chips, and silicon wafers. Alternatively, a procedure analogous to a dot or slot blot may also be used to arrange and link elements to the surface of a substrate using thermal, UV, chemical, or mechanical bonding procedures. A typical array may be produced using available methods and machines well known to those of ordinary skill in the art and may contain any appropriate number of elements (Schena, M. et al. (1995) Science 270:467-470;

Shalon, D. et al. (1996) Genome Res. 6:639-645; Marshall, A. and J. Hodgson (1998) Nat. Biotechnol. 16:27-31).

Full length cDNAs, Expressed Sequence Tags (ESTs), or fragments or oligomers thereof may comprise the elements of the microarray. Fragments or oligomers suitable for hybridization can be selected using software well known in the art such as LASERGENE software (DNASTAR). The array elements are hybridized with polynucleotides in a biological sample. The polynucleotides in the biological sample are conjugated to a fluorescent label or other molecular tag for ease of detection. After hybridization, nonhybridized nucleotides from the biological sample are removed, and a fluorescence scanner is used to detect hybridization at each array element. Alternatively, laser desorbtion and mass spectrometry may be used for detection of hybridization. The degree of complementarity and the relative abundance of each polynucleotide which hybridizes to an element on the microarray may be assessed. In one embodiment, microarray preparation and usage is described in detail below.

Tissue or Cell Sample Preparation

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Total RNA is isolated from tissue samples using the guanidinium thiocyanate method and poly(A)⁺ RNA is purified using the oligo-(dT) cellulose method. Each poly(A)⁺ RNA sample is reverse transcribed using MMLV reverse-transcriptase, 0.05 pg/μl oligo-(dT) primer (21mer), 1X first strand buffer, 0.03 units/μl RNase inhibitor, 500 μM dATP, 500 μM dGTP, 500 μM dTTP, 40 μM dCTP, 40 μM dCTP-Cy3 (BDS) or dCTP-Cy5 (Amersham Biosciences). The reverse transcription reaction is performed in a 25 ml volume containing 200 ng poly(A)⁺ RNA with GEMBRIGHT kits (Incyte Genomics). Specific control poly(A)⁺ RNAs are synthesized by *in vitro* transcription from non-coding yeast genomic DNA. After incubation at 37°C for 2 hr, each reaction sample (one with Cy3 and another with Cy5 labeling) is treated with 2.5 ml of 0.5M sodium hydroxide and incubated for 20 minutes at 85°C to the stop the reaction and degrade the RNA. Samples are purified using two successive CHROMA SPIN 30 gel filtration spin columns (Clontech, Palo Alto CA) and after combining, both reaction samples are ethanol precipitated using 1 ml of glycogen (1 mg/ml), 60 ml sodium acetate, and 300 ml of 100% ethanol. The sample is then dried to completion using a SpeedVAC (Savant Instruments Inc., Holbrook NY) and resuspended in 14 μl 5X SSC/0.2% SDS.

Microarray Preparation

Sequences of the present invention are used to generate array elements. Each array element is amplified from bacterial cells containing vectors with cloned cDNA inserts. PCR amplification uses primers complementary to the vector sequences flanking the cDNA insert. Array elements are

amplified in thirty cycles of PCR from an initial quantity of 1-2 ng to a final quantity greater than 5 μ g. Amplified array elements are then purified using SEPHACRYL-400 (Amersham Biosciences).

Purified array elements are immobilized on polymer-coated glass slides. Glass microscope slides (Corning) are cleaned by ultrasound in 0.1% SDS and acetone, with extensive distilled water washes between and after treatments. Glass slides are etched in 4% hydrofluoric acid (VWR Scientific Products Corporation (VWR), West Chester PA), washed extensively in distilled water, and coated with 0.05% aminopropyl silane (Sigma) in 95% ethanol. Coated slides are cured in a 110°C oven.

Array elements are applied to the coated glass substrate using a procedure described in U.S. Patent No. 5,807,522, incorporated herein by reference. 1 μ l of the array element DNA, at an average concentration of 100 ng/ μ l, is loaded into the open capillary printing element by a high-speed robotic apparatus. The apparatus then deposits about 5 nl of array element sample per slide.

Microarrays are UV-crosslinked using a STRATALINKER UV-crosslinker (Stratagene). Microarrays are washed at room temperature once in 0.2% SDS and three times in distilled water. Non-specific binding sites are blocked by incubation of microarrays in 0.2% casein in phosphate buffered saline (PBS) (Tropix, Inc., Bedford MA) for 30 minutes at 60°C followed by washes in 0.2% SDS and distilled water as before.

Hybridization

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Hybridization reactions contain 9 μ 1 of sample mixture consisting of 0.2 μ g each of Cy3 and Cy5 labeled cDNA synthesis products in 5X SSC, 0.2% SDS hybridization buffer. The sample mixture is heated to 65°C for 5 minutes and is aliquoted onto the microarray surface and covered with an 1.8 cm² coverslip. The arrays are transferred to a waterproof chamber having a cavity just slightly larger than a microscope slide. The chamber is kept at 100% humidity internally by the addition of 140 μ 1 of 5X SSC in a corner of the chamber. The chamber containing the arrays is incubated for about 6.5 hours at 60°C. The arrays are washed for 10 min at 45°C in a first wash buffer (1X SSC, 0.1% SDS), three times for 10 minutes each at 45°C in a second wash buffer (0.1X SSC), and dried.

Detection

Reporter-labeled hybridization complexes are detected with a microscope equipped with an Innova 70 mixed gas 10 W laser (Coherent, Inc., Santa Clara CA) capable of generating spectral lines at 488 nm for excitation of Cy3 and at 632 nm for excitation of Cy5. The excitation laser light is focused on the array using a 20X microscope objective (Nikon, Inc., Melville NY). The slide containing the array is placed on a computer-controlled X-Y stage on the microscope and raster-

scanned past the objective. The 1.8 cm x 1.8 cm array used in the present example is scanned with a resolution of 20 micrometers.

In two separate scans, a mixed gas multiline laser excites the two fluorophores sequentially. Emitted light is split, based on wavelength, into two photomultiplier tube detectors (PMT R1477, Hamamatsu Photonics Systems, Bridgewater NJ) corresponding to the two fluorophores. Appropriate filters positioned between the array and the photomultiplier tubes are used to filter the signals. The emission maxima of the fluorophores used are 565 nm for Cy3 and 650 nm for Cy5. Each array is typically scanned twice, one scan per fluorophore using the appropriate filters at the laser source, although the apparatus is capable of recording the spectra from both fluorophores simultaneously.

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Expression

The sensitivity of the scans is typically calibrated using the signal intensity generated by a cDNA control species added to the sample mixture at a known concentration. A specific location on the array contains a complementary DNA sequence, allowing the intensity of the signal at that location to be correlated with a weight ratio of hybridizing species of 1:100,000. When two samples from different sources (e.g., representing test and control cells), each labeled with a different fluorophore, are hybridized to a single array for the purpose of identifying genes that are differentially expressed, the calibration is done by labeling samples of the calibrating cDNA with the two fluorophores and adding identical amounts of each to the hybridization mixture.

The output of the photomultiplier tube is digitized using a 12-bit RTI-835H analog-to-digital (A/D) conversion board (Analog Devices, Inc., Norwood MA) installed in an IBM-compatible PC computer. The digitized data are displayed as an image where the signal intensity is mapped using a linear 20-color transformation to a pseudocolor scale ranging from blue (low signal) to red (high signal). The data is also analyzed quantitatively. Where two different fluorophores are excited and measured simultaneously, the data are first corrected for optical crosstalk (due to overlapping emission spectra) between the fluorophores using each fluorophore's emission spectrum.

A grid is superimposed over the fluorescence signal image such that the signal from each spot is centered in each element of the grid. The fluorescence signal within each element is then integrated to obtain a numerical value corresponding to the average intensity of the signal. The software used for signal analysis is the GEMTOOLS gene expression analysis program (Incyte Genomics). Array elements that exhibited at least about a two-fold change in expression, a signal-to-background ratio of at least 2.5, and an element spot size of at least 40% were identified as differentially expressed.

For example, SEQ ID NO:30 showed differential expression in bone osteosarcoma tissues

versus normal osteocytes, as determined by microarray analysis. Messenger RNA from normal human osteoblasts was compared with mRNA from biopsy specimens, osteosarcoma tissues, primary cultures, or metastasized tissues. A normal osteoblast primary culture, NHOst 5488, was chosen as the reference in the initial experiments. One basic set of experiments is defined as the comparison of mRNA from biopsy specimen with that of definitive surgical specimen from the same patient. Extended study of this basic set includes mRNA from primary cell cultures of the definitive surgical specimen, muscle, or cartilage tissue from the same patient. Biopsy specimens, definitive surgical specimens, or lung metastatic tissues from different individuals were also included to reveal individual variability. The expression of SEQ ID NO:30 was increased by at least two-fold in bone osteosarcoma tissues relative to normal osteocytes. Therefore, in various embodiments, SEQ ID NO:30 can be used for one or more of the following: i) monitoring treatment of bone cancer, ii) diagnostic assays for bone cancer, and iii) developing therapeutics and/or other treatments for bone cancer.

In an alternative example, SEQ ID NO:33 showed differential expression in lung squamous carcinoma tissues versus normal lung tissues as determined by microarray analysis. In matched tissue experiments, the expression of SEQ ID NO:33 was decreased by at least two-fold in lung squamous carcinoma tissues relative to grossly uninvolved normal lung tissues from the same donors. Therefore, in various embodiments, SEQ ID NO:33 can be used for one or more of the following: i) monitoring treatment of lung cancer, ii) diagnostic assays for lung cancer, and iii) developing therapeutics and/or other treatments for lung cancer.

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SEQ ID NO:33 also showed differential expression in ovarian adenocarcinoma tissues versus normal ovarian tissues as determined by microarray analysis. The expression of SEQ ID NO:33 was decreased by at least two-fold in ovarian adenocarcinoma tissues relative to grossly uninvolved normal ovarian tissues from the same donor. Therefore, in various embodiments, SEQ ID NO:33 can be used for one or more of the following: i) monitoring treatment of ovarian cancer, ii) diagnostic assays for ovarian cancer, and iii) developing therapeutics and/or other treatments for ovarian cancer.

SEQ ID NO:33 and SEQ ID NO:40 showed differential expression in association with immune and inflammatory responses as determined by microarray analysis. The expression of SEQ ID NO:33 was increased by at least two-fold in human umbilical vein cells treated with PMA and ionomycin relative to untreated human umbilical vein cells. Human umbilical vein cells are derived from the endothelium of the human umbilical vein, and have been used as an experimental model for investigating the functional biology of human endothelial cells in vitro. PMA is a broad activator of

protein kinase C-dependent pathways and ionomycin is a calcium ionophore that permits the entry of calcium in the cell, hence increasing the cytosolic calcium concentration. The expression of SEQ ID NO:40 was increased by at least 2.5-fold in vascular endothelial tissue treated with TNFα and IL-1β compared with untreated vascular endothelial tissue, as determined by microarray analysis. Human coronary artery endothelial cells and human coronary artery smooth muscle cells (BioWhittaker, Inc., San Diego CA) obtained from the same donor were cultured in tissue culture flasks in Endothelium Growth Medium or Smooth Muscle Growth Medium, respectively (BioWhittaker). Cultures at 85% confluency were either treated with recombinant human TNFα and IL-1β (R&D Systems, Minneapolis MN) at 10 ng/ml each for 24 hours at 37° C or were left untreated. Therefore, in various embodiments, SEQ ID NO:33 and SEQ ID NO:40 can each be used for one or more of the following: i) monitoring treatment of immune/inflammatory responses, ii) diagnostic assays for immune/inflammatory responses, and iii) developing therapeutics and/or other treatments for immune/inflammatory responses.

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In an alternative example, SEQ ID NO:38 showed differential expression in breast carcinoma cell lines versus primary mammary epithelial cells as determined by microarray analysis. The breast carcinoma cell lines include MCF7, a breast adenocarcinoma cell line derived from the pleural effusion of a 69-year-old female; T-47D, a breast carcinoma cell line derived from a pleural effusion from a 54-year-old female with an infiltrating ductal carcinoma of the breast; Sk-BR-3, a breast adenocarcinoma cell line isolated from a malignant pleural effusion of a 43-year-old female; MDAmb-231, a metastatic breast tumor cell line derived from the pleural effusion of a 51-year-old female with metastatic breast carcinoma; and MDA-mb-435S, a spindle shaped strain that evolved from a cell line isolated from the pleural effusion of a 31 year old female with metastatic, ductal adenocarcinoma of the breast. The primary mammary epithelial cell line HMEC was derived from normal human mammary tissue (Clonetics, San Diego, CA). All cell cultures were propagated in a chemicallydefined medium, according to the supplier's recommendations and grown to 70-80% confluence prior to RNA isolation. The microarray experiments showed that the expression of SEQ ID NO:38 was decreased by at least two-fold in all five breast carcinoma lines (MCF7, T-47D, Sk-BR-3, MDA-mb-231, and MDA-mb-435S) relative to primary mammary epithelial cells. Therefore, in various embodiments, SEQ ID NO:38 can be used for one or more of the following: i) monitoring treatment of breast cancer, ii) diagnostic assays for breast cancer, and iii) developing therapeutics and/or other treatments for breast cancer.

SEQ ID NO:38 also showed differential expression in certain prostate carcinoma cell lines

versus normal prostate epithelial cells as determined by microarray analysis. The prostate carcinoma cell lines include CA-HPV-10, DU 145, LNCaP, PC-3, and MDAPCa2b. CA-HPV-7 was derived from cells from a 63 year old male with prostate adenocarcinoma and was transformed by transfection with HPV18 DNA. DU 145 was isolated from a metastatic site in the brain of a 69 year old male with widespread metastatic prostate carcinoma. DU 145 has no detectable sensitivity to hormones; forms colonies in semi-solid medium; is only weakly positive for acid phosphatase; and cells are negative for prostate specific antigen (PSA). LNCaP is a prostate carcinoma cell line isolated from a lymph node biopsy of a 50 year old male with metastatic prostate carcinoma. LNCaP expresses PSA, produces prostate acid phosphatase, and expresses androgen receptors. PC-3, a prostate adenocarcinoma cell line, was isolated from a metastatic site in the bone of a 62 year old male with grade IV prostate adenocarcinoma. MDAPCa2b is a prostate adenocarcinoma cell line isolated from a metastatic site in the bone of a 63 year old male. The MDAPCa2b cell line expresses PSA and androgen receptor and is androgen sensitive. The normal epithelial cell line, PrEC, is a primary prostate epithelial cell line isolated from a normal donor. The expression of SEQ ID NO:38 was decreased by at least two-fold in three out of five prostate carcinoma lines (DU 145, LNCaP, and PC-3) relative to cells from the normal prostate epithelial cell line, PrEC. Therefore, in various embodiments, SEQ ID NO:38 can be used for one or more of the following: i) monitoring treatment of prostate cancer, ii) diagnostic assays for prostate cancer, and iii) developing therapeutics and/or other treatments for prostate cancer.

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In addition, SEQ ID NO:38 and SEQ ID NO:43 showed differential expression in toxicology studies as determined by microarray analysis. The expression of SEQ ID NO:43 was increased by at least two-fold in C3A hepatoblastoma cells treated with 1-100 μM beclomethazone as compared with untreated C3A hepatoblastoma cells. The human C3A cell line is a clonal derivative of HepG2/C3 (hepatoma cell line, isolated from a 15-year-old male with liver tumor), which was selected for strong contact inhibition of growth. The use of a clonal population enhances the reproducibility of the cells. C3A cells have many characteristics of primary human hepatocytes in culture: i) expression of insulin receptor and insulin-like growth factor II receptor; ii) secretion of a high ratio of serum albumin compared with α-fetoprotein iii) conversion of ammonia to urea and glutamine; iv) metabolism of aromatic amino acids; and v) proliferation in glucose-free and insulin-free medium. The C3A cell line is now well established as an in vitro model of the mature human liver (Mickelson et al. (1995) Hepatology 22:866-875; Nagendra et al. (1997) Am J Physiol 272:G408-G416). C3A cells were treated with 1-100 μM beclomethazone for 1hr, 3hr, 6hr and compared with untreated cells. In

addition, the expression of SEQ ID NO:34 was increased by at least two-fold in early confluent C3A cells treated with progesterone, beclomethasone, medroxyprogesterone, budesonide, prednisone, dexamethasone, or betamethasone, for 1, 3, or 6 hours, as compared to untreated C3A cells. In addition, the expression of SEQ ID NO:38 was decreased by at least two-fold in early confluent C3A cells treated with progesterone, beclomethasone, medroxyprogesterone, budesonide, prednisone, dexamethasone, or betamethasone, for 1, 3, or 6 hours, as compared to untreated C3A cells. The effects of steroids on liver metabolism are important to the understanding of the pharmacodynamics of drugs. Therefore, in various embodiments, SEQ ID NO:34, SEQ ID NO:38 and SEQ ID NO:43 can each be used for one or more of the following: i) monitoring treatment of liver toxicity, diseases and disorders, ii) diagnostic assays for liver toxicity, diseases and disorders, and iii) developing therapeutics and/or other treatments for liver toxicity, diseases and disorders.

In yet another example, the expression of SEQ ID NO:48 was differentially expressed in a specific region of human brain tissue as compared to pooled brain tissue control. Characterization of region-specific gene expression in the human brain provides a context and background for molecular neurobiology research in general. This knowledge may provide insight into the genetic basis of brain structure and function. The expression of SEQ ID NO:48 was decreased by at least two-fold in normal human amygdala, entorhinal cortex, brain tissue as compared to the normal human pooled brain tissue used as a control. These experiments indicate that SEQ ID NO:48 exhibited significant differential expression patterns using microarray techniques, and further establishes its utility as a diagnostic marker or therapeutic agent which may be useful in neurological disorders. Therefore, in various embodiments, SEQ ID NO:48 can be used for one or more of the following: i) monitoring treatment of neurological disorders, ii) diagnostic assays for neurological disorders, and iii) developing therapeutics and/or other treatments for neurological disorders.

XII. Complementary Polynucleotides

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Sequences complementary to the TRICH-encoding sequences, or any parts thereof, are used to detect, decrease, or inhibit expression of naturally occurring TRICH. Although use of oligonucleotides comprising from about 15 to 30 base pairs is described, essentially the same procedure is used with smaller or with larger sequence fragments. Appropriate oligonucleotides are designed using OLIGO 4.06 software (National Biosciences) and the coding sequence of TRICH. To inhibit transcription, a complementary oligonucleotide is designed from the most unique 5' sequence and used to prevent promoter binding to the coding sequence. To inhibit translation, a complementary oligonucleotide is designed to prevent ribosomal binding to the TRICH-encoding transcript.

XIII. Expression of TRICH

Expression and purification of TRICH is achieved using bacterial or virus-based expression systems. For expression of TRICH in bacteria, cDNA is subcloned into an appropriate vector containing an antibiotic resistance gene and an inducible promoter that directs high levels of cDNA transcription. Examples of such promoters include, but are not limited to, the trp-lac (tac) hybrid promoter and the T5 or T7 bacteriophage promoter in conjunction with the *lac* operator regulatory element. Recombinant vectors are transformed into suitable bacterial hosts, e.g., BL21(DE3). Antibiotic resistant bacteria express TRICH upon induction with isopropyl beta-Dthiogalactopyranoside (IPTG). Expression of TRICH in eukaryotic cells is achieved by infecting insect or mammalian cell lines with recombinant Autographica californica nuclear polyhedrosis virus (AcMNPV), commonly known as baculovirus. The nonessential polyhedrin gene of baculovirus is replaced with cDNA encoding TRICH by either homologous recombination or bacterial-mediated transposition involving transfer plasmid intermediates. Viral infectivity is maintained and the strong polyhedrin promoter drives high levels of cDNA transcription. Recombinant baculovirus is used to infect Spodoptera frugiperda (Sf9) insect cells in most cases, or human hepatocytes, in some cases. Infection of the latter requires additional genetic modifications to baculovirus (Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945).

In most expression systems, TRICH is synthesized as a fusion protein with, e.g., glutathione S-transferase (GST) or a peptide epitope tag, such as FLAG or 6-His, permitting rapid, single-step, affinity-based purification of recombinant fusion protein from crude cell lysates. GST, a 26-kilodalton enzyme from *Schistosoma japonicum*, enables the purification of fusion proteins on immobilized glutathione under conditions that maintain protein activity and antigenicity (Amersham Biosciences). Following purification, the GST moiety can be proteolytically cleaved from TRICH at specifically engineered sites. FLAG, an 8-amino acid peptide, enables immunoaffinity purification using commercially available monoclonal and polyclonal anti-FLAG antibodies (Eastman Kodak). 6-His, a stretch of six consecutive histidine residues, enables purification on metal-chelate resins (QIAGEN). Methods for protein expression and purification are discussed in Ausubel et al. (*supra*, ch. 10 and 16). Purified TRICH obtained by these methods can be used directly in the assays shown in Examples XVII, XVIII, and XIX, where applicable.

XIV. Functional Assays

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TRICH function is assessed by expressing the sequences encoding TRICH at physiologically

elevated levels in mammalian cell culture systems. cDNA is subcloned into a mammalian expression vector containing a strong promoter that drives high levels of cDNA expression. Vectors of choice include PCMV SPORT plasmid (Invitrogen, Carlsbad CA) and PCR3.1 plasmid (Invitrogen), both of which contain the cytomegalovirus promoter. 5-10 μ g of recombinant vector are transiently transfected into a human cell line, for example, an endothelial or hematopoietic cell line, using either liposome formulations or electroporation. 1-2 μ g of an additional plasmid containing sequences encoding a marker protein are co-transfected. Expression of a marker protein provides a means to distinguish transfected cells from nontransfected cells and is a reliable predictor of cDNA expression from the recombinant vector. Marker proteins of choice include, e.g., Green Fluorescent Protein (GFP; Clontech), CD64, or a CD64-GFP fusion protein. Flow cytometry (FCM), an automated, laser optics-based technique, is used to identify transfected cells expressing GFP or CD64-GFP and to evaluate the apoptotic state of the cells and other cellular properties. FCM detects and quantifies the uptake of fluorescent molecules that diagnose events preceding or coincident with cell death. These events include changes in nuclear DNA content as measured by staining of DNA with propidium iodide; changes in cell size and granularity as measured by forward light scatter and 90 degree side light scatter; down-regulation of DNA synthesis as measured by decrease in bromodeoxyuridine uptake; alterations in expression of cell surface and intracellular proteins as measured by reactivity with specific antibodies; and alterations in plasma membrane composition as measured by the binding of fluorescein-conjugated Annexin V protein to the cell surface. Methods in flow cytometry are discussed in Ormerod, M.G. (1994; Flow Cytometry, Oxford, New York NY).

The influence of TRICH on gene expression can be assessed using highly purified populations of cells transfected with sequences encoding TRICH and either CD64 or CD64-GFP. CD64 and CD64-GFP are expressed on the surface of transfected cells and bind to conserved regions of human immunoglobulin G (IgG). Transfected cells are efficiently separated from nontransfected cells using magnetic beads coated with either human IgG or antibody against CD64 (DYNAL, Lake Success NY). mRNA can be purified from the cells using methods well known by those of skill in the art. Expression of mRNA encoding TRICH and other genes of interest can be analyzed by northern analysis or microarray techniques.

XV. Production of TRICH Specific Antibodies

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TRICH substantially purified using polyacrylamide gel electrophoresis (PAGE; see, e.g., Harrington, M.G. (1990) Methods Enzymol. 182:488-495), or other purification techniques, is used to immunize animals (e.g., rabbits, mice, etc.) and to produce antibodies using standard protocols.

Alternatively, the TRICH amino acid sequence is analyzed using LASERGENE software (DNASTAR) to determine regions of high immunogenicity, and a corresponding oligopeptide is synthesized and used to raise antibodies by means known to those of skill in the art. Methods for selection of appropriate epitopes, such as those near the C-terminus or in hydrophilic regions are well described in the art (Ausubel et al., *supra*, ch. 11).

Typically, oligopeptides of about 15 residues in length are synthesized using an ABI 431A peptide synthesizer (Applied Biosystems) using FMOC chemistry and coupled to KLH (Sigma-Aldrich, St. Louis MO) by reaction with N-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS) to increase immunogenicity (Ausubel et al., *supra*). Rabbits are immunized with the oligopeptide-KLH complex in complete Freund's adjuvant. Resulting antisera are tested for antipeptide and anti-TRICH activity by, for example, binding the peptide or TRICH to a substrate, blocking with 1% BSA, reacting with rabbit antisera, washing, and reacting with radio-iodinated goat anti-rabbit IgG.

XVI. Purification of Naturally Occurring TRICH Using Specific Antibodies

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Naturally occurring or recombinant TRICH is substantially purified by immunoaffinity chromatography using antibodies specific for TRICH. An immunoaffinity column is constructed by covalently coupling anti-TRICH antibody to an activated chromatographic resin, such as CNBr-activated SEPHAROSE (Amersham Biosciences). After the coupling, the resin is blocked and washed according to the manufacturer's instructions.

Media containing TRICH are passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of TRICH (e.g., high ionic strength buffers in the presence of detergent). The column is eluted under conditions that disrupt antibody/TRICH binding (e.g., a buffer of pH 2 to pH 3, or a high concentration of a chaotrope, such as urea or thiocyanate ion), and TRICH is collected.

XVII. Identification of Molecules Which Interact with TRICH

TRICH, or biologically active fragments thereof, are labeled with ¹²⁵I Bolton-Hunter reagent (Bolton, A.E. and W.M. Hunter (1973) Biochem. J. 133:529-539). Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the labeled TRICH, washed, and any wells with labeled TRICH complex are assayed. Data obtained using different concentrations of TRICH are used to calculate values for the number, affinity, and association of TRICH with the candidate molecules.

Alternatively, molecules interacting with TRICH are analyzed using the yeast two-hybrid system as described in Fields, S. and O. Song (1989; Nature 340:245-246), or using commercially

available kits based on the two-hybrid system, such as the MATCHMAKER system (Clontech).

TRICH may also be used in the PATHCALLING process (CuraGen Corp., New Haven CT) which employs the yeast two-hybrid system in a high-throughput manner to determine all interactions between the proteins encoded by two large libraries of genes (Nandabalan, K. et al. (2000) U.S. Patent No. 6,057,101).

XVII. Identification of Molecules Which Interact with TRICH

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Molecules which interact with TRICH may include transporter substrates, agonists or antagonists, modulatory proteins such as Gβγ proteins (Reimann, *supra*) or proteins involved in TRICH localization or clustering such as MAGUKs (Craven, *supra*). TRICH, or biologically active fragments thereof, are labeled with ¹²⁵I Bolton-Hunter reagent. (See, e.g., Bolton A.E. and W.M. Hunter (1973) Biochem. J. 133:529-539.) Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the labeled TRICH, washed, and any wells with labeled TRICH complex are assayed. Data obtained using different concentrations of TRICH are used to calculate values for the number, affinity, and association of TRICH with the candidate molecules.

Alternatively, proteins that interact with TRICH are isolated using the yeast 2-hybrid system (Fields, S. and O. Song (1989) Nature 340:245-246). TRICH, or fragments thereof, are expressed as fusion proteins with the DNA binding domain of Gal4 or lexA, and potential interacting proteins are expressed as fusion proteins with an activation domain. Interactions between the TRICH fusion protein and the TRICH interacting proteins (fusion proteins with an activation domain) reconstitute a transactivation function that is observed by expression of a reporter gene. Yeast 2-hybrid systems are commercially available, and methods for use of the yeast 2-hybrid system with ion channel proteins are discussed in Niethammer, M. and M. Sheng (1998, Meth. Enzymol. 293:104-122).

TRICH may also be used in the PATHCALLING process (CuraGen Corp., New Haven CT) which employs the yeast two-hybrid system in a high-throughput manner to determine all interactions between the proteins encoded by two large libraries of genes (Nandabalan, K. et al. (2000) U.S. Patent No. 6,057,101).

Potential TRICH agonists or antagonists may be tested for activation or inhibition of TRICH ion channel activity using the assays described in section XVIII.

XVIII. Demonstration of TRICH Activity

Ion channel activity of TRICH is demonstrated using an electrophysiological assay for ion conductance. TRICH can be expressed by transforming a mammalian cell line such as COS7, HeLa or CHO with a eukaryotic expression vector encoding TRICH. Eukaryotic expression vectors are

commercially available, and the techniques to introduce them into cells are well known to those skilled in the art. A second plasmid which expresses any one of a number of marker genes, such as β -galactosidase, is co-transformed into the cells to allow rapid identification of those cells which have taken up and expressed the foreign DNA. The cells are incubated for 48-72 hours after transformation under conditions appropriate for the cell line to allow expression and accumulation of TRICH and β -galactosidase.

Transformed cells expressing \(\beta\)-galactosidase are stained blue when a suitable colorimetric substrate is added to the culture media under conditions that are well known in the art. Stained cells are tested for differences in membrane conductance by electrophysiological techniques that are well known in the art. Untransformed cells, and/or cells transformed with either vector sequences alone or \(\beta\)-galactosidase sequences alone, are used as controls and tested in parallel. Cells expressing TRICH will have higher anion or cation conductance relative to control cells. The contribution of TRICH to conductance can be confirmed by incubating the cells using antibodies specific for TRICH. The antibodies will bind to the extracellular side of TRICH, thereby blocking the pore in the ion channel, and the associated conductance.

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Alternatively, ion channel activity of TRICH is measured as current flow across a TRICH-containing *Xenopus laevis* oocyte membrane using the two-electrode voltage-clamp technique (Ishi et al., *supra*; Jegla, T. and L. Salkoff (1997) J. Neurosci. 17:32-44). TRICH is subcloned into an appropriate *Xenopus* oocyte expression vector, such as pBF, and 0.5-5 ng of mRNA is injected into mature stage IV oocytes. Injected oocytes are incubated at 18 °C for 1-5 days. Inside-out macropatches are excised into an intracellular solution containing 116 mM K-gluconate, 4 mM KCl, and 10 mM Hepes (pH 7.2). The intracellular solution is supplemented with varying concentrations of the TRICH mediator, such as cAMP, cGMP, or Ca⁺² (in the form of CaCl₂), where appropriate. Electrode resistance is set at 2-5 MΩ and electrodes are filled with the intracellular solution lacking mediator. Experiments are performed at room temperature from a holding potential of 0 mV. Voltage ramps (2.5 s) from -100 to 100 mV are acquired at a sampling frequency of 500 Hz. Current measured is proportional to the activity of TRICH in the assay.

Transport activity of TRICH is assayed by measuring uptake of labeled substrates into *Xenopus laevis* oocytes. Oocytes at stages V and VI are injected with TRICH mRNA (10 ng per oocyte) and incubated for 3 days at 18°C in OR2 medium (82.5mM NaCl, 2.5 mM KCl, 1mM CaCl₂, 1mM MgCl₂, 1mM Na₂HPO₄, 5 mM Hepes, 3.8 mM NaOH, 50µg/ml gentamycin, pH 7.8) to allow expression of TRICH. Oocytes are then transferred to standard uptake medium (100mM NaCl, 2

mM KCl, 1mM CaCl₂, 1mM MgCl₂, 10 mM Hepes/Tris pH 7.5). Uptake of various substrates (e.g., amino acids, sugars, drugs, ions, and neurotransmitters) is initiated by adding labeled substrate (e.g. radiolabeled with ³H, fluorescently labeled with rhodamine, etc.) to the oocytes. After incubating for 30 minutes, uptake is terminated by washing the oocytes three times in Na⁺-free medium, measuring the incorporated label, and comparing with controls. TRICH activity is proportional to the level of internalized labeled substrate.

ATPase activity associated with TRICH can be measured by hydrolysis of radiolabeled ATP- $[\gamma^{-32}P]$, separation of the hydrolysis products by chromatographic methods, and quantitation of the recovered ³²P using a scintillation counter. The reaction mixture contains ATP- $[\gamma^{-32}P]$ and varying amounts of TRICH in a suitable buffer incubated at 37 °C for a suitable period of time. The reaction is terminated by acid precipitation with trichloroacetic acid and then neutralized with base, and an aliquot of the reaction mixture is subjected to membrane or filter paper-based chromatography to separate the reaction products. The amount of ³²P liberated is counted in a scintillation counter. The amount of radioactivity recovered is proportional to the ATPase activity of TRICH in the assay.

Lipocalin activity of TRICH is measured by ligand fluorescence enhancement spectrofluorometry (Lin et al. (1997) Molecular Vision 3:17). Examples of ligands include retinol (Sigma, St. Louis MO) and 16-anthryloxy-palmitic acid (16-AP) (Molecular Probes Inc., Eugene OR). Ligand is dissolved in 100% ethanol and its concentration is estimated using known extinction coefficents (retinol: 46,000 A/M/cm at 325 nm; 16-AP: 8,200 A/M/cm at 361 nm). A 700 μ l aliquot of 1 μ M TRICH in 10 mM Tris (pH 7.5), 2 mM EDTA, and 500 mM NaCl is placed in a 1 cm path length quartz cuvette and 1 μ l aliquots of ligand solution are added. Fluorescence is measured 100 seconds after each addition until readings are stable. Change in fluorescence per unit change in ligand concentration is proportional to TRICH activity.

XIX. Identification of TRICH Agonists and Antagonists

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TRICH is expressed in a eukaryotic cell line such as CHO (Chinese Hamster Ovary) or HEK (Human Embryonic Kidney) 293. Ion channel activity of the transformed cells is measured in the presence and absence of candidate agonists or antagonists. Ion channel activity is assayed using patch clamp methods well known in the art or as described in Example XVIII. Alternatively, ion channel activity is assayed using fluorescent techniques that measure ion flux across the cell membrane (Velicelebi, G. et al. (1999) Meth. Enzymol. 294:20-47; West, M.R. and C.R. Molloy (1996) Anal. Biochem. 241:51-58). These assays may be adapted for high-throughput screening using microplates. Changes in internal ion concentration are measured using fluorescent dyes such as the

Ca²⁺ indicator Fluo-4 AM, sodium-sensitive dyes such as SBFI and sodium green, or the Cl⁻ indicator MQAE (all available from Molecular Probes) in combination with the FLIPR fluorimetric plate reading system (Molecular Devices). In a more generic version of this assay, changes in membrane potential caused by ionic flux across the plasma membrane are measured using oxonyl dyes such as DiBAC₄ (Molecular Probes). DiBAC₄ equilibrates between the extracellular solution and cellular sites according to the cellular membrane potential. The dye's fluorescence intensity is 20-fold greater when bound to hydrophobic intracellular sites, allowing detection of DiBAC₄ entry into the cell (Gonzalez, J.E. and P.A. Negulescu (1998) Curr. Opin. Biotechnol. 9:624-631). Candidate agonists or antagonists may be selected from known ion channel agonists or antagonists, peptide libraries, or combinatorial chemical libraries.

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Various modifications and variations of the described compositions, methods, and systems of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. It will be appreciated that the invention provides novel and useful proteins, and their encoding polynucleotides, which can be used in the drug discovery process, as well as methods for using these compositions for the detection, diagnosis, and treatment of diseases and conditions.

Although the invention has been described in connection with certain embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Nor should the description of such embodiments be considered exhaustive or limit the invention to the precise forms disclosed. Furthermore, elements from one embodiment can be readily recombined with elements from one or more other embodiments. Such combinations can form a number of embodiments within the scope of the invention. It is intended that the scope of the invention be defined by the following claims and their equivalents.

Incyte Project ID	Polypeptide	Incyte	Polynucleotide	Incyte	
	SEQ ID NO:	Polypeptide ID	SEQ ID NO:	Polynucleotide	Incyte Full Length
				m l	CIOIICS
1853191	1	1853191CD1	27	1853191CB1	
7497369	2	7497369CD1	28	7497369CB1	3508438CA2
1700438	3	1700438CD1	29	1700438CB1	1700438CA2,
					90166551CA2,
					90167174CA2,
					90167190CA2
535939	4	535939CD1	30	535939CB1	90129304CA2
55118067	5	55118067CD1	31	55118067CB1	
7502087	9	7502087CD1	32	7502087CB1	
7500819	7	7500819CD1	33	7500819CB1	
7503413	8	7503413CD1	34	7503413CB1	
7500007	6	7500007CD1	35	7500007CB1	
7500025	10	7500025CD1	36	7500025CB1	
7502736	11	7502736CD1	37	7502736CB1	
7503570	12	7503570CD1	38	7503570CB1	
7504008	13	7504008CD1	39	7504008CB1	90010166CA2,
					90010266CA2,
					90010442CA2
7503559	14	7503559CD1	40	7503559CB1	90198141CA2
6243872	15	6243872CD1	41	6243872CB1	
90011608	16	90011608CD1	42	90011608CB1	90011075CA2,
					90011083CA2,
					90011608CA2,
					90011624CA2
90024583	17	90024583CD1	43	90024583CB1	90024583CA2
90113658	18	90113658CD1	44	90113658CB1	
3942766	19	3942766CD1	45	3942766CB1	

Incyte Project ID	Polypeptide	Incyte	Polynucleotide	Incyte	
3	SEQ ID NO:	Polypeptide ID	SEQ ID NO:	Polynucleotide ID	Incyte Full Length Clones
7501987	20	7501987CD1	46	7501987CB1	5842557CA2,
				٠	90119749CA2,
					90119849CA2,
					90119857CA2,
					90120041CA2,
					90152509CA2
7503223	21	7503223CD1	47	7503223CB1	90041459CA2,
					90041551CA2
7503566	22	7503566CD1	48	7503566CB1	90066119CA2,
					90066211CA2
7505122	23	7505122CD1	49	7505122CB1	1803394CA2,
					1805561CA2
7511620	24	7511620CD1	50	7511620CB1	
7506995	25	7506995CD1	51	7506995CB1	90119757CA2
7506996	26	7506996CD1	52	7506996CB1	

Polypeptide SEQ Incyte ID NO: Polype	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	Probability Score	Annotation
1	1853191CD1	g6996442	9.0E-168	[Homo sapiens] CTL1 protein
				O'Regan, S. et al., An electric lobe suppressor for a yeast choline transport
				mutation belongs to a new family of transporter-like proteins. Proc. Natl. Acad.
				Sci. U.S.A. 97 (4), 1835-1840 (2000).
2	7497369CD1	g9663117	6.7E-44	[Homo sapiens] organic cation transporter
3	1700438CD1		4.7E-25	[Homo sapiens] SLC11A3 iron transporter
				Abboud, S. et al., A novel mammalian iron-regulated protein involved in
				intracellular iron metabolism J. Biol. Chem. 275 (26), 19906-19912 (2000).
4	535939CD1	g2621849	2.9E-47	[Methanothermobacter thermautotrophicus] Na+/H+-exchanging protein:Na+/H+
				antiporter
				Smith, D.R., et al., Complete genome sequence of Methanobacterium
				thermoautotrophicum deltaH: functional analysis and comparative genomics J.
				Bacteriol. 179 (22), 7135-7155 (1997).
5	55118067CD1	9758059	5.0E-25	[Arabidopsis thaliana] amino acid transporter-like protein
				Sato, S. et al., Structural analysis of Arabidopsis thaliana chromosome 5. IV.
				Sequence features of the regions of 1,456,315 bp covered by nineteen physically
				assigned P1 and TAC clones. DNA Res. 5 (1), 41-54 (1998).
9	7502087CD1	g2745729	0.0	[Rattus norvegicus] potassium channel
				Shi, W. et al., Identification of two nervous system-specific members of the erg
				potassium channel gene family. J. Neurosci. 17 (24), 9423-9432 (1997).
7	7500819CD1	g17391438	9.0E-31	[Homo sapiens] FXYD domain-containing ion transport regulator 6
		g15216228	2.9E-24	[Mus musculus] (AB032010) PLM-like protein
8	7503413CD1	g3065814	2.3E-160	[Rattus norvegicus] sodium-dicarboxylate cotransporter SDCT1
				Chen, X.Z. et al., Characterization of a rat Na+-dicarboxylate cotransporter. J.
				Biol. Chem. 273 (33), 20972-20981 (1998).

Polypeptide SEQ Incyte ID NO:	ptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	Probability Score	Annotation
8 cont.		338046 SLC13A2	8.6E-161	[Homo sapiens][Active transporter, secondary; Transporter] [Plasma membrane] Rena Na(+)-dicarboxylate cotransporter, reabsorbs Krebs cycle intermediates, such as succinate and citrate, from the glomerular filtrate.
6	7500007CD1	g202523	1.4 <u>E-261</u>	[Rattus norvegicus] GABA transporter Borden, L.A., et al. (1992) J. Biol. Chem. 267:21098-21104. Molecular heterogeneity of the GABA transport System: Cloning of two novel high-affinity GABA transporters from rat brain.
			2.5E-220	[Homo sapiens] Gamma-aminobutyric acid (GABA) transporter, expressed in brain, retina, and peripheral tissues, requires sodium and chloride for activity.
10	7500025CD1	g11878259 571102 KCNH2	0.0 1.3E-290	[Homo sapiens] Eag-related gene member 2 [Homo sapiens] [Channel (passive transporter); Transporter; Small molecule-binding protein][Plasma membrane] Voltage-gated (delayed rectifier) potassium channel, forms the Ikr channels that are important for cardiac rhythm; mutations in the gene are associated with the long QT form of cardiac arrhythmia.
	7502736CD1	g2745729	0:0	[Rattus norvegicus] potassium channel (Shi, W. et al. (1997) Identification of two nervous system-specific members of the erg potassium channel gene family. J. Neurosci. 17:9423-9432.)
		331276 Rn.10875	0.0	[Rattus norvegicus][Channel (passive transporter); Transporter][Plasma membrane] Ether-a-go-go related 2, a member of the erg family of potassium channels, contains six putative transmembrane domains, a slowly activating delayed rectifier potassium channel, may facilitate the differentiation of prevertebral neurons (Shi, W. et al. (1997) supra.)

Table 2

Polypeptide SEQ Incyte ID NO: Polype	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:		Annotation
11 cont.		331274 Rn.10874	5.2E-233	[Rattus norvegicus][Channel (passive transporter), Transporter] [Plasma membrane] Ether-a-go-go-related gene 3, an inward rectifier potassium channel that functions in potassium transport specifically in the nervous system (Shi, W. et al. (1997) supra.)
12	7503570CD1	g5926734 567834 SLC3A2 321548 Mdu1	9.6E-265 1.5E-259 4.0E-200	[Homo sapiens] 4F2 heavy chain [Homo sapiens] 4F2 heavy chain [Homo sapiens] 4F2 cell-surface antigen, a component of system L amino acid transport activity, involved in T-cell activation, cell proliferation and the regulation of HIV-mediated cell fusion; over-expression is associated with malignant transformation (Rajan, D. P. et al. (2000) Differential influence of the 4F2 heavy chain and the protein related to b(0,+) amino acid transport on substrate affinity of the heteromeric b(0,+) amino acid transporter. J. Biol. Chem. 275:14331-14335.) [Mus musculus][Protein kinase; Transferase; Active transporter, secondary; Transporter; Receptor (signaling)][Plasma membrane] Heavy chain of 4F2 cellsurface antigen, a component of system L amino acid transport activity, involved in cellular activation and proliferation; over-expression of human MDU1 (SLC3A2) is associated with malignant transformation (Rajan, D. P. et al. (2000) supra.)
13	7504008CD1	g13344999	0.0	[Homo sapiens] solute carrier family 26 member 6 (Waldegger, S. et al. (2001) Cloning and Characterization of SLC26A6, a Novel Member of the Solute Carrier 26 Gene Family. Genomics 72:43-50.)

Polypeptide SEQ Incyte ID NO: Polype	ptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	Probability Score	Annotation
13 cont.		662428 SLC26A6	0.0	[Homo sapiens] Member of a family of anion transporters that exchange chloride and bicarbonate ions across membranes in an electroneutral manner, expressed on apical plasma membranes of duct cells in pancreas. (Lohi, H. et al. (2000) Mapping of five new putative anion transporter genes in human and characterization of SLC26A6, A candidate gene for pancreatic anion exchanger. Genomics 70, 102-112; Waldegger, S. et al. (2001) supra.).
		336854 SLC26A4	2.8E-121	[Homo sapiens] [Transporter] Pendrin, a chloride and iodide transporter; mutation of corresponding gene causes Pendred syndrome and a common form of nonsyndromic recessive deafness. (Scott, D. A. et al. (1999) The Pendred syndrome gene encodes a chloride-iodide transport protein. Nat. Genet. 21:440-443; Usami, S. et al. (1999) Non-syndromic hearing loss associated with enlarged vestibular aqueduct is caused by PDS mutations. Hum. Genet. 104:188-192.)
		339298 SLC26A3	9.0E-112	[Homo sapiens][Transcription factor; Transporter] Solute carrier family 26 member 3, involved in anion and sulfate transport, down-regulated in colon adenomas and adenocarcinoma; gene mutation causes congenital chloride diarrhea. (Hoglund, P. et al. (1998) Clustering of private mutations in the congenital chloride diarrhea/down-regulated in adenoma gene. Hum. Mutat. 11:321-327.)
14	7503559CD1	63880560	5.6E-199	[Caenorhabditis elegans] Similarity to Yeast E1-E2 ATPase (SW:YED1_YEAST), contains similarity to Pfam domain: PF00122 (E1-E2 ATPase), Score=102.4, E-value=2.7e-28
		313777 K07E3.7	7E-204	[Caenorhabditis elegans] [Hydrolase; ATPase] Member of the P-type ATPase, Ca2+-type subfamily protein family.

14 cont.	rolypepuae 1D	TEOME	Score	
14 cont.		ED NO:		
		251962 W08D2.5	4.9E-200	[Caenorhabditis elegans] [Active transporter, primary, Hydrolase; Transporter; ATPase] [Unspecified membrane] Member of the P-type ATPase, Ca2+-type subfamily protein family.
			5.2E-192	Saccharomyces cerevisiae] [Active transporter, primary; Hydrolase; Transporter;
		YOR291W		ATPase] [Unspecified membrane] Member of the cation-translocating P-type
				ATPase superfamily of membrane transporters.
15 62	6243872CD1		2E-150	[Rattus norvegicus] testis-specific transporter TST1
			1.7E-85	Homo sapiens] [Transporter] [Unspecified membrane] Organic anion transporter
		SLC21A12		(solute carrier family 21 member 12), a member of the OATP family that
				transports thyroid hormone by a Na+independent process, may be involved in the
				transport of bile acids and prostaglandins.
				Fujiwara, K. et al. (2001) Endocrinology 142:2005-2012
				Identification of thyroid hormone transporters in humans: different molecules are
				involved in a tissue-specific manner.
		252769	1.3E-58	[Caenorhabditis elegans] [Transporter] [Unspecified membrane] Member of the
		Y70G10A.3		prostaglandin transporter protein family.
		343800	2E-52	[Homo sapiens] [Active transporter, secondary; Transporter] [Unspecified
×		SLC21A6		membrane; Plasma membrane] Organic anion transporter, has a broad substrate
				specificity and is expressed in the liver.
		618842	4.2E-52	[Mus musculus] [Transporter] Protein with high similarity to human SLC21A6,
		Slc21a14		which is an organic anion transporter with broad substrate specificity and
				expressed in the liver.
		245952	5.8E-50	[Caenorhabditis elegans] [Transporter] [Unspecified membrane] Member of the
		F53B1.8		prostaglandin transporter protein family.
16 90	90011608CD1	4	8.8E-222	[Rattus norvegicus] lysosomal amino acid transporter 1
				Sagne, C. et al. (2001) Proc. Natl. Acad. Sci. U S A 98:7206-7211
				Identification and characterization of a lysosomal transporter for small neutral
				amino acids.

Polypeptide SEQ Incyte ID NO: Polype	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	Probability Score	Annotation
16 cont.		252408 Y43F4B.7	2.2E-65	[Caenorhabditis elegans] [Active transporter, secondary; Transporter] [Unspecified membrane] Putative membrane transporter of the amino acid/auxin permease-like (AAAP-like) protein family.
		251329 T27A1.5	3E-63	[Caenorhabditis elegans] Protein with similarity to membrane transporters of the amino acid/auxin permease family, putative paralog of C. elegans Y43F4B.7.
		240958 C44B7.6	3.1E-43	[Caenorhabditis elegans] [Active transporter, secondary; Transporter] [Unspecified membrane] Member of the amino acid/auxin permease protein family.
		247128 H32K16.1	2.8E-42	[Caenorhabditis elegans] Putative paralog of C.elegans Y38H6C.17, has weak similarity to C. elegans UNC-47, vesicular GABA transporter (VGAT).
		313559 F59B2.2	4.8E-40	[Caenorhabditis elegans] Protein with strong similarity to C. elegans Y38H6C.17, has similarity to C. elegans UNC-47, vesicular GABA transporter.
17	90024583CD1	g1098557 338046 SLC13A2	4.2E-167	[Homo sapiens] renal sodium/dicarboxylate cotransporter [Homo sapiens] [Active transporter, secondary; Transporter] [Unspecified membrane; Plasma membrane] Sodium-dependent dicarboxylate cotransporter (solute carrier family 13 member 2), reabsorbs Krebs cycle intermediates, such as succinate and citrate, from the renal tubular filtrate. Pajor, A.M. (1996) Am. J. Physiol. 270:F642-648 Molecular cloning and functional expression of a sodium-dicarboxylate cotransporter from human kidney.
		331190 Rn.10821	1E-165	[Rattus norvegicus] [Active transporter, secondary; Transporter] [Unspecified membrane] Sodium-dependent dicarboxylate cotransporter (solute carrier family 13 member 2), reabsorbs Krebs cycle intermediates, such as succinate and citrate, from the renal tubular filtrate.

Polypeptide SEQ Incyte ID NO:	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:		Annotation
17 cont.		742868 SLC13A3	1.3E-142	[Homo sapiens] Na(+)-dependent high-affinity dicarboxylate transporter 3, mediates succinate transport in the presence of sodium.
			9.2E-140	[Rattus norvegicus] [Active transporter, secondary; Transporter] [Unspecified membrane] Sodium-dependent high-affinity dicarboxylate transporter 3, may function in di- andtri-carboxylic acid transport and elimination of organic anions and therapeutic drugs in the kidney.
		742872 SLC13A4	2.3E-132	[Homo sapiens] Sulfate transporter, mediates sodium-dependent sulfate transport at high levels, may function in sulfate incorporation in high endothelial venules.
18	90113658CD1	g6010763	8.1E-180	[Rattus norvegicus] ion transporter protein
		430266 Slc22a3	5.8E-29	[Mus musculus] [Active transporter, secondary; Major Facilitator Superfamily; Transporter] [Unspecified membrane; Plasma membrane] Solute carrier family 22 member 3 (extraneuronal monoamine transporter), regulates monoamine transport in the heart and placenta. Grundemann, D. et al. (1999) Selective substrates for non-neuronal monoamine transporters. Mol. Pharmacol. 56:1-10.
19	3942766CD1	g8132324	0.0	[Homo sapiens] sodium-dependent high-affinity dicarboxylate transporter. Wang, H. et al. (2000) Structure, function, and genomic organization of human Na+-dependent high-affinity dicarboxylate transporter. Am. J. Physiol. 278:C1019-C1030.
		742868 SLC13A3	0.0	[Homo sapiens] Na(+)-dependent high-affinity dicarboxylate transporter 3, mediates succinate transport in the presence of sodium.
		661462 Nadc3	2.6E-286	[Rattus norvegicus] [Active transporter, secondary; Transporter] [Unspecified membrane] Sodium-dependent high-affinity dicarboxylate transporter 3, may function in di- and tri-carboxylic acid transport and elimination of organic anions and therapeutic drugs in the kidney. (Wang, H. et al. (2000) supra.)

Polypeptide SEQ Incyte ID NO: Polypeptide ID	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score	Probability Score	Annotation
		D NO:		
19 cont.		331190	3.2E-137	[Rattus norvegicus] [Active transporter, secondary; Transporter] [Unspecified
		Rn.10821		membrane] Sodium-dependent dicarboxylate co-transporter (solute carrier family
				13 member 2), reabsorbs Krebs cycle intermediates, such as succinate and citrate,
				from the renal tubular filtrate.
		338046	8.8E-135	[Homo sapiens] [Active transporter, secondary; Transporter] [Unspecified
		SLC13A2		membrane; Plasma membrane] Sodium-dependent dicarboxylate cotransporter
				(solute carrier family 13 member 2), reabsorbs Krebs cycle intermediates, such as
				succinate and citrate, from the renal tubular filtrate.
		742872	1.7E-121	[Homo sapiens] Sulfate transporter, mediates sodium-dependent sulfate transport
		SLC13A4		at high levels, may function insulfate incorporation in high endothelial venules.
20	7501987CD1	g20384654	1.0E-151	[Homo sapiens] two-pore calcium channel protein 2
		g14041819	8.5E-17	[Arabidopsis thaliana] (AB053952) two-pore calcium channel.
)		Furuichi, T. et al. (2001) A Putative Two Pore Channel AtTPC1 Mediates Ca(2+)
				Flux in Arabidopsis Leaf Cells. Plant Cell Physiol. 42: 900-905.
		367634	2.1E-16	[Rattus norvegicus] Member of the ion transport family, which contains calcium,
		Rn.24484		sodium, and potassium channels, has weak similarity to a region of the alpha-1
				subunit of voltage-gated calcium channels.
21	7503223CD1	g2739503	6.8E-80	[Homo sapiens] potassium channel
		g2739503	6.8E-80	[Homo sapiens] potassium channel.
				Su, K. et al. (1997) Isolation, characterization, and mapping of two human
				potassium
				Biochem. Biophys. Res. Commun. 241: 675-681.

Annotation	[Homo sapiens] [Channel (passive transporter); Transporter] [Plasma membrane; Unspecified membrane] Potassium voltage channel subfamily G 1, functions as a regulatory subunit, slows deactivation of the potassium channel upon association with Kv2.1 (KCNB1), may play a role in regulating membrane excitability. Su, K. et al. (1997) Isolation, characterization, and mapping of two human potassium channels. Biochem. Biophys. Res. Commun. 241: 675-81.	[Homo sapiens] [Channel (passive transporter); Transporter] [Plasma membrane] Potassium voltage channel subfamily gamma 2, a member of the Kv6 family of ion channels, functions as a votage-gated potassium channel upon interaction with Kv2.1 alpha subunit, may contribute to cardiac action potentiation repolarization.	Rattus norvegicus] [Channel (passive transporter); Transporter] [Plasma membrane; Unspecified membrane] Potassium voltage-gated channel (Shabrelated subfamily, member 1), a delayed rectifier potassium channel which plays a role in vasoconstriction, hypertension, and probably regulates neuronal excitability; associated with type I diabetic cardiomyopathy.	[Mus musculus] [Channel (passive transporter); Transporter] [Plasma membrane; Unspecified membrane] Potassium voltage-gated channel (Shab-related subfamily, member 1), a delayed rectifier channel which plays a role in Schwann cell proliferation, glucose-stimulated insulin secretion, and probably regulates neuronal excitability.	[Homo sapiens] [Channel (passive transporter); Transporter] [Plasma membrane; Unspecified membrane] Potassium voltage-gated channel (Shab-related subfamily, member 1), functions as a delayed rectifier potassium channel.
Probability Score	6.0E-81	2.4E-24	1.2E-23	1.3E-23	1.3E-23
GenBank ID NO: Probability or PROTEOME Score ID NO:	336122 KCNG1	428816 KCNG2	430588 Kcnb1	429486 Kcnb1	341660 KCNB1
Incyte Polypeptide ID					
Polypeptide SEQ Incyte ID NO: Polype	21 cont.				

OTO -1:1- C	1	Canbant ID MO. Brokehility	Γ	Amodation
Folypeptide SEQ incyte ID NO: Polype	mcyte Polypeptide ID	or PROTEOME S		
22	7503566CD1	g1314290	2.0E-169	[Homo sapiens] vesicular monoamine transporter VMAT1. Erickson, J.D. et al.(1996) Distinct pharmacological properties and distribution in neurons and endocrine cells of two isoforms of the human vesicular monoamine transporter. Proc. Natl. Acad. Sci. U.S.A. 93: 5166-5171.
		338052 SLC18A1	1.7E-170	[Homo sapiens] Solute carrier family 18 member 1(vesicular monoamine transporter 1), transports and stores monoamine neurotransmitters in adrenal chromaffin cells, and is inhibited by the drugs fenfluramine, reserpine, and amphetamines.
		329252 Slc18a2	3.1E-165	[Rattus norvegicus] [Transporter] [Unspecified membrane; Other vesicles of the secretory/endocytic pathways] Solute carrier family18 A2 (vesicular monoamine transporter 2), pumps monoamines and neurotrasmitters into synaptic vesicles; human SLC18A2 genetic variants may be associated with neurobehavioral disorders.
		338054 SLC18A2	8.3E-165	[Homo sapiens] [Transporter] [Secretory vesicles; Cytoplasmic; Unspecified membrane; Plasma membrane; Otherv esicles of the secretory/endocytic pathways] Solute carrier family18 A2 (vesicular monoamine transporter 2), pumps monoamines and neurotransmitters into synaptic vesicles, activity is sensitive to inhibition by tetrabenazine; genetic variants may be associated with neurobehavioral disorders.
		331476 Slc18a1	1.2E-146	[Rattus norvegicus] [Transporter] [Secretory vesicles; Cytoplasmic; Unspecified membrane; Other vesicles of the secretory/endocytic pathways] Solute carrier family 18 member 1 (vesicular monoamine transporter 1), transports and stores monoamine neurotransmitters in adrenal chromaffin cells, and is inhibited by the drug reserpine.

	in enopus	nnsporter] i. channel in breast varies in oma cell east	embrane] nannel or ımary and	is an renal ression is	nember of
	nan-like prote fuctance in X	nsporter); Tr nt regulator (3) y, expressed te regulation nmary carcir gression of b	r] [Plasma n ve chloride c initiated mar ifferentiation	e transporter) actor, contain K-ATPase ir neostasis; ex	sa6, a novel r
	hospholemm chloride con	I (passive tra g ion transponannel activit gression; ger is of two man	r);Transporte ator 3, putati nue and ras- in cellular di	unel (passive el-inducing f egulates Na, r and K+ hor	ation of slc26
	t-8, a novel p s, induces a 76-2182.	unit; Channe ain containin se chloride chest cancer procular analys as a model sy (129-39.	ve transporte ansport regul pecifically in a is involved	subunit; Charanarane] Chann , binds and r nance of Na-	mily 26 men 1 characteriz
	T8 protein. I. (1995) Mabreast tumor breast tumor m. 270 : 217	gulatory sub FXYD dom that mediate arker of brea arker of thea les. (1998) Mole tional level a fetastasis 16	tannel (passi aining ion tra pregulated sl at the protein	[Regulatory cified memb XYD domain le in mainte one.	te carrier fa Cloning and
tion	[Homo sapiens] MAT8 protein. Morrison, B.W. et al. (1995) Mat-8, a novel phospholemman-like protein expressed in human breast tumors, induces a chloride conductance in Xenopus oocytes. J. Biol. Chem. 270: 2176-2182.	[Homo sapiens] [Regulatory subunit; Channel (passive transporter); Transporter] [Plasma membrane] FXYD domain containing ion transport regulator 3, channel or channel regulator that mediates chloride channel activity, expressed in breast cancers, potential marker of breast cancer progression; gene regulation varies in breast cancer cell lines. Schiemann, S. et al. (1998) Molecular analysis of two mammary carcinoma cell lines at the transcriptional level as a model system for progression of breast cancer. Clin. Exp. Metastasis 16:129-39.	[Mus musculus] [Channel (passive transporter); Transporter] [Plasma membrane] FXYD domain containing ion transport regulator 3, putative chloride channel or channel regulator; upregulated specifically in nue and ras-initiated mammary tumors indicating that the protein is involved in cellular differentiation and signaling.	[Rattus norvegicus] [Regulatory subunit; Channel-inducing factor, contains an Transporter] [Unspecified membrane] Channel-inducing factor, contains an extracytoplasmic FXYD domain, binds and regulates Na, K-ATPase in renal tissue, may play a role in maintenance of Na+ and K+ homeostasis; expression is induced by aldosterone.	[Homo sapiens] solute carrier family 26 member 6 Waldegger, S. et al., Cloning and characterization of slc26a6, a novel member of the solute carrier 26 oene family. Genomics 72, 43-50, (2001).
Annotation	[Homo Morrisc express oocytes	[Homo Plasma or chan cancers breast c Schiem lines at cancer.	[Mus mus FXYD do channel re tumors in signaling.	[Rattus Transpo extracy tissue, 1	[Homo Waldeg
Probability Score	7.0E-23	6.1E-24	8.5E-18	1.2E-13	4.0E-76
GenBank ID NO: Probability or PROTEOME Score ID NO:	g1085026	343156 FXYD3	319786 Fxyd3	628631 LOC64190	g13344999
ptide ID	7505122CD1 g	<u>ж</u>	Е Н	7	7511620CD1 g
Polypeptide SEQ Incyte ID NO: Polype	23				24

Polypeptide SEQ Incyte D NO: Polype	ptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	Probability Score	Annotation
24 cont.		662428 SLC26A6	1.6E-66	[Homo sapiens] Solute carrier family 26 member 6, a member of the anion transporter family that is present on the apical plasma membrane and may act in exchange of chloride and bicarbonate ions across membranes in an electroneutral manner.
				Lohi, H. et al., Mapping of five new putative anion transporter genes in human and characterization of SLC26A6, A candidate gene for pancreatic anion exchanger, Genomics 70, 102-12 (2000).
		430122 S1c26a4	1.8E-23	[Mus musculus][Transporter][Plasma membrane; Unspecified membrane] Solute carrier family 26 member 4 (pendrin), a putative bicarbonate transporter; mutation of the human SLC26A4 gene causes Pendred syndrome, enlarged vestibular aqueduct syndrome, and DFNB4 form of non syndromic hearing loss.
				Royaux, I. E. et al., Pendrin, encoded by the Pendred syndrome gene, resides in the apical region of renal intercalated cells and mediates bicarbonate secretion., Proc Natl Acad Sci U S A 98, 4221-6. (2001).
25	7506995CD1	g20384654 o4586963	1.0E-129 8.6E-27	[Homo sapiens] two-pore calcium channel protein 2 Rattus norvegicus] voltage-gated ca channel
		20,000		Ishibashi, K. et al. Molecular cloning of a novel form (Two-repeat) protein related to voltage-gated sodium and calcium channels
				Biochem. Biophys. Res. Commun. 270, 370-376 (2000)
		367634 Rn.24484	7.4E-28	[Rattus norvegicus] Member of the ion transport family, which contains calcium, sodium, and potassium channels, has weak similarity to a region of low voltage-activated T-type calcium channel alpha 1I subunit (rat Cacna1i).
				Ishibashi, K. et al. (supra)

									
Annotation	[Homo sapiens][Channel (passive transporter); Transporter][Plasma membrane; Unspecified membrane] Calcium channel (voltage-dependent, T-type) alpha 1G subunit, generates T-type calcium currents, mediates increases in intracellular calcium concentrations; may play a role in tumor development.	Perez-Reyes, E. et al. Molecular characterization of a neuronal low-voltage-activated T-type calcium channel. Nature 391, 896-900 (1998).	Toyota, M. et al. Inactivation of CACNA1G, a T-type calcium channel gene, by aberrant methylation of its 5' CpG island in human tumors. Cancer Res 59, 4535-	41. (1999).	[Homo sapiens] two-pore calcium channel protein 2	[Arabidopsis thaliana] putative calcium channel	[Rattus norvegicus] Member of the ion transport family, which contains calcium, sodium, and potassium channels, has weak similarity to a region of low voltage-	activated T-type calcium channel alpha 11 subunit (rat Cacna1i) Ishibashi, K. et al. Biochem Biophys Res Commun 270, 370-6 (2000).	
Probability Score	7.0E-11		:		0.0	5.2E-31	7.4E-32		
GenBank ID NO: Probability or PROTEOME Score ID NO:	605816 CACNA1G				g20384654	g7270679	367634 Rn.24484		
Incyte Polypeptide ID					7506996CD1				
Polypeptide SEQ Incyte ID NO: Polypeptide ID	25 cont.				26				

PCT/US02/26323

SEQ	Incyte	Amino Acid Potential		Potential	Signature Sequences, Domains and Motifs	Analytical Methods
А	ID Polypeptide	Residues	Phosphorylation	Glycosylation Sites		and Databases
Ö.	П		Sites			
	1853191CD1	652	S77 S85 S127 S155 N136 N151 N412	N136 N151 N412	Oytosolic domain: M1-T32, T237-L240, R306-S325, TMHMMER	TMHMMER
			S210 S414 S509	N502 N520	T408-D532, S583-R652	
			S583 S607 S624		Transmeinbrane domain: A33-A55, T214-I236,	
			S638 T30 T90		V241-Y263, C283-L305, A326-V348, I385-V407,	
			T232 T276 T319		F533-N555, F560-L582	
			T587		Non-cytosolic domain: G56-D213, Y264-K282,	
					A349-L384, Y556-A559	
2	7497369CD1	345	S89 S104 S144	NI3	M1-G128, D179-R184, A235-	TMHMMER
			S305 T124		P240, E296-H345	
					Non-cytosolic domain:R147-T155, L208-W211,	·
					L264-G272	
					Transmembrane domain: L129-R146, F156-A178,	
					P185-Y207, T212-F234, T241-P263, F273-P295	
3	1700438CD1	150	S125		Signal Peptide: M19-G36; M19-G38; M7-G36	HMMER
					Cytosolic domains: M1-M19, R77-L82	TMHMMER
					Transmembrane domains: V20-T42, I57-L76, V83-	
					L105	
					Non-cytosolic domains: G43-S56, A106-G150	
					NADH-UBIQUINONE/PLASTOQUINONE	BLIMPS_PFAM
					OXIDOREDUCTASE CHAIN 6 PF00499: W12-Y32	
4	535939CD1	537	S55 S175 S276	06N	Sodium/hydrogen exchanger family: 1119-V520	HIMMER_PFAM

Analytical Methods	and Databases	TMHMMER					HMMER_PFAM		TMHMMER							BLAST_DOMO		HIMMER_PFAM	HIMMER PFAM	HMMER_PFAM	HIMMER PFAM	TMHIMMER				
Signature Sequences, Domains and Motifs		Cytosolic domains: S107-G112, I162-W172, P230-W233, S329-T340, S412-T417, G513-V537	Transmembrane domains: V87-G106, G113-I135,	G306-P328, F341-A374, I389-A411, V418-V440,	V490-L512	Non-cytosolic domains: M1-R86, K136-P138, D193-R206, Q257-R305, G375-I388, C441-D489	Transmembrane amino acid transporter protein: C30-HMMER_PFAM	S398	Cytosolic domains: M1-N8, K59-K82, Q143-A148,	T253-K322, K367-Q377	Transmembrane domains: W9-F31, I36-V58, M83-	G105, V120-L142, S149-S171, F230-F252, A323-	P340, T344-Y366, V378-V397	Non-cytosolic domains: K32-G35, D106-Q119, S172-	1229, N341-E343, S398-S1119	NEUROFILAMENT; TRIPLET;	DM04498 P12036 434-1019: V389-E877	PAC motif: K93-L135	PAS domain: I41-M60			Cytosolic domains: D284-A399, T486-V491			Non-cytosolic domains: M1-D260, G423-K462, I515-	H947
Potential	Glycosylation Sites			-														N218 N449 N510	N731							
Potential	Phosphorylation Sites	S380 S412 S477 T9 T62 T105 T474					S64 S172 S187	S217 S222 S251	S396 S429 S545	S685 S965 T68	T123 T847							S26 S55 S105 S140	S145 S200 S283	S288 S458 S488	S670 S706 S740	S763 S777 S853	S861 S868 S886	S918 T13 T170	T202 T220 T301	T326 T363 T377
Amino Acid Potential	Residues						1119											947						1275		
Incyte							55118067CD1 1119											7502087CD1								
SEQ	의 ²	4 cont.					5											9			-					

SEQ	Incyte	Amino Acid Potential	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
요 일 유	Polypeptide ID	Residues	Phosphorylation Sites	Glycosylation Sites		and Databases
6 cont.			T486 T522 T678		POTASSIUM CHANNEL IONIC CHANNEL PD118772: E702-S944	BLAST_PRODOM
	,				CHANNEL IONIC POTASSIUM K+ SUBUNIT HYPERPOLARIZATION-ACTIVATED PROTEIN	BLAST_PRODOM
					PUTATIVE EAG LONG PD001039: S228-V339	-
	,				CHANNEL PROTEIN IONIC POTASSIUM	BLAST_PRODOM
					NONPHOTOTROPIC HYPOCOTYL PUTATIVE	
					SUBUNIT REPEAT EAG PD009483; MI-L86	
					CHANNEL IONIC POTASSIUM SUBUNIT	BLAST_PRODOM
					MERG1A RERG VENTRICULAR ERG K+	
					PUTATIVE PD009699: K174-L227	
					CAMP RECEPTOR PROTEIN CYCLIC	BLAST_DOMO
					NUCLEOTIDE-BINDING DOMAIN	
					DM01165 I38465 562-948; H413-F727	
					DM01165 I48912 391-786: H413-R715	
					DM01165 Q02280 384-776: H413-R715	
					POTASSIUM; CHANNEL; KST1; AKT1;	BLAST_DOMO
					DM02383 I38465 353-560: T201-A412	
7	7500819CD1	80	S19 S58 T36 Y32		signal_cleavage: M1-A18	SPSCAN
					Signal Peptides: M1-A18, M1-A20, M1-A21, M1-	HMMER
					K25, M1-K23	
					ATP1G1/PLM/MAT8 family: A21-C75	HMMER PFAM
					Cytosolic domains: M1-E2, R59-Q80	TMHMMER
					Transmembrane domains: L3-A21, T36-S58	
					Non-cytosolic domain: E22-Q35	
_					ATP1G1 / PLM / MAT8 family proteins BL01310:	BLIMPS_BLOCKS
					D28-C63	

Analytical Methods	ally Dalabases	BLAST_PRODOM	BLAST_PRODOM	BLAST_DOMO	MOTIFS	HMMER	HMMER_PFAM	TMHMMER						
Signature Sequences, Domains and Motifs		PHOSPHOLEMMAN PRECURSOR TRANSMEMBRANE PHOSPHORYLATION SIGNAL IONIC CHANNEL ION TRANSPORT PD058885; L5-F65	TRANSMEMBRANE TRANSPORT PRECURSOR SIGNAL CHANNEL ION SODIUM/POTASSIUM-TRANSPORTING ATPASE GAMMA CHAIN PD005989: D28-P69	CAMP; MAJOR; SODIUM; TRANSMEMBRANE; DM02637 A40533 19-92: A21-F65	ATP1G1 / PLM / MAT8 family signature: D28-G41 MOTIFS	Signal Peptide: M1-A33, M1-C38	Sodium: sulfate symporter transmembrane: S6-W341, P356-L517	Cytosolic domains: M1-K12, P68-T86, N141-L210,	Y276-K308, T423-N428, G481-T491 Transmembrane domains:	\$13-L30, M45-F67, N87-H106, P121-S140,	L403-F422, V429-L448, T458-Y480,	G492-I514	Non-cytosolic domains:	M31-L44, K107-K120, L234-S252, S332-P402, N449-C457, F515-T531
Potential	Glycosylation Sites					N525								
Potential	Phosphorylation Sites					S291 S313 T527				-				
Amino Acid Potential	Residues					531								
SEQ Incyte	Polypeptide ID					7503413CD1								
SEQ	a ë	7 cont.				∞								

SEO	Incyte	Amino Acid Potential	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
A	Polypeptide	Residues	Phosphorylation	Glycosylation Sites		and Databases
NO:	А		Sites			
∞					Sodium: sulfate symporter family proteins	BLIMPS_BLOCKS
cont.					BL01271: T132-I151, M208-V232, P370-G391,	(P< 1.3E-10)
					S443-I497	
			ŷ.		PROTEIN TRANSMEMBRANE TRANSPORT	BLAST_PRODOM
				,	MEMBRANE INNER TRANSPORTER SODIUM	
					SYMPORT OF COTRANSPORTER	
			•		PD000549: M208-W510, V15-V173	
					RENAL; BOUND; PRO-SER-ALA; NA;	BLAST_DOMO
					DM02914	
					P46556 1-520: R37-W341, L320-F515	
					A47714 28-576; P358-F515, T144-W341,	
					I29-K206	•
					S43561 28-507: R37-M159, L359-T509,	
					V152-W341, A4-V41	
4.05					P32739 25-517: G336-F515, M125-W341,	
					R37-M159, P21-T86	
					Na Sulfate symporter: A461-V477	MOTIFS
6	7500007CD1 510	510	S10 T138 T433	N77 N81 N86	Sodium: neurotransmitter symporter family: E68-	HIMMER_PFAM
			T463 T495 T504	N177	P478, R32-G67	

Analytical Methods	and Databases	TMHMMER	BLIMPS_BLOCKS	PROFILESCAN	BLIMPS_PRINTS	BLAST_PRODOM	BLAST_PRODOM
Signature Sequences, Domains and Motifs	a	Cytosolic domains: K133-K140, K213-A224, D297-E325, A381-P398, L464-C510 Transmembrane domains: G110-W132, V141-L163, A190-N212, L225-M247, V274-L296, V326-M348, A358-G380, L399-I421, L441-T463 Non-cytosolic domains: M1-L109, P164-D189, S248-A273, Y349-Y357, K422-A440	Sodium: neurotransmitter symporter family proteins BL00610: W114-G165, R180-T232, A273-P315, S372-L426, Y432-P454	Sodium: neurotransmitter symporter family rignatures:N36-F129	SODIUM/NEUROTRANSMITTER PR00176: V141-I158, I223-I243, L277-L296, G361-A381, E40-L61, K401-I421	TRANSPORTER NEUROTRANSMITTER TRANSPORT TRANSMEMBRANE SYMPORT GLYCOPROTEIN SODIUM CHLORIDE- DEPENDENT SODIUM DEPENDENT GABA PD000448: G134-P478, N54-Y150, R32-G67	TRANSMEMBRANE TRANSPORT PROTEIN TRANSPORTER AMINO ACID PERMEASE AMINO ACID GLYCOPROTEIN MEMBRANE PD000214: A166-P454, A111-S261
Potential S	Glycosylation Sites	O H Z	N	<u> </u>	S	1 0 0	T T
Potential	Phosphorylation Sites						
Amino Acid Potential	Residues						
Incyte	Polypeptide ID						
SEQ	A 8	cont.					

SE	SEO Incyte	Amino Acid Potential	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
A S	Polypeptide	Residues	Phosphorylation Sites	Glycosylation Sites		and Databases
6	1		20170		SODIUM CHLORIDE-DEPENDENT GABA	BLAST PRODOM
cont.				1	TRANSPORTER NEUROTRANSMITTER	
					TRANSPORT TRANSMEMBRANE	
					GLYCOPROTEIN SYMPORT MULTIGENE	
					PD037836: T463-C510	
					SODIUM: NEUROTRANSMITTER SYMPORTER BLAST_DOMO	BLAST_DOMO
					FAMILY	
					DM00572	•
					P31646 22-576:E68-R485, E22-G67	
					P31649 22-576:E68-R485, E22-G67	
					P27799 26-581:E68-Q484, E22-G67	
					P48066 40-596:T8-R485, K24-G67	
10	7500025CD1	894	S2 S26 S55 S105	N218 N457 N678	PAC motif: K93-L135	HMMER_PFAM
			S140 S145 S200			
			S283 S288 S435			
			S617 S653 S687			
			S710 S724 S800			
	-		S808 S815 S833			
			S865 T13 T170			
			T202 T220 T301			
			T326 T363 T377			
			T433 T469 T625			
					Cyclic nucleotide-binding domain: K556-A646	HMMER PFAM
					Ion transport protein: P299-V458	HIMMER PFAM

Analytical Methods and Databases	TMHMMER	BLAST_PRODOM	BLAST_PRODOM	BLAST_PRODOM	BLAST_PRODOM	BLAST_DOMO
Signature Sequences, Domains and Motifs	Cytosolic domains: M1-A257, R320-A338, C419-V438 Transmembrane domains: V258-F280, L300-F319, V339-F361, Y396-I418, F439-I461 Non-cytosolic domains: L281-P299, R362-E395, I462-H894	POTASSIUM CHANNEL IONIC CHANNEL PD118772: E649-S891	CHANNEL IONIC POTASSIUM K+ SUBUNIT HYPERPOLARIZATION ACTIVATED PROTEIN PUTATIVE EAG LONG PD001039: S228-V339	CHANNEL, PROTEIN IONIC POTASSIUM NON- PHOTOTROPIC HYPOCOTYL PUTATIVE SUBUNIT REPEAT EAG · PD009483: R4-L86	CHANNEL IONIC POTASSIUM SUBUNIT MERG1A RERG VENTRICULAR ERG K+ PUTATIVE PD009699; K174-L227	CAMP RECEPTOR PROTEIN CYCLIC NUCLEOTIDE-BINDING DOMAIN DM01165 I38465 562-948: S420-F674, H413-I418 Q02280 384-776: A412-R662, H413-I418 I48912 391-786: S420-R662, H413-I418
Potential Glycosylation Sites						
Potential Phosphorylation Sites						
Amino Acid Potential Residues Phosphor Sites						
Incyte Polypeptide ID						·
اند ا	cont.					

PCT/US02/26323

Incyte	Amino Acid Potential	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
_	Residues	Phosphorylation Sites	Glycosylation Sites		and Databases
				POTASSIUM; CHANNEL; KST1; AKT1; DM02383 138465 353-560; T201-A412	BLAST_DOMO
	788	S19 S24 S47 S60	N290 N351 N572	Cyclic nucleotide-binding domain: K450-A540	HMMER_PFAM
	3	S64 S79 S85 S124		Ion transport protein: P140-V352	HMMER PFAM
		S129 S299 S329		Cytosolic domains: M1-D101, N159-A240, T327-	TMHMMER
		S511 S547 S581		V332	
		S604 S618 S694		Transmembrane domains: W102-S124, S139-I158,	
		S702 S709 S727		V241-I263, Y304-N326, F333-I355	
		S759 T33 T142		Non-cytosolic domains: D125-C138, G264-K303,	
		T167 T204 T218		I356-H788	
		T327 T363 T519		POTASSIUM CHANNEL IONIC CHANNEL	BLAST_PRODOM
				PD118772: E543-S785	
				CHANNEL IONIC POTASSIUM K+ SUBUNIT	BLAST_PRODOM
				HYPERPOLARIZATIONACTIVATED PROTEIN	
				PUTATIVE EAG LONG PD001039: A94-V180	÷
				CAMP RECEPTOR PROTEIN CYCLIC	BLAST_DOMO
				NUCLEOTIDE-BINDING DOMAIN	
				DM01165 138465 562-948: H254-F568	
				DM01165 148912 391-786: H254-R556	
				DM01165 Q02280 384-776; H254-R556	
				POTASSIUM; CHANNEL; KST1; AKT1;	BLAST_DOMO
				DM02383 138465 353-560: A94-A253	
7503570CD1	501	S64 S152 S185	N264 N280 N323	Cytosolic domain: M1-R81	TMHMMER
		S195 S200 S289		Transmembrane domain: W82-V104	
		S293 S309 S366		Non-cytosolic domain: R105-A501	

Analytical Methods	and Databases	BLAST_PRODOM	BLAST_DOMO	BLAST_DOMO	HMMER_PFAM	HIMMER PFAM	TIMHIMMER					BI IMPS BLOCKS		BLAST_PRODOM		
Signature Sequences, Domains and Motifs		HEAVY CHAIN CD98 PROTEIN 4F2 CELL SURFACE ANTIGEN 4F2HC GLYCOPROTEIN TRANSMEMBRANE PD013690: Q221-P387, L373-A501	CELL SURFACE ANTIGEN 4F2 HEAVY CHAIN DM08844 P08195 113-428: P113-Q381, S366-R401 DM08844 P10852 107-423: P113-Q381, Q381-R401	1LAT; CYCLOMALTODEXTRINASE; DM04732 P08195 430-528: S402-A501 DM04732 S57719 424-527: R401-Y499	STAS domain: Y493-A700	Sulfate transporter family: L198-T470	Cytosolic domains: T138-T186, R331-E341, F399-	K404, R469-L721	Transmembrane domains: A115-G137, L187-Y209, G308-L330, L342-M364, V379-L398, A405-L422,	L437-V468	Non-cytosolic domains: M1-L114, L210-V307, S365-	Q378, S423-L436	Sulfate transporters proteins BL01130: S96-V149, T186-L237	SULFATE TRANSPORTER TRANSPORT	PROTEIN TRANSMEMBRANE GLYCOPROTEIN	AFFINITY SULPHATE HIGH PERMEASE PD001121: L71-V196
Potential	Glycosylation Sites				N167 N172 N576											
Potential	Phosphorylation Sites	S398 S478 T5 T220			S156 S338 S536	8565 8578 S596	S641 S650 S676	S696 T138 T174	T251 T577 T619	1007 1000						
Amino Acid Potential	Residues				721	1										
Incyte	Polypeptide	<u>a</u>			7504008CD1								_			
OH.	A S	cont.			13	7										

Analytical Methods	and Databases	BLAST_PRODOM	BLAST_PRODOM	BLAST_DOMO	HMMER_PFAM
Signature Sequences. Domains and Motifs		PROTEIN TRANSPORT SULFATE TRANSPORTER TRANSMEMBRANE PERMEASE INTERGENIC REGION AFFINITY GLYCOPROTEIN PD001255: H278-R469, L198-L237	SULFATE TRANSPORTER PROTEIN TRANSPORT TRANSMEMBRANE AFFINITY GLYCOPROTEIN SULPHATE HIGH DISEASE PD001755: P473-L523, O627-R709	SULFATE TRANSPORTERS DM01229P45380 10-468: L264-K430, C62-A175, E159-L264, L422-L478 DM01229P50443 49-505: L264-W429, Q61-L187, S160-L264 DM01229P40879 5-462: Y29-G317, A267-K430 DM01229 Q02920 1-447: R66-W257, P260-K430	E1-E2 ATPase: N278-V365
Potential	Glycosylation Sites				N23 N150 N300 N312 N318 N704 N1045 N1053 N1059 N1073 N1217
	ylation	·			S98 S103 S130 S144 S170 S227 S252 S523 S802 S817 S899 S901 S1055 T74 T269 T353 T358 T387 T502 T549 T576 T912 T1061 T1159 T1182 T1206 Y349 Y407
Amino Acid Potential	Residues				1226
SEO Incute	ptide				7503559CD1
GEO	A S	cont.			14

Analytical Methods	and Databases	TMHMMER							BLIMPS_BLOCKS			PROFILESCAN	BLIMPS_PRINTS			BLAST_PRODOM						BLAST_PRODOM				
Signature Sequences, Domains and Motifs		Cytosolic domains: M1-K28, R254-Y411, A469-A30, N986-A1005, K1093-N1104, E1161-T1226	Transmembrane domains: L29-P51, Y231-I253,	L412-L434, 1449-Y468, A931-V953, F968-L985,	L1006-V1028, T1075-S1092, Y1105-A1127, W1142-	V1160	Non-cytosolic domains: E52-E230, N435-D448,	T954-Q967, K1029-T1074, S1128-Q1141	E1-E2 ATPases phosphorylation site proteins	BL00154: V454-G490, L492-L510, K652-C662,	N724-M764, V878-S901, A905-V938	E1-E2 ATPases phosphorylation site: 1478-E526	P-type cation-transporting atpase superfamily	signature PR00119: N318-T332, C496-L510, A740-	D750, C881-L900	ATPASE PROBABLE CALCIUM-	TRANSPORTING PROTEIN HYDROLASE	CALCIUM TRANSPORT TRANSMEMBRANE	PHOSPHORYLATION MAGNESIUM PD090368:	D1064-L1114, Q995-Y1034; PD150086: G178-	L236; PD023991: E904-I1016, E1062-D1165	ATPASE HYDROLASE TRANSMEMBRANE	PHOSPHORYLATION ATP-BINDING	TRANSPORT PUMP CALCIUM MAGNESIUM	MEMBRANE PD000132: E282-D506, Q672-I765,	D715-M764
Potential	Glycosylation Sites																								_	
Potential	Phosphorylation Sites																									
Amino Acid Potential	Residues																									
SEO Incyte	Polypeptide ID	-																								
SEO	A È	14	-																							(

Analytical Methods and Databases	BLAST_DOMO	MOTIFS	HMMER_PFAM	TMHMMER BLAST_PRODOM	
Signature Sequences, Domains and Motifs	E1-E2 ATPASES PHOSPHORYLATION SITE DM00115 P22189 49-801: V623-P767, H800-S984, S202-K331, P401-E505, S556-A575 DM00115 P13586 78-773: S202-N527, E840-D972, Q607-H790, G557-H583 DM00115 P54678 80-795: I622-I765, A235-D506, M823-Y957, V1132-D1165 DM00115 P47317 26-695: F628-K780, I238-E566, D844-L963	E1-E2 ATPases phosphorylation site: D498-T504	Organic Anion Transporter Polypeptide (OATP): G156-I394, N465-L638	Cytosolic domains: M1-C107, D176-Y187, N258-V341, R397-P498, Y610-L638 Transmembrane domains: F108-I130, I153-I175, L188-V210, W235-P257, A342-I364, A374-V396, L499-A521, L587-I609 Non-cytosolic domains: G131-D152, K211-T234, V365-K373, M522-F586 TRANSPORTER PROTEIN TRANSMEMBRANE TRANSPORT SIMILAR MATRIN F/G ORGANIC ANION SODIUMINDEPENDENT PD005488:	C423-T618
Potential Glycosylation Sites			11 S41 S75 S129 N215 N221 N418 363 S366 S371 N465 N580		
Potential Phosphorylation Sites			S11 S41 S75 S129 S263 S366 S371	·	
Amino Acid Potential Residues Phospho Sites				·	
SEQ Incyte D Polypeptide NO: ID			6243872CD1 638		
SEQ NO ID SEQ	cont.		15		

Table 3

Analytical Methods and Databases	BLAST_DOMO	HMMER_PFAM	TMHMMER			BLAST_PRODOM	BLAST_PRODOM	BLAST_PRODOM	BLAST_PRODOM
Signature Sequences, Domains and Motifs	ANION; ORGANIC; TRANSPORTER; INDEPENDENT; DM03420 Q00910 1-642; G161-T618, N103-V159 DM03420 P46721 1-638; G156-I591 DM03420 A49580 1-638; G156-I591	Transmembrane amino acid transporter protein: A102-G482	Cytosolic domains: K131-R171, R243-L248, L309-P320, S395-D406, S457-D468 Transmembrane domains: 1108-V130, R172-A194.	S225-1242, S249-V271, W286-P308, L321-L343, I372-V394, L407-I424, V434-Y456, A469-I491	Non-cytosolic domains: M1-G107, D195-D224, Q272-P285, Q344-S371, P425-L433, Q492-I507	ACID AMINO PROTEIN TRANSPORTER PERMEASE TRANSMEMBRANE INTERGENIC REGION PUTATIVE PROLINE PD001875: W80- I.380	PROTEIN F59B2.2 Y43F4B.7 H32K16.1 SIMILAR C ELEGANS CHROMOSOME III PD150950: V174 L333	PROTEIN F59B2.2 T27A1.5 Y43F4B.7 H32K16.1 SIMILAR C ELEGANS CHROMOSOME III PD150568: V328-Y456	PROTEIN Y43F4B,7 H32K16.1 SIMILAR C ELEGANS F59B2.2 PD018165: N76-R164
Potential Glycosylation Sites		N62 N76 N141 N205 N214 N256							
Potential Phosphorylation Sites		S4 S46 S51 S55 S143 S365 T34							
Amino Acid Potential Residues Phospho Sites		507							
Incyte Polypeptide ID		90011608CD1 507							
SEQ ID	cont.	16							

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Analytical Methods and Databases	HMMER	HMMER_PFAM	TMHMMER	BLIMPS_BLOCKS	BLAST_PRODOM	BLAST_DOMO
Signature Sequences, Domains and Motifs	Signal Peptide: M1-A33, M1-C38	Sodium: sulfate symporter transmembrane: S6-L554	Cytosolic domains: M1-K12, P68-N87, Q157-L210, Y276-K308, P371-G409, N465-G484, N545-T568 Transmembrane domains: S13-L30, M45-F67, M88-L105, P134-L156, C211-L233, W253-V275, L309-F331, Y353-V370, I410-W432, A442-S464, L485-A507, V522-V544 Non-cytosolic domains: M31-L44, H106-A133, L234-S252, S332-K352, M433-H441, T508-K521	Sodium: sulfate symporter family proteins BL01271: T132-I151, M208-V232, P407-G428, S480-I534	PROTEIN TRANSMEMBRANE TRANSPORT MEMBRANE INNER TRANSPORTER SODIUM SYMPORT OF COTRANSPORTER PD000549: F323-W547, V15-V173, M208-K374, I16-L170	RENAL; BOUND; PRO-SER-ALA; NA; DM02914 P46556 1-520; R37-F552 DM02914 A47714 28-576; T144-F552, I29-K206 DM02914 S43561 28-507; E290-T546, R37-M159, V152-K388, A4-V41 DM02914 P32739 25-517; D174-F552, R37-M159, P21-T86
Potential Glycosylation Sites	N562					
Potential Phosphorylation Sites	S291 S313 S372 S381 T383 T564					
Amino Acid Potential Residues Phospho Sites	895					
Incyte Polypeptide ID	90024583CD1 568					
S B S	17					

Analytical Methods and Databases	S	MER	S	R	HIMMER_PFAM	MER	S_BLOCKS
Analytical Meand Databases	MOTIFS	TMEMMER	MOTIFS	HMMER	HIMIME	TMHMMER	BLIMP
Signature Sequences, Domains and Motifs	Sodium: sulfate symporter family signature: A498-V514	Cytosolic domains: MI-S144, MI91-R196, L247-Q252, R401-G404, E470-V475, P528-M595 Transmembrane domains: L145-G164, V168-T190, F197-R214, F224-A246, V253-F272, Y378-G400, L405-G427, 1447-A469, 1476-1498, L508-L527 Non-cytosolic domains: R165-P167, I215-R223, P273-D377, K428-S446, E499-F507	Sugar transport proteins signature 1: T156-S172	Signal Peptide: M1-A33	Sodium: sulfate symporter transmembrane: P213-L573, A7-L162	Cytosolic domains: M1-V11, P77-K80, K160-L235, Y301-N334, P393-A467, P528-T547 Transmembrane domains: W12-L34, T54-L76, V81-1103, F137-L159, I236-I258, W278-L300, L335-F357, L370-F392, V468-I490, H505-T527, G548-I570 Non-cytosolic domains: P35-C53, E104-S136, L259-S277, T358-S369, F491-V504, F571-L602	Sodium: sulfate symporter family proteins BL01271: BLIMPS_BLOCKS T135-I154, F234-I258, P426-G447, A499-L553
Potential Glycosylation Sites		N60 N79 N82 N103 N188 N538		13 S78 S101 S220 N312 N586 N596 305 S394 S398 318 T410 T417 575 T598			
Potential Phosphorylation Sites		S84 S96 S105 S126 N60 N79 N82 S275 S287 S319 N103 N188 N S440 S540 T52 T194 T296 T563 T582		S13 S78 S101 S220 S305 S394 S398 T318 T410 T417 T575 T598			
Amino Acid Potential Residues Phosphor Sites				602			
Incyte Polypeptide ID		90113658CD1 595		3942766CD1			
S 0 S	17 cont.	18		19			

Analytical Methods	and Databases	BLAST_PRODOM	BLAST_DOMO	TMFMMER	HMMER_PFAM	BLAST_PRODOM	BLAST_DOMO
Signature Sequences, Domains and Motifs		PROTEIN TRANSMEMBRANE TRANSPORT MEMBRANE INNER TRANSPORTER SODIUM SYMPORT OF CO-TRANSPORTER PD000549: 1328-W566, L17-L162, L235-W402	RENAL; BOUND; PRO-SER-ALA; NA; DM02914 BLAST_DOMO A47714 28-576: A562-F571 S43561 28-507: G233-A567, E38-L162, L549-N596 P46556 1-520: K204-F571, E38-E177 P32739 25-517: K232-F571, E38-A182, L21-A63	Cytosolic domains: M1-A77, D244-D254 Transmembrane domains: I78-Y100, Y221-L243, F255-L274 Non-cytosolic domains: S101-Y220, K275-V372	K+ channel tetramerisation domain: R63-S159	POTASSIUM CHANNEL IONIC CHANNEL PD060713: M1-R62	CHANNEL; POTASSIUM; CDRK; SHAW; DM00490 JH0595 26-142: R63-D151 P15387 18-134: R63-D151 P17970 268-384: R64-D149 P17972 1-102: D60-S159
Potential	Glycosylation Sites			;			
Potential	Phosphorylation Sites			S24 S29 S162 S180	S9 S146 T79 T80 T96 T111		
Amino Acid Potential	Residues			372	165		
SEQ Incyte	Polypeptide ID			7501987CD1 372	7503223CD1 165		
SEQ	a ë	cont.		70	21		

Analytical Methods	and Databases	TMHMMER			PROFILESCAN		PROFILESCAN	BLIMPS BLOCKS	BLAST DOMO	1			MOTIFS	TMHMMER			BLIMPS_BLOCKS	BLAST_PRODOM				BLAST_DOMO				
Signature Sequences, Domains and Motifs			Transmembrane domain: W34-T56	Non-cytosolic domain: M1-D33	per/Zinc superoxide dismutase signatures: L12-	R60	Nickel-dependent hydrogenases large subunit	F8 family BL01310: S28-1.63	山		IA55571[20-87: A20-G52	I48648 20-88; A20-G52	ATP1G1 / PLM / MAT8 family signature: S28-G41	Cytosolic domain: T138-Q152	Transmembrane domain: A115-G137	Non-cytosolic domain: M1-L114	Sulfate transporter IPB001902: L105-Q152, L75-Y84 BLIMPS_BLOCKS	SULFATE TRANSPORTER, TRANSPORT	PROTEIN, TRANSMEMBRANE	GLYCOPROTEIN, HIGH AFFINITY SULPHATE,	PERMEASE PD001121: L71-G145	RS DM01229	P50443 49-505: Q61-L146	P40879 5-462: Y29-G145	P45380 10-468: C62-L146	P38359 53-515: H54-G145
	Glycosylation Sites																									
Potential	Phosphorylation Sites													T138												
Amino Acid Potential	Residues													152												
Incyte														7511620CD1												
SEQ	<u>.</u> .	23	cont.											24												

3-P 191, 191, 19, P	3.1 3.1 3.1 3.1 3.1 3.1	19 19 13 15 16 16 16 16 16 16 16 16 16 16 16 16 16		E 1346	
Cytosolic domains: M1-L22, T192-T203, S253-P272, TMHMMER N333-W338, D391-H394, L450-E497 Transmembrane domains: V23-V45, P169-Y191, A204-V226, A230-P252, Y273-L295, L310-A332, L339-I361, L371-V390, T395-P417, I427-Y449 Non-cytosolic domains: P46-I168, G227-S229, P296-Q309, F362-G370, S418-A426 CHROMAFFIN GRANULE AMINE TRANSPORTER VESICULAR VAT1 TRANSPORT TRANSMEMBRANE GLYCOPROTEIN NEUROTRANSMITTER	1-L22, T192-T203, S253-1 394, L450-E497 uins: V23-V45, P169-Y191 52, Y273-L295, L310-A37 0, T395-P417, 1427-Y449 s: P46-1168, G227-S229, F 18-A426 NULE AMINE SICULAR VAT1 SICULAR VAT1 SICULAR VAT1 SICULAR VAT1	1-L22, T192-T203, S253-394, L450-E497 ins: V23-V45, P169-Y19 52, Y273-L295, L310-A3 0, T395-P417, I427-Y449 s: P46-1168, G227-S229, I 18-A426 NULE AMINE SICULAR VAT1 SMEMBRANE SUROTRANSMITTER NULE AMINE	1-L22, T192-T203, S253 394, L450-E497 ins: V23-V45, P169-Y19 52, Y273-L295, L310-A- 0, T395-P417, 1427-Y44 s: P46-1168, G227-S229, 18-A426 NULE AMINE SICULAR VAT1 SMEMBRANE SUROTRANSMITTER SICULAR 1 VAT1 SMEMBRANE SUROTRANSMITTER 6 5177 NULE AMINE SICULAR 1 VAT1 SMEMBRANE FICULAR 1 VAT1 SMEMBRANE 8-E497 8-E497	1-L22, T192-T203, S253 394, L450-E497 ins: V23-V45, P169-Y19 52, Y273-L295, L310-A-70, T395-P417, 1427-Y44 s: P46-1168, G227-S229, 18-A426 NULE AMINE SIMEMBRANE SICULAR VAT1 SIMEMBRANE SICULAR I VAT1 SIMEMBRANE SICULAR I VAT1 SIMEMBRANE SICULAR I VAT1 SIMEMBRANE SICULAR I VAT1 SIMEMBRANE TTC VESICLE AMINE 6 77 PTIC VESICLE AMINE 170805: M186-D271 N DOMAIN DM03018 8-E497 8-G486 8-S452	1-L22, T192-T203, S253 394, L450-E497 ins: V23-V45, P169-Y19 52, Y273-L295, L310-A3 0, T395-P417, I427-Y44 s: P46-I168, G227-S229, 18-A426 NULE AMINE SICULAR VAT1 SMEMBRANE SUROTRANSMITTER NULE AMINE SICULAR 1 VAT1 SMEMBRANE 51CULAR 1 VAT1 SMEMBRANE 77 77 8-E497 8-E497 8-G86 8-S452 56-R451
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nbrane domains: V23-V ² 26, A230-P252, Y273-L ² 1, L371-V390, T395-P41 solic domains: P46-I168, 62-G370, S418-A426 AFFIN GRANULE AMI ORTER VESICULAR VORT TRANSMEMBRA PROTEIN NEUROTRAN	nbrane domains: V23-V ² 26, A230-P252, Y273-L ² 1, L371-V390, T395-P41 solic domains: P46-1168, 62-G370, S418-A426 AFFIN GRANULE AMI ORTER VESICULAR V ORT TRANSMEMBRA PROTEIN NEUROTRAN 6: M1-G128	nbrane domains: V23-V ² 26, A230-P252, Y273-L ² 1, L371-V390, T395-P41 solic domains: P46-I168, 62-G370, S418-A426 AFFIN GRANULE AMI ORTER VESICULAR V ORT TRANSMEMBRA PROTEIN NEUROTRAN 6: M1-G128 AFFIN GRANULE AMI	nbrane domains: V23-V26, A230-P252, Y273-L2 1, L371-V390, T395-P41 solic domains: P46-1168, 62-G370, S418-A426 AFFIN GRANULE AMI ORTER VESICULAR V ORT TRANSMEMBRA ROTEIN NEUROTRAN 6: M1-G128 AFFIN GRANULE AMI ORTER VESICULAR 1 ORT TRANSMEMBRA YROTEIN NEUROTRAN 5: P217-F266 9: A461-E497 CTO SYNAPTIC VESIC ORTER PD170805: M1 LAR LUMEN DOMAIN 172-524: F178-E497 140-492: F178-G486 57-516: F178-S452	nbrane domains: V23-V26, A230-P252, Y273-L2 1, L371-V390, T395-P41 solic domains: P46-1168, 62-G370, S418-A426 AFFIN GRANULE AMI ORTER VESICULAR VORT TRANSMEMBRA ROTEIN NEUROTRAN 6: M1-G128 AFFIN GRANULE AMI ORTER VESICULAR 1 ORTER VESICULAR 1 ORTER VESICULAR 1 CRT TRANSMEMBRA AFFIN GRANULE AMI ORTER VESICULAR 1 CRT TRANSMEMBRA AFFIN GRANULE AMI ORTER VESICULAR 1 CRT TRANSMEMBRA 10-R71-F266 19: A461-E497 10-524: F178-E497 172-524: F178-E497 140-492: F178-G486 57-516: F178-S452	nbrane domains: V23-V26, A230-P252, Y273-L2 1, L371-V390, T395-P41 solic domains: P46-1168, 62-G370, S418-A426 AFFIN GRANULE AMI ORTER VESICULAR V ORT TRANSMEMBRA PROTEIN NEUROTRAN 6: M1-G128 AFFIN GRANULE AMI ORT TRANSMEMBRA PROTEIN NEUROTRAN 7: P217-F266 9: A461-E497 TO SYNAPTIC VESIC ORTER PD170805: M1 LAR LUMEN DOMAIN (72-524: F178-E497 (40-492: F178-G486 57-516: F178-S452 (35-500: Y166-R451) eavage: M1-A20
NEUDIANG WOMANDS. V226, A230-P252, Y 361, L371-V390, T3 rtosolic domains: P4 F362-G370, S418-A MAFFIN GRANUL SPORTER VESICU SPORT TRANSME OPROTEIN NEURG	HEMORANG WOMANNS. V226, A230-P252, Y 1361, L371-V390, T3 ytosolic domains: P4(F362-G370, S418-A MAAFFIN GRANUL. ISPORTER VESICU ISPORT TRANSME COPROTEIN NEURC 1226: M1-G128	A204-V226, A230-P252, Y L339-I361, L371-V390, T3 Non-cytosolic domains: P40 Q309, F362-G370, S418-A CHROMAFFIN GRANULI TRANSPORT TRANSMEI GLYCOPROTEIN NEURC PD034226: M1-G128 CHROMAFFIN GRANULI	A204-V226, A230-P252, Y L339-I361, L371-V390, T3 Non-cytosolic domains: P46 Q309, F362-G370, S418-A Q309, F362-G370, S418-A CHROMAFFIN GRANULI TRANSPORTER VESICU TRANSPORTER VESICU TRANSPORTER VESICU TRANSPORTER VESICU TRANSPORTER VESICU TRANSPORTER VESICU TRANSPORTER PO178 GLYCOPROTEIN NEURC PD034225: P217-F266 PD164279: A461-E497 SIMILAR TO SYNAPTIC TRANSPORTER PD1708 VESICULAR LUMEN DO [P54219]172-524: F178-E4 [S43685]140-492: F178-G4	226, A230-P252, Y 61, L371-V390, T3 osolic domains: P4 362-G370, S418-A IAFFIN GRANULI PORT TRANSMEI	A204-V226, A230-P252, Y L339-I361, L371-V390, T3 Non-cytosolic domains: P4(Q309, F362-G370, S418-A CHROMAFFIN GRANULI TRANSPORT TRANSMEI GLYCOPROTEIN NEURC PD034226: M1-G128 CHROMAFFIN GRANULI TRANSPORT TRANSMEI GLYCOPROTEIN NEURC PD034226: P217-F266 PD164279: A461-E497 SIMILAR TO SYNAPTIC TRANSPORTER PD1708 VESICULAR LUMEN DO P54219 172-524: F178-E4 S43685 140-492: F178-G4 I38658 157-516: F178-G4
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361, L371-V390, T ytosolic domains: Pt F362-G370, S418-4 MAFFIN GRANUI SPORTER VESICI SPORT TRANSME OPROTEIN NEUR	1361, L371-V390, T ytosolic domains: P4 F362-G370, S418-4 MAAFFIN GRANUI ISPORTER VESICI ISPORT TRANSME COPROTEIN NEUR 1226: M1-G128	340, L371-V390, T sytosolic domains: Por F362-G370, S418-4 OMAFFIN GRANUI SPORTER VESICK SPORT TRANSME COPROTEIN NEUR 4226: M1-G128 OMAFFIN GRANUI	361, L371-V390, T tosolic domains: P4 e362-G370, S418-VAFEIN GRANUI SPORTER VESICU SPORT TRANSME DOROTEIN NEUR AAFEIN GRANUI SPORT TRANSME SPORT TRANSME DPROTEIN NEUR 225: P217-F266 279: A461-E497 AR TO SYNAPTIC SPORTER PD170: ULAR LUMEN DV	61, L371-V390, T osolic domains: P4 362-G370, S418-P IAFFIN GRANUI PORTER VESICU PORT TRANSME 126: M1-G128 IAFFIN GRANUI PORT TRANSME PORT TRANSME PORT TRANSME PORTER VESICU PORTER VESICU PORTER VESICU PORTER PD170! TA SYNAPTIC PORTER PD170! TAS LUMEN D170! J172-524: F178-E- [140-492: F178-G-157-516: F178-G-157-516: F178-G-157-516-F178-G-157-516-F178-G-157-516-F178-G-157-516-F178-G-157-516-F178-G-157-516-F178-G-157-516-F178-G-157-516-F178-G-157-516-F178-G-157-516-F178-G-157-516-F178-G-157-516-F178-G-157-516-F178-G-F178-516-F178-G-F178-516-F178-S-516-F178-	61, L371-V390, T osolic domains: P4 362-G370, S418-/ IAFFIN GRANUI PORT TRANSME PORT TRANSME A4FIN GRANUI PORTER VESICI PORT TRANSME 25: P217-F266 25: P217-F266 17: A461-E497 R TO SYNAPTIC PORTER PD1703 ILAR LUMEN DI 172-524: F178-E 140-492: F178-G 157-516: F178-G 1135-500: Y166-R
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SPORTER VESICULAR VAT1 SPORT TRANSMEMBRANE OPROTEIN NEUROTRANSMITTER	ISPORTER VESICULAR VAT1 ISPORT TRANSMEMBRANE YOPROTEIN NEUROTRANSMITTER 1226: M1-G128	ASPORTER VESICULAR VAT1 ASPORT TRANSMEMBRANE COPROTEIN NEUROTRANSMITTER 4226: M1-G128 OMAFFIN GRANULE AMINE	TRANSPORTER VESICULAR VAT1 TRANSPORT TRANSMEMBRANE GLYCOPROTEIN NEUROTRANSMITTER PD034226: MI-G128 CHROMAFFIN GRANULE AMINE TRANSPORTER VESICULAR 1 VAT1 TRANSPORT TRANSMEMBRANE GLYCOPROTEIN NEUROTRANSMITTER PD034225: P217-F266 PD164279: A461-E497 SIMILAR TO SYNAPTIC VESICLE AMINI TRANSPORTER PD170805: M186-D271 VESICULAR LUMEN DOMAIN DM03018 [P54219]172-524: F178-E497 [S43685]140-492: F178-G486	PORTER VESICULAR VAT1 PORT TRANSMEMBRANE PROTEIN NEUROTRANSMITTER 26: MI-G128 IAFFIN GRANULE AMINE PORT TRANSMEMBRANE PORT TRANSMEMBRANE 25: P217-F266 T9: A461-E497 R TO SYNAPTIC VESICLE AMINI PORTER PD170805: M186-D271 JLAR LUMEN DOMAIN DM03018 172-524: F178-E497 140-492: F178-G486 157-516: F178-S452	PORTER VESICULAR VAT1 PORT TRANSMEMBRANE PROTEIN NEUROTRANSMITTER 26: MI-G128 IAFFIN GRANULE AMINE PORTER VESICULAR I VAT1 PORT TRANSMEMBRANE PORTER VESICULAR I VAT1 PORT TRANSMEMBRANE 125: P217-F266 T9: A461-E497 R TO SYNAPTIC VESICLE AMIN PORTER PD170805: M186-D271 JLAR LUMEN DOMAIN DM03018 172-524: F178-E497 140-492: F178-G486 157-516: F178-S452 1135-500: Y166-R451
SPORT TRANSMEMBRANE OPROTEIN NEUROTRANSMITTER	ISPORT TRANSMEMBRANE COPROTEIN NEUROTRANSMITTER 1226: MI-G128	TRANSPORT TRANSMEMBRANE GLYCOPROTEIN NEUROTRANSMITTER PD034226: M1-G128 CHROMAFFIN GRANULE AMINE	TRANSPORT TRANSMEMBRANE GLYCOPROTEIN NEUROTRANSMITTER PD034226: M1-G128 CHROMAFFIN GRANULE AMINE TRANSPORT TRANSMEMBRANE GLYCOPROTEIN NEUROTRANSMITTER PD034225: P217-F266 PD164279: A461-E497 SIMILAR TO SYNAPTIC VESICLE AMINE TRANSPORTER PD170805: M186-D271 VESICULAR LUMEN DOMAIN DM03018 [P54219]172-524: F178-G486 [I38658]157-516: F178-S452	PORT TRANSMEMBRANE PPROTEIN NEUROTRANSMITTER 26: M1-G128 IAFFIN GRANULE AMINE PORTER VESICULAR I VATI PORT TRANSMEMBRANE PORT TRANSMEMBRANE 125: P217-F266 T9: A461-E497 R TO SYNAPTIC VESICLE AMINE PORTER PD170805: M186-D271 ILAR LUMEN DOMAIN DM03018 177-524: F178-E497 157-516: F178-S452 157-516: F178-S451	PORT TRANSMEMBRANE PPROTEIN NEUROTRANSMITTER 26: M1-G128 IAFFIN GRANULE AMINE PORTER VESICULAR 1 VAT1 PORT TRANSMEMBRANE PPROTEIN NEUROTRANSMITTER 25: P217-F266 79: A461-E497 R TO SYNAPTIC VESICLE AMINE PORTER PD170805: M186-D271 JLAR LUMEN DOMAIN DM03018 [172-524: F178-E497 [140-492: F178-G486 157-516: F178-S452 [135-500: Y166-R451
OPROTEIN NEUROTRANSMITTER	OPROTEIN NEUROTRANSMITTER 1226: M1-G128	COPROTEIN NEUROTRANSMITTER 1226: M1-G128 MAFFIN GRANULE AMINE	DPROTEIN NEUROTRANSMITTER 226: MI-G128 MAFFIN GRANULE AMINE SPORT TRANSMEMBRANE DPROTEIN NEUROTRANSMITTER 225: P217-F266 279: A461-E497 AR TO SYNAPTIC VESICLE AMINE SPORTER PD170805: M186-D271 ULAR LUMEN DOMAIN DM03018 9 172-524: F178-E497 5 140-492: F178-G486	PROTEIN NEUROTRANSMITTER 26: MI-G128 IAFFIN GRANULE AMINE PORTER VESICULAR I VAT1 PORT TRANSMEMBRANE PROTEIN NEUROTRANSMITTER 25: P217-F266 79: A461-E497 R TO SYNAPTIC VESICLE AMINE PORTER PD170805: M186-D271 JLAR LUMEN DOMAIN DM03018 [172-524: F178-E497 [140-492: F178-G486 [157-516: F178-S452	PROTEIN NEUROTRANSMITTER 26: MI-G128 IAFFIN GRANULE AMINE PORTER VESICULAR 1 VAT1 PORT TRANSMEMBRANE PROTEIN NEUROTRANSMITTER 25: P217-F266 79: A461-E497 R TO SYNAPTIC VESICLE AMINE PORTER PD170805: M186-D271 ILAR LUMEN DOMAIN DM03018 172-524: F178-E497 1135-500: Y166-R451 liasvage: M1-A20
	:226: M1-G128	MAFFIN GRANULE AMINE	226: M1-G128 MAFFIN GRANULE AMINE SPORTER VESICULAR 1 VAT1 SPORT TRANSMEMBRANE 225: P217-F266 279: A461-E497 AR TO SYNAPTIC VESICLE AMINE SPORTER PD170805: M186-D271 ULAR LUMEN DOMAIN DM03018 9 172-524: F178-E497 5 140-492: F178-G486	26: M1-G128 IAFFIN GRANULE AMINE PORT TRANSMEMBRANE PPROTEIN NEUROTRANSMITTER 25: P217-F266 79: A461-E497 R TO SYNAPTIC VESICLE AMINE PORTER PD170805: M186-D271 ILAR LUMEN DOMAIN DM03018 [172-524: F178-E497 [140-492: F178-G486 157-516: F178-S452	26: M1-G128 IAFFIN GRANULE AMINE PORT TRANSMEMBRANE PPROTEIN NEUROTRANSMITTER 25: P217-F266 79: A461-E497 R TO SYNAPTIC VESICLE AMINE PORTER PD170805: M186-D271 ILAR LUMEN DOMAIN DM03018 172-524: F178-E497 140-492: F178-G486 157-516: F178-S452 135-500: Y166-R451
CHRUMAFFIN GRANULE AMINE TRANSPORTER VESICULAR 1 VAT1 TRANSPORT TRANSMEMBRANE	IKANSPORTER VESICULAR I VATI TRANSPORT TRANSMEMBRANE		225. P217-F266 279: A461-E497 AR TO SYNAPTIC VESICLE AMINE SPORTER PD170805: M186-D271 ULAR LUMEN DOMAIN DM03018 9 172-524: F178-E497 5 140-492: F178-G486	25: P217-F266 79: A461-E497 R TO SYNAPTIC VESICLE AMINE PORTER PD170805: M186-D271 JLAR LUMEN DOMAIN DM03018 [172-524: F178-E497 140-492: F178-G486 157-516: F178-S452	25: P217-F266 79: A461-E497 R TO SYNAPTIC VESICLE AMINE PORTER PD170805: M186-D271 JLAR LUMEN DOMAIN DM03018 [172-524: F178-E497 [140-492: F178-G486 157-516: F178-S452 [135-500: Y166-R451
MALFIN GRANULE AMINE SPORTER VESICULAR I VATI SPORT TRANSMEMBRANE OPROTEIN NEUROTRANSMITTER	ISPORTER VESICOLAR I VAII ISPORT TRANSMEMBRANE OOROTEIN NEUROTRANSMITTER	GLYCOPROTEIN NEUROTRANSMITTER	279: A461-E497 AR TO SYNAPTIC VESICLE AMINE SPORTER PD170805: M186-D271 ULAR LUMEN DOMAIN DM03018 9 172-524: F178-E497 5 140-492: F178-G486 1 57-516: F178-S452	79: A461-E497 R TO SYNAPTIC VESICLE AMINE PORTER PD170805: M186-D271 JLAR LUMEN DOMAIN DM03018 [172-524: F178-E497 [140-492: F178-G486 157-516: F178-S452 [135-500: Y166-R451	79: A461-E497 R TO SYNAPTIC VESICLE AMINE PORTER PD170805: M186-D271 JLAR LUMEN DOMAIN DM03018 [172-524: F178-E497 [140-492: F178-G486 157-516: F178-S452 [135-500: Y166-R451
MAFFIN GRANULE AMINE SPORTER VESICULAR I VATI SPORT TRANSMEMBRANE OPROTEIN NEUROTRANSMITTER 225: P217-F266	SPORTER VESTCOLARI I VATI SPORT TRANSMEMBRANE OPROTEIN NEUROTRANSMITTER 225: P217-F266	OPROTEIN NEUROTRANSMITTER 225: P217-F266	AR TO SYNAPTIC VESICLE AMINE SPORTER PD170805; M186-D271 ULAR LUMEN DOMAIN DM03018 9 172-524; F178-E497 5 140-492; F178-G486 1 57-516; F178-S452	R TO SYNAPTIC VESICLE AMINE PORTER PD170805: M186-D271 LAR LUMEN DOMAIN DM03018 172-524: F178-E497 157-516: F178-S452 157-516: F178-S451	R TO SYNAPTIC VESICLE AMINE PORTER PD170805: M186-D271 JLAR LUMEN DOMAIN DM03018 [172-524: F178-E497 [140-492: F178-G486 157-516: F178-S452 [135-500: Y166-R451
AAFFIN GRANULE AMINE PORTER VESICULAR I VATI PORT TRANSMEMBRANE 125: P217-F266 179: A461-E497	PORT TRANSMEMBRANE PORT TRANSMEMBRANE PROTEIN NEUROTRANSMITTER 25: P217-F266	DPROTEIN NEUROTRANSMITTER 25: P217-F266 79: A461-E497	PORTER PD170805: M186-D271 JLAR LUMEN DOMAIN DM03018 172-524: F178-E497 140-492: F178-G486 157-516: F178-S452	PORTER PD170805: M186-D271 JLAR LUMEN DOMAIN DM03018 [172-524: F178-E497 [140-492: F178-G486 157-516: F178-S452 [135-500: Y166-R451	PORTER PD170805: M186-D271 JLAR LUMEN DOMAIN DM03018 [172-524: F178-E497 [140-492: F178-G486 157-516: F178-S452 [135-500: Y166-R451 sleavage: M1-A20
PAFFIN GRANULE AMINE PORTER VESICULAR I VATI PORT TRANSMEMBRANE PROTEIN NEUROTRANSMITTER 25: P217-F266 79: A461-E497 R TO SYNAPTIC VESICLE AMINE	PORTER VESICULAR I VALI PORT TRANSMEMBRANE PROTEIN NEUROTRANSMITTER 25: P217-F266 P9: A461-E497 R TO SYNAPTIC VESICLE AMINE	PROTEIN NEUROTRANSMITTER 25: P217-F266 79: A461-E497 R TO SYNAPTIC VESICLE AMINE	LAK LUMEN DUMAIN DM03018 172-524: F178-E497 140-492: F178-G486 157-516: F178-S452	LAR LUMEN DOMAIN DM03018 172-524: F178-E497 140-492: F178-G486 157-516: F178-S452 135-500: Y166-R451	LAR LUMEN DOMAIN DM03018 172-524: F178-E497 140-492: F178-G486 157-516: F178-S452 135-500: Y166-R451 leavage: M1-A20
PORTER VESICULAR I VATI PORT TRANSMEMBRANE PROTEIN NEUROTRANSMITTER 25. P217-F266 79. A461-E497 R TO SYNAPTIC VESICLE AMINE PORTER PD170805: M186-D271	PORT ER VESICOLAR I VALI PORT TRANSMEMBRANE PROTEIN NEUROTRANSMITTER 25: P217-F266 79: A461-E497 R TO SYNAPTIC VESICLE AMINE PORTER PD170805: M186-D271	PROTEIN NEUROTRANSMITTER 25: P217-F266 79: A461-E497 R TO SYNAPTIC VESICLE AMINE PORTER PD170805: M186-D271	172-224. F178-E497 140-492: F178-G486 157-516: F178-S452	172-224. F178-E497 140-492: F178-G486 157-516: F178-S452 135-500: Y166-R451	172-224, F178-E497 140-492; F178-G486 157-516; F178-S452 135-500; Y166-R451 leavage; M1-A20
PORTER VESICULAR I VATI PORTER VESICULAR I VATI PORT TRANSMEMBRANE PPROTEIN NEUROTRANSMITTER 25: P217-F266 79: A461-E497 R TO SYNAPTIC VESICLE AMINE PORTER PD170805: M186-D271 JLAR LUMEN DOMAIN DM03018	PORT TRANSMEMBRANE PORT TRANSMEMBRANE PPROTEIN NEUROTRANSMITTER 25: P217-F266 79: A461-E497 IR TO SYNAPTIC VESICLE AMINE PORTER PD170805: M186-D271 ILAR LUMEN DOMAIN DM03018	PPROTEIN NEUROTRANSMITTER 25: P217-F266 79: A461-E497 IR TO SYNAPTIC VESICLE AMINE PORTER PD170805: M186-D271 JLAR LUMEN DOMAIN DM03018	157-516: F178-S452	157-516: F178-S452 135-500: Y166-R451	157-516; F178-S452 135-500; Y166-R451 Jeavage: M1-A20
MAFFIN GRANULE AMINE SPORTER VESICULAR I VATI SPORT TRANSMEMBRANE DPROTEIN NEUROTRANSMITTER 225: P217-F266 279: A461-E497 AR TO SYNAPTIC VESICLE AMINE SPORTER PD170805: M186-D271 ULAR LUMEN DOMAIN DM03018 9 172-524: F178-E497 5 140-492: F178-G486	SPORTER VESICULAR I VALI SPORT TRANSMEMBRANE DPROTEIN NEUROTRANSMITTER 225: P217-F266 279: A461-E497 AR TO SYNAPTIC VESICLE AMINE SPORTER PD170805: M186-D271 ULAR LUMEN DOMAIN DM03018 9 172-524: F178-E497	DPROTEIN NEUROTRANSMITTER 225: P217-F266 279: A461-E497 AR TO SYNAPTIC VESICLE AMINE SPORTER PD170805: M186-D271 ULAR LUMEN DOMAIN DM03018 9 172-524: F178-E497 5 140-492: F178-G486		135-500: Y166-R451	135-500: Y166-R451 leavage: M1-A20

SEQ	SEQ Incyte	Amino Acid Potential		Potential	Signature Sequences, Domains and Motifs	Analytical Methods
<u>A</u>	Polypeptide Residues	Residues	Phosphorylation	Glycosylation Sites		and Databases
ÖN.	a a		Sites			
25	7506995CD1 467	467	S24 S29 S150 S254 N326 N333	N326 N333	Cytosolic domains: M1-A77, N293-G298, D375-	TIMHIMIMER
			S276 S381 T143		K386	
			T454		Transmembrane domains: I78-Y100, F270-Q292,	
					G299-V321, A355-L374, I387-L409	
					Non-cytosolic domains: S101-R269, A322-F354,	
					E410-R467	
<u> </u>	7506996CD1 490	490	S24 S29 S162 S180 N349 N356	N349 N356	Cytosolic domains: M1-A77, D244-D254, Q315-	TMHMMER
			S404 T477		F320, D398-K409	
					Transmembrane domains: I78-Y100, Y221-L243,	
					F255-F277, F292-V314, G321-I343, A378-L397,	
					I410-L432	
					Non-cytosolic domains: S101-Y220, A278-V291,	
					V344-F377, E433-R490	

Polynucleotide	Sequence Fragments
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	1253-1770, 1266-1423, 1276-1705, 1286-1623, 1290-1938, 1299-2189, 1303-2066, 1309-1514, 1309-1550, 1309-
	1940, 1313-1877, 1317-1901, 1327-1596, 1362-2028, 1367-1631, 1370-2072, 1373-2066, 1377-1653, 1377-1837,
	1377-1911, 1377-1929, 1391-2024, 1408-1696, 1414-1923, 1425-2308, 1439-1694, 1444-2063, 1458-1766, 1460-
	2132, 1461-1913, 1461-2312, 1480-2294, 1482-1853, 1493-1961, 1499-2008, 1499-2312, 1518-1841, 1519-2107,
	1525-2262, 1529-2312, 1533-1805, 1536-2024, 1545-2144, 1553-2158, 1557-2160, 1562-2312, 1601-2062, 1603-
	2266, 1606-2253, 1608-2174, 1616-2285, 1617-1935, 1617-2035, 1617-2112, 1617-2186, 1617-2190,
	1617-2265, 1618-2160, 1655-2185, 1687-2009, 1687-2274, 1691-2275, 1702-1982, 1707-1970, 1707-2169, 1714-
	2219, 1721-2213, 1730-2264, 1774-2285, 1800-2048, 1810-2066, 1813-2059, 1817-2238, 1817-2240, 1818-2308,
	1835-2294, 1886-2154, 1896-2246, 1910-2218, 1941-2236, 1980-2262, 2002-2285, 2059-2219

Table 5

Polynucleotide SEQ	Incyte Project ID:	Representative Library
ID NO:		
27	1853191CB1	PROSNOT15
28	7497369CB1	KIDEUNE02
29	1700438CB1	BLADTUT05
30	535939CB1	HEARFET01
31	55118067CB1	LIVRTUT13
33	7500819CB1	CORPNOT02
34	7503413CB1	LPARNOT02
35	7500007CB1	BRAITUT02
36	7500025CB1	COLCTUT02
37	7502736CB1	BRANDIN01
38	7503570CB1	CARGDIT01
39	7504008CB1	THP1TXT03
40	7503559CB1	MCLDTXN03
41	6243872CB1	TESTNOT17
44	90113658CB1	PROSTUT04
45	3942766CB1	FIBRTXS07
46	7501987CB1	BRAENOT04
47	7503223CB1	PROSTUT10
48	7503566CB1	BSTMNON02
49	7505122CB1	SINTNOT13
50	7511620CB1	COLNNOT23
51	7506995CB1	BRAENOT04
52	7506996CB1	BRAENOT04

Table (

Library	Vector	Library Description
	NONE	NONE
BLADTUT05	pINCY	Library was constructed using RNA isolated from bladder tumor tissue removed from a 66-year-old Caucasian male during a radical prostatectomy, radical cystectomy, and urinary diversion. Pathology indicated grade 3 transitional cell carcinoma on the anterior wall of the bladder. Patient history included lung neoplasm and tobacco abuse in remission. Family history included malignant breast neoplasm, tuberculosis, cerebrovascular disease, atherosclerotic coronary artery disease, and lung cancer.
BRAENOT04	pINCY	Library was constructed using RNA isolated from inferior parietal cortex tissue removed from the brain of a 35-year-old Caucasian male who died from cardiac failure. Pathology indicated moderate leptomeningeal fibrosis and multiple microinfarctions of the cerebral neocortex. Patient history included dilated cardiomyopathy, congestive heart failure, cardiomegaly and an enlarged spleen and liver.
BRAITUT02	PSPORT1	Library was constructed using RNA isolated from brain tumor tissue removed from the frontal lobe of a 58-year-old Caucasian male during excision of a cerebral meningeal lesion. Pathology indicated a grade 2 metastatic hypernephroma. Patient history included a grade 2 renal cell carcinoma, insomnia, and chronic airway obstruction. Family history included a malignant neoplasm of the kidney.
BRANDIN01	pINCY	This normalized pineal gland tissue library was constructed from .4 million independent clones from a pineal gland tissue library throm two different donors. Starting RNA was made from pooled pineal gland tissue removed from two Caucasian females: a 68-year-old (donor A) who died from congestive heart failure and a 79-year-old (donor B) who died from pineumonia. Neuropathology for donor A indicated mild to moderate Alzheimer disease, atherosclerosis, and multiple infarctions. Neuropathology for donor B indicated severe Alzheimer disease, arteriolosclerosis, cerebral amyloid angiopathy and multiple infarctions. There were diffuse and neuritic amyloid plaques and neurofibrillary tangles throughout the brain sections examined in both donors. Patient history included diabetes mellitus, rheumatoid arthritis, hyperthyroidism, amyloid heart disease, and dementia in donor A; and pseudophakia, gastritis with bleeding, glaucoma, peripheral vascular disease, COPD, delayed onset tonic/clonic seizures, and transient ischemic attack in donor B. The library was normalized in one round using conditions adapted from Soares et al., PNAS (1994) 91:9228-9232 and Bonaldo
BSTMNON02	PSPORTI	This normalized brain stem library was constructed from 2.84 million independent clones from a brain stem library. Starting RNA was made from the brain stem tissue of a 72-year-old Caucasian male who died from myocardial infarction. Patient history included coronary artery disease, insulin-dependent diabetes mellitus, and arthritis. Normalization and hybridization conditions were adapted from Soares et al. (PNAS (1994) 91:9228).
CARGDIT01	pINCY	Library was constructed using RNA isolated from diseased cartilage tissue. Patient history included osteoarthritis.

Library	Vector	Library Description
COLCTUT02	pincy	Library was constructed using RNA isolated from colon tumor tissue removed from the cecum of a 30-year-old Caucasian female during partial colectomy, open liver biopsy, incidental appendectomy, and permanent colostomy. Pathology indicated carcinoid tumor (grade 1 neuroendocrine carcinoma) arising in the terminal ileum, forming a mass in the right colon. Patient history included chronic sinus infections and endometriosis. Family history included hyperlipidemia, anxiety, upper lobe lung cancer, stomach cancer, liver cancer, and cirrhosis.
COLNNOT23	pINCY	Library was constructed using RNA isolated from diseased colon tissue removed from a 16-year-old Caucasian male during a total colectomy with abdominal/perineal resection. Pathology indicated gastritis and pancolonitis consistent with the acute phase of ulcerative colitis. Inflammation was more severe in the transverse colon, with inflammation confined to the mucosa. There was only mild involvement of the ascending and sigmoid colon, and no significant involvement of the cecum, rectum, or terminal ileum. Family history included irritable bowel syndrome.
CORPNOT02	pINCY	Library was constructed using RNA isolated from diseased corpus callosum tissue removed from the brain of a 74-year-old Caucasian male who died from Alzheimer's disease.
FIBRTXS07	pINCY	This subtracted library was constructed using 1.3 million clones from a dermal fibroblast library and was subjected to two rounds of subtraction hybridization with 2.8 million clones from an untreated dermal fibroblast tissue library. The starting library for subtraction was constructed using RNA isolated from treated dermal fibroblast tissue removed from the breast of a 31-year-old Caucasian female. The cells were treated with 9CIS retinoic acid. The hybridization probe for subtraction was derived from a similarly constructed library from RNA isolated from untreated dermal fibroblast tissue from the same donor. Subtractive hybridization conditions were based on the methodologies of Swaroop et al., NAR (1991) 19:1954 and Bonaldo, et al., Genome Research (1996) 6:791.
HEARFET01	pINCY	Library was constructed using RNA isolated from heart tissue removed from a Hispanic male fetus, who died at 18 weeks' gestation.
KIDEUNE02	pINCY	This 5' biased random primed library was constructed using RNA isolated from an untreated transformed embryonal cell line (293-EBNA) derived from kidney epithelial tissue (Invitrogen). The cells were transformed with adenovirus 5 DNA.

Library	Vector	Library Description
JT13	pINCY	Library was constructed using RNA isolated from liver tumor tissue removed from a 62-year-old Caucasian female during partial hepatectomy and exploratory laparotomy. Pathology indicated metastatic intermediate grade neuroendocrine carcinoma, consistent with islet cell tumor, forming nodules ranging in size, in the lateral and medial left liver lobe. The pancreas showed fibrosis, chronic inflammation and fat necrosis consistent with pseudocyst. The gall bladder showed mild chronic cholecystitis. Patient history included malignant neoplasm of the pancreas tail, pulmonary embolism, hyperlipidemia, thrombophlebitis, joint pain in multiple joints, type II diabetes, benign hypertension, and cerebrovascular disease. Family history included pancreas cancer, secondary liver cancer, benign hypertension, and hyperlipidemia.
LPARNOT02	pINCY	Library was constructed using RNA isolated from tissue obtained from the left parotid (salivary) gland of a 70-year-old male with parotid cancer.
MCLDTXN03 pinCY	pINCY	This normalized dendritic cell library was constructed from one million independent clones from a pool of two derived dendritic cell libraries. Starting libraries were constructed using RNA isolated from untreated and treated derived dendritic cells from umbilical cord blood CD34+ precursor cells removed from a male. The cells were derived with granulocyte/macrophage colony stimulating factor (GM-CSF), tumor necrosis factor alpha (TNF alpha), and stem cell factor (SCF). The GM-CSF was added at time 0 at 100 ng/ml, the TNF alpha was added at time 0 at 2.5 ng/ml, and the SCF was added at time 0 at 25 ng/ml. Incubation time was 13 days. The treated cells were then exposed to phorbol myristate acetate (PMA), and Ionomycin. The PMA and Ionomycin were added at 13 days for five hours. The library was normalized in two rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228-9232 and Bonaldo et al., Genome Research (1996) 6:791, except that a significantly longer (48 hours/round) reannealing hybridization was used.
PROSNOT15	pINCY	Library was constructed using RNA isolated from diseased prostate tissue removed from a 66-year-old Caucasian male during radical prostatectomy and regional lymph node excision. Pathology indicated adenofibromatous hyperplasia. Pathology for the associated tumor tissue indicated an adenocarcinoma (Gleason grade 2+3). The patient presented with elevated prostate specific antigen (PSA). Family history included prostate cancer, secondary bone cancer, and benign hypertension.
PROSTUT04	PSPORT1	Library was constructed using RNA isolated from prostate tumor tissue removed from a 57-year-old Caucasian male during radical prostatectomy, removal of both testes and excision of regional lymph nodes. Pathology indicated adenocarcinoma (Gleason grade 3+3). Patient history included a benign neoplasm of the large bowel and type I diabetes. Family history included a malignant neoplasm of the prostate and type I diabetes.

Library	Vector	Library Description
PROSTUT10 pINCY	pINCY	Library was constructed using RNA isolated from prostatic tumor tissue removed from a 66-year-old Caucasian male during radical prostatectomy and regional lymph node excision. Pathology indicated an adenocarcinoma (Gleason grade 2+3). Adenofibromatous hyperplasia was also present. The patient presented with elevated prostate specific antigen (PSA). Family history included prostate cancer and secondary bone cancer.
SINTNOT13	pINCY	Library was constructed using RNA isolated from ileum tissue obtained from a 25-year-old Asian female during a partial colectomy and temporary ileostomy. Pathology indicated moderately active chronic ulcerative colitis, involving colonic mucosa from the distal margin to the ascending colon. Family history included hyperlipidemia, depressive disorder, malignant cervical neoplasm, viral hepatitis A, and depressive disorder.
TESTNOT17 pINCY	pINCY	Library was constructed from testis tissue removed from a 26-year-old Caucasian male who died from head trauma due to a motor vehicle accident. Serologies were negative. Patient history included a hernia at birth, tobacco use (1 1/2 ppd), marijuana use, and daily alcohol use (beer and hard liquor).
THP1TXT03	pINCY	Library was constructed using RNA isolated from treated THP-1 cells. THP-1 is a human promonocyte line derived from the peripheral blood of a 1-year- old Caucasian male with acute monocytic leukemia (ref. Int. J. Cancer (1980) 26:171). The THP-1 cultured cells were differentiated with PMA(100ng/ml) for 48 hours, incubated with Mycobacteria tuberculosis, strain H37Rv, for 4 hours at 37C, washed and RNA extracted.

Program	Description	Reference	Parameter Threshold
ABI FACTURA	A program that removes vector sequences and masks Applied Biosystems, Foster City, CA. ambiguous bases in nucleic acid sequences.	Applied Biosystems, Foster City, CA.	
ABI/PARACEL FDF	A Fast Data Finder useful in comparing and annotating amino acid or nucleic acid sequences.	Applied Biosystems, Foster City, CA; Paracel Inc., Pasadena, CA.	Mismatch <50%
ABI AutoAssembler	A program that assembles nucleic acid sequences.	Applied Biosystems, Foster City, CA.	
BLAST	A Basic Local Alignment Search Tool useful in sequence similarity search for amino acid and nucleic 215:403-410; Altschul, S.F. et al. (1997) acid sequences. BLAST includes five functions: Nucleic Acids Res. 25:3389-3402. blastp, blastn, blastx, tblastn, and tblastx.	Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410; Altschul, S.F. et al. (1997) Nucleic Acids Res. 25:3389-3402.	ESTs: Probability value = 1.0E-8 or less; Full Length sequences: Probability value = 1.0E-10 or less
FASTA	A Pearson and Lipman algorithm that searches for similarity between a query sequence and a group of sequences of the same type. FASTA comprises as least five functions: fasta, tfasta, fastx, tfastx, and ssearch.	Pearson, W.R. and D.J. Lipman (1988) Proc. ESTs: fasta E value = 1.06E-6; Natl. Acad Sci. USA 85:2444-2448; Pearson, Assembled ESTs: fasta Identity W.R. (1990) Methods Enzymol. 183:63-98; = 95% or greater and Match and Smith, T.F. and M.S. Waterman (1981) length = 200 bases or greater; fastx. Adv. Appl. Math. 2:482-489. Full Length sequences: fastx score = 100 or greater	ESTs: fasta E value = 1.06E-6; Assembled ESTs: fasta Identity = 95% or greater and Match length = 200 bases or greater; fastx E value = 1.0E-8 or less; Full Length sequences: fastx score = 100 or greater
BLIMPS	A BLocks IMProved Searcher that matches a sequence against those in BLOCKS, PRINTS, DOMO, PRODOM, and PFAM databases to search for gene families, sequence homology, and structural fingerprint regions.	Henikoff, S. and J.G. Henikoff (1991) Nucleic Acids Res. 19:6565-6572; Henikoff, J.G. and S. Henikoff (1996) Methods Enzymol. 266:88-105; and Attwood, T.K. et al. (1997) J. Chem. Inf. Comput. Sci. 37:417- 424.	Probability value = 1.0E-3 or less

Program	Description	Reference	Parameter Threshold
HMMER	An algorithm for searching a query sequence against hidden Markov model (HMM)-based databases of protein family consensus sequences, such as PFAM, INCY, SMART and TIGRFAM.	 Krogh, A. et al. (1994) J. Mol. Biol. 235:1501-1531; Sonnhammer, E.L.L. et al. (1988) Nucleic Acids Res. 26:320-322; Durbin, R. et al. (1998) Our World View, in a Nutshell, Cambridge Univ. Press, pp. 1-350. 	PFAM, INCY, SMART or TIGRFAM hits: Probability value = 1.0E-3 or less; Signal peptide hits: Score = 0 or greater
ProfileScan	An algorithm that searches for structural and sequence motifs in protein sequences that match sequence patterns defined in Prosite.	Gribskov, M. et al. (1988) CABIOS 4:61-66; Normalized quality score ≥ GCG Gribskov, M. et al. (1989) Methods specified "HIGH" value for that Enzymol. 183:146-159; Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221.	Normalized quality score \geq GCG specified "HIGH" value for that particular Prosite motif. Generally, score = 1.4-2.1.
Phred	A base-calling algorithm that examines automated Ewing, B. et al. (1998) Genome Res. 8:175-sequencer traces with high sensitivity and probability. 185; Ewing, B. and P. Green (1998) Genome Res. 8:186-194.	Ewing, B. et al. (1998) Genome Res. 8:175-185; Ewing, B. and P. Green (1998) Genome Res. 8:186-194.	
Phrap	A Phils Revised Assembly Program including SWAT and CrossMatch, programs based on efficient implementation of the Smith-Waterman algorithm, useful in searching sequence homology and assembling DNA sequences. Smith, T.F. and M.S. Waterman (1981) T.F. and and (1981) J. Mol. Biol. 147 and Green, P., University of Washington, Seattle, WA.	Smith, T.F. and M.S. Waterman (1981) Adv. Score = 120 or greater; Match Appl. Math. 2:482-489; Smith, T.F. and M.S. Waterman (1981) J. Mol. Biol. 147:195-197; and Green, P., University of Washington, Seattle, WA.	Score = 120 or greater; Match length = 56 or greater
Consed	A graphical tool for viewing and editing Phrap assemblies.	Gordon, D. et al. (1998) Genome Res. 8:195-202.	
SPScan	A weight matrix analysis program that scans protein sequences for the presence of secretory signal peptides.	Nielson, H. et al. (1997) Protein Engineering 10:1-6; Claverie, J.M. and S. Audic (1997) CABIOS 12:431-439.	Score = 3.5 or greater
TMAP	A program that uses weight matrices to delineate transmembrane segments on protein sequences and determine orientation.	Persson, B. and P. Argos (1994) J. Mol. Biol. 237:182-192; Persson, B. and P. Argos (1996) Protein Sci. 5:363-371.	

Program	Description	Reference	Parameter Threshold
TMHMMER	that uses a hidden Markov model (HMM) transmembrane segments on protein nd determine orientation.	Sonnhammer, E.L. et al. (1998) Proc. Sixth Intl. Conf. On Intelligent Systems for Mol. Biol., Glasgow et al., eds., The Am. Assoc. for Artificial Intelligence (AAAI) Press, Menlo Park, CA, and MIT Press, Cambridge, MA, pp. 175-182.	
Motifs	A program that searches amino acid sequences for patterns that matched those defined in Prosite.	Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221; Wisconsin Package Program Manual, version 9, page M51-59, Genetics Computer Group, Madison, WI.	

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Hispanic Allele 1	frequency	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a
Asian Allele 1	frequency	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a
African Allele 1	frequency	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a
Caucasian Allele 1	frequency	n/a	n/a	n/a	1.00	0.17	n/a	n/a	n/a	n/a	1.00	0.17	n/a	n/a	n/a	n/a
EST Allele Allele Amino Acid Caucasian Allele 1 2 Allele 1		noncoding	noncoding	noncoding	noncoding	V390	noncoding	A296	A296	G175	noncoding	V413	noncoding	A319	A319	K164
Allele 2		G	G	Ğ	T	A	G	m L	2	A	T	A	G	T	C	A
Allele		A	A	A	Э	G	A	G	G	Ö	ນ	G	A	ප	Ŋ	G
EST		A	A	G	${f T}$	ව	A	T	ß	ß	H	ß	A	E	ڻ ڻ	A
CB1	!	2028	2039	2027	2120	1463	1871	1181	1181	819	2191	1534	1942	1252	1252	786
EST	5	413		348	143	158	132	25	53	174	143	158	132	25	53	234
SNP ID		SNP00099024 413	SNP00099024 430	SNP00099024 348	SNP00034867 143	SNP00039549 158	SNP00010061 132	SNP00039548 25	SNP00147613 53	SNP00039547 174	SNP00034867 143	SNP00039549 158	SNP00010061	SNP00039548	SNP00147613 53	SNP00135254 234
ESTID		1733477T6	2416475T6	7685191H1	1878368H1	2601563F6	2753227H1	3051974H1	5884565H1	1293671H1	1878368H1	2601563F6	2753227H1	3051974H1	5884565H1	6060785H1
PID		7511620	7511620	7511620	7506995	7506995	7506995	7506995	7506995	750696	7506996	7506996	7506996	9669052	7506996	7506996
SEQ	a ë	55	20	50	Г	5.			İ	53	52	52	53	3 6	53	52

What is claimed is:

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- 1. An isolated polypeptide selected from the group consisting of:
- a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26,
- b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-8, 11, 14-18, 20, 22-23, and 25-26,
- a polypeptide comprising a naturally occurring amino acid sequence at least 93% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:9 and SEQ ID NO:21,
- a polypeptide comprising a naturally occurring amino acid sequence at least 96%
 identical to the amino acid sequence of SEQ ID NO:24,
- e) a polypeptide comprising a naturally occurring amino acid sequence at least 99% identical to the amino acid sequence of SEQ ID NO:19,
- f) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and
- g) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26.
- 2. An isolated polypeptide of claim 1 comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26.
 - 3. An isolated polynucleotide encoding a polypeptide of claim 1.
 - 4. An isolated polynucleotide encoding a polypeptide of claim 2.
- 5. An isolated polynucleotide of claim 4 comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52.
- 6. A recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide of claim 3.

- 7. A cell transformed with a recombinant polynucleotide of claim 6.
- 8. A transgenic organism comprising a recombinant polynucleotide of claim 6.
- 9. A method of producing a polypeptide of claim 1, the method comprising:
 - a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide, and said recombinant polynucleotide comprises a promoter sequence operably linked to a polynucleotide encoding the polypeptide of claim 1, and
 - b) recovering the polypeptide so expressed.

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- 10. A method of claim 9, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-26.
- 15 11. An isolated antibody which specifically binds to a polypeptide of claim 1.
 - 12. An isolated polynucleotide selected from the group consisting of:
 - a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52,
 - b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-44, SEQ ID NO:46-47, SEQ ID NO:49, and SEQ ID NO:51-52,
 - c) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 92% identical to the polynucleotide sequence of SEQ ID NO:48,
 - d) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 99% identical to the polynucleotide sequence of SEQ ID NO:45,
 - a polynucleotide consisting essentially of a naturally occurring polynucleotide
 sequence at least 90% identical to the polynucleotide sequence of SEQ ID NO:50,
 - d) a polynucleotide complementary to a polynucleotide of a),
 - e) a polynucleotide complementary to a polynucleotide of b),
 - f) a polynucleotide complementary to a polynucleotide of c),
 - g) a polynucleotide complementary to a polynucleotide of d),

h) a polynucleotide complementary to a polynucleotide of e), and

i) an RNA equivalent of a)-h).

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- 13. An isolated polynucleotide comprising at least 60 contiguous nucleotides of apolynucleotide of claim 12.
 - 14. A method of detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 12, the method comprising:
 - a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and
 - b) detecting the presence or absence of said hybridization complex, and, optionally, if present, the amount thereof.
 - 15. A method of claim 14, wherein the probe comprises at least 60 contiguous nucleotides.
- 16. A method of detecting a target polynucleotide in a sample, said target polynucleotide
 20 having a sequence of a polynucleotide of claim 12, the method comprising:
 - a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and
 - b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.
 - 17. A composition comprising a polypeptide of claim 1 and a pharmaceutically acceptable excipient.
- 18. A composition of claim 17, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-26.
 - 19. A method for treating a disease or condition associated with decreased expression of

functional TRICH, comprising administering to a patient in need of such treatment the composition of claim 17.

- 20. A method of screening a compound for effectiveness as an agonist of a polypeptide of claim 1, the method comprising:
 - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
 - b) detecting agonist activity in the sample.
- 21. A composition comprising an agonist compound identified by a method of claim 20 and a pharmaceutically acceptable excipient.
 - 22. A method for treating a disease or condition associated with decreased expression of functional TRICH, comprising administering to a patient in need of such treatment a composition of claim 21.

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- 23. A method of screening a compound for effectiveness as an antagonist of a polypeptide of claim 1, the method comprising:
 - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
 - b) detecting antagonist activity in the sample.

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- 24. A composition comprising an antagonist compound identified by a method of claim 23 and a pharmaceutically acceptable excipient.
- 25. A method for treating a disease or condition associated with overexpression of functional TRICH, comprising administering to a patient in need of such treatment a composition of claim 24.
 - 26. A method of screening for a compound that specifically binds to the polypeptide of claim 1, the method comprising:
 - a) combining the polypeptide of claim 1 with at least one test compound under suitable conditions, and
 - b) detecting binding of the polypeptide of claim 1 to the test compound, thereby identifying a compound that specifically binds to the polypeptide of claim 1.

27. A method of screening for a compound that modulates the activity of the polypeptide of claim 1, the method comprising:

- a) combining the polypeptide of claim 1 with at least one test compound under conditions permissive for the activity of the polypeptide of claim 1,
- b) assessing the activity of the polypeptide of claim 1 in the presence of the test compound, and

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- c) comparing the activity of the polypeptide of claim 1 in the presence of the test compound with the activity of the polypeptide of claim 1 in the absence of the test compound, wherein a change in the activity of the polypeptide of claim 1 in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide of claim 1.
- 28. A method of screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a sequence of claim 5, the method comprising:
 - exposing a sample comprising the target polynucleotide to a compound, under conditions suitable for the expression of the target polynucleotide,
 - b) detecting altered expression of the target polynucleotide, and
 - c) comparing the expression of the target polynucleotide in the presence of varying amounts of the compound and in the absence of the compound.
 - 29. A method of assessing toxicity of a test compound, the method comprising:
 - a) treating a biological sample containing nucleic acids with the test compound,
 - b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide of claim 12 under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide comprising a polynucleotide sequence of a polynucleotide of claim 12 or fragment thereof,
 - c) quantifying the amount of hybridization complex, and
- d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is

indicative of toxicity of the test compound.

30. A diagnostic test for a condition or disease associated with the expression of TRICH in a biological sample, the method comprising:

- a) combining the biological sample with an antibody of claim 11, under conditions suitable for the antibody to bind the polypeptide and form an antibody:polypeptide complex, and
- b) detecting the complex, wherein the presence of the complex correlates with the presence of the polypeptide in the biological sample.

31. The antibody of claim 11, wherein the antibody is:

- a) a chimeric antibody,
- b) a single chain antibody,
- c) a Fab fragment,

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- d) a F(ab')₂ fragment, or
- e) a humanized antibody.
- 32. A composition comprising an antibody of claim 11 and an acceptable excipient.
- 33. A method of diagnosing a condition or disease associated with the expression of TRICH in a subject, comprising administering to said subject an effective amount of the composition of claim 32.
 - 34. A composition of claim 32, wherein the antibody is labeled.
 - 35. A method of diagnosing a condition or disease associated with the expression of TRICH in a subject, comprising administering to said subject an effective amount of the composition of claim 34.
- 36. A method of preparing a polyclonal antibody with the specificity of the antibody of claim 11, the method comprising:
 - a) immunizing an animal with a polypeptide consisting of an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, or an immunogenic fragment

- thereof, under conditions to elicit an antibody response,
- b) isolating antibodies from the animal, and

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- c) screening the isolated antibodies with the polypeptide, thereby identifying a polyclonal antibody which specifically binds to a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26.
- 37. A polyclonal antibody produced by a method of claim 36.
- 38. A composition comprising the polyclonal antibody of claim 37 and a suitable carrier.
- 39. A method of making a monoclonal antibody with the specificity of the antibody of claim 11, the method comprising:
 - a) immunizing an animal with a polypeptide consisting of an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, or an immunogenic fragment thereof, under conditions to elicit an antibody response,
 - b) isolating antibody producing cells from the animal,
 - c) fusing the antibody producing cells with immortalized cells to form monoclonal antibody-producing hybridoma cells,
 - d) culturing the hybridoma cells, and
 - e) isolating from the culture monoclonal antibody which specifically binds to a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26.
 - 40. A monoclonal antibody produced by a method of claim 39.
 - 41. A composition comprising the monoclonal antibody of claim 40 and a suitable carrier.
- 42. The antibody of claim 11, wherein the antibody is produced by screening a Fab expression library.
- 43. The antibody of claim 11, wherein the antibody is produced by screening a recombinant immunoglobulin library.

44. A method of detecting a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26 in a sample, the method comprising:

- a) incubating the antibody of claim 11 with a sample under conditions to allow specific
 binding of the antibody and the polypeptide, and
- b) detecting specific binding, wherein specific binding indicates the presence of a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26 in the sample.
- 45. A method of purifying a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26 from a sample, the method comprising:
 - a) incubating the antibody of claim 11 with a sample under conditions to allow specific binding of the antibody and the polypeptide, and
 - separating the antibody from the sample and obtaining the purified polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26.
 - 46. A microarray wherein at least one element of the microarray is a polynucleotide of claim 13.
- 47. A method of generating an expression profile of a sample which contains polynucleotides, the method comprising:
 - a) labeling the polynucleotides of the sample,

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- b) contacting the elements of the microarray of claim 46 with the labeled polynucleotides of the sample under conditions suitable for the formation of a hybridization complex, and
- c) quantifying the expression of the polynucleotides in the sample.
- 48. An array comprising different nucleotide molecules affixed in distinct physical locations on a solid substrate, wherein at least one of said nucleotide molecules comprises a first oligonucleotide or polynucleotide sequence specifically hybridizable with at least 30 contiguous nucleotides of a target polynucleotide, and wherein said target polynucleotide is a polynucleotide of claim 12.
 - 49. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is

completely complementary to at least 30 contiguous nucleotides of said target polynucleotide.

50. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to at least 60 contiguous nucleotides of said target polynucleotide.

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- 51. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to said target polynucleotide.
 - 52. An array of claim 48, which is a microarray.

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- 53. An array of claim 48, further comprising said target polynucleotide hybridized to a nucleotide molecule comprising said first oligonucleotide or polynucleotide sequence.
- 54. An array of claim 48, wherein a linker joins at least one of said nucleotide molecules to said solid substrate.
 - 55. An array of claim 48, wherein each distinct physical location on the substrate contains multiple nucleotide molecules, and the multiple nucleotide molecules at any single distinct physical location have the same sequence, and each distinct physical location on the substrate contains nucleotide molecules having a sequence which differs from the sequence of nucleotide molecules at another distinct physical location on the substrate.
- 56. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:1.

- 57. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:2.
- 58. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:3.
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- 59. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:4.
- 60. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:5.

61. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:6. 62. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:7. 5 63. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:8. 64. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:9. 65. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:10. 10 66. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:11. 67. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:12. 15 68. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:13. 69. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:14. 70. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:15. 20 71. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:16. 72. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:17. 25 73. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:18. 74. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:19. 75. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:20. 30 76. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:21. 77. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:22.

78. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:23. 79. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:24. 80. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:25. 5 81. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:26. 82. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:27. 10 83. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEO ID NO:28. 84. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEO ID NO:29. 85. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:30. 15 86. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:31. 87. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:32. 20 88. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:33. 89. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:34. 90. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:35. 25 91. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:36. 92. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:37. 30 93. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:38. 94. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:39.

95. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:40.

- 96. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:41.
- 5 97. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:42.
 - 98. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:43.
 - 99. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:44.

100. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:45.

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- 101. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:46.
 - 102. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:47.
- 20 103. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:48.
 - 104. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:49.
 - 105. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:50.
- 106. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:51.
 - 107. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:52.

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Gln	Arg	G1n	Glu	Asp 740	Glu	Glu	·Asp	Lys		Arg	Gln	Va1	Glu	
His	Gln	Glu	Pro	Gly 755	Ala	Ala	Va1	Pro		Gly	Gln	Glu	Ala	
Glu	Gly	Lys	Ala	Arg 770	Glu	Thr	Val	Glu		Leu	Pro	Pro	Leu	
Leu	qaA	Pro	Val	Leu 785	Arg	Ala	Pro	Gly		Arg	Pro	Ala	Pro	
Gln	Asp	Leu	Asn	Gln 800	Arg	Ser	Leu	Glu		Ser	Glu	Gly	Pro	
Gly	Arg	Asp	Pro	Ala 815	Gly	Pro	Pro	Asp		Gly	Pro	Asp	Thr	
Pro	Arg	Ala	Ala	Gln 830	Ala	Lys	Leu	Arg		Gly	Gln	Lys	Asp	
Ala	Pro	Arg	Ala	Ala 845	Gly	Thr	Val	Lys		Leu	Pro	Lys	Gly	
Glu	Gln	Val	Pro	Val 860	Pro	Asp	Pro	Ala		G1u	Ala	Gly	Gly	
Glu	G1u	Arg	Leu	Ala 875	Glu	Glu	Phe	Pro		Gln	Ser	Gln	Asp	
Thr	Gly	Gly	Ser	Gln 890	Asp	Arg	Lys	Lys		Gly	Lys	Glu	Va1	
Ala	Thr	Gly	Thr	Ser 905	Ile	Leu	Lys	Glu	Ala 910	Asn	Trp	Leu	Val	Ala 915
Gly	Pro	Gly	Ala	Glu 920	Thr	Gly	Asp	Pro	Arg 925	Met	Lys	Pro	Lys	Gln 930
Val	Ser	Arg	qaA	Leu 935	Gly	Leu	Ala		Asp 940	Leu	Pro	Gly	Gly	Ala 945
Glu	Gly	Ala	Ala	Ala 950	Gln	Pro	Gln	Ala	Val 955	Leu	Arg	Gln	Pro	Glu 960
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Gly	Asp	His	Val	Pro 995	Va1	Ser	His		Gln 1000	Pro	Arg	Gly		Glu 1005
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Gly	Leu	Lys	Arg	Ala 1025	Val	Pro	G1y	Gly		Arg	Pro	Asp		Ala 1035
Lys	Pro	Asn	Arg	Asp 1040	Leu	Lys	Leu	Gln		G1y	Ser	Asp	Leu	
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Gln Val Asn Asp Leu Arg Gly Ala Leu Asp Ala Gln Leu Arg Gln
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Asp Gly Phe Cys Glu Leu Phe Gly Tyr Ser Arg Val Glu Val Met
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Ser Ser Ala Val Ser Arg Leu Ala Gln Ala Leu Leu Gly Ala Glu
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Phe Arg Cys Leu Val Asp Val Val Pro Val Lys Asn Glu Asp Gly
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Ile Pro Gln Phe Thr Leu Asn Phe Val Glu Phe Asn Leu Glu Lys
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His Arg Ser Ser Ser Thr Thr Glu Ile Glu Ile Ile Ala Pro His
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Lys Val Val Glu Arg Thr Gln Asn Val Thr Glu Lys Val Thr Gln
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Val Leu Ser Leu Gly Ala Asp Val Leu Pro Glu Tyr Lys Leu Gln
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Ala Pro Arg Ile His Arg Trp Thr Ile Leu His Tyr Ser Pro Phe
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Ala Val Phe Thr Pro Tyr Ser Ala Ala Phe Leu Leu Ser Asp Gln
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Val	Ile	Asn	Phe	Arg 320	Thr	Thr	Tyr	Val	Asn 325	Thr	Asn	Asp	Glu	Val 330
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Cys	Thr	Phe	Ala	Leu 410	Ile	Ala	His	Trp	Leu 415	Ala	Cys	Ile	Trp	Tyr 420
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			Glu	575					580					585
			Leu	590					595					600
Gly	Cys	Leu	Arg	Ala 605	Leu	Ala	Val	Lys	Phe 610	Lys	Thr	Thr	His	Ala 615
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			Ile	635					640					645
			Ala	650					655					660
			His	665					670					675
			Tyr	680					685					690
Leu	Glu	Val	Leu	Asp	Met	Tyr	Pro	Ala	Phe	Ala	Glu	Ser	Phe	Trp

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Ser Lys Leu Glu Val Thr Phe Asn Leu Arg Asp Ala Pro Gly Ser
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Gln Asp His Gln Gly Phe Phe Leu Ser Asp Asn Gln Ser Asp Ala
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Leu Gln Glu Met Pro Pro Arg His Ser Pro Gln Ser Pro Gln
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Glu Asp Pro Asp Cys Trp Pro Leu Lys Leu Gly Ser Arg Leu Glu
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Gly His Ala Ser Tyr Ile Leu Glu Ala Pro Ala Ser Asn Asp Leu
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Leu Pro Gln Gly Phe Leu Pro Pro Ala Gln Thr Pro Ser Tyr Gly
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Leu Leu Gly Gly Gly Phe Ala Leu Ala Lys Gly Ser Glu Ala Ser
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Gly Leu Ser Val Trp Met Gly Lys Gln Met Glu Pro Leu His Ala
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Leu Tyr Ile Met Leu Pro Cys Thr Leu Ser Ala Ser Phe Ala Phe
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Leu Cys Tyr Lys Asn Gly Gly Glu His Cys Met Glu Phe Gln Lys
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Thr Asn Gly Ser Leu Asn Gly Thr Ser Glu Asn Ala Thr Ser Pro
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Val Ile Glu Phe Trp Glu Arg Arg Val Leu Lys Ile Ser Asp Gly
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Ala	Ala	Gln	Gly	Ile 170	Gln	Phe	Tyr	Leu	Tyr 175	Pro	Asn	Leu	Thr	Arg 180
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				200					205				Gly	210
				215					220				Ala	225
				230					235				Ala	240
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				260					265				Ala Ala	270
				275					280				Gln	285
				290					295				Tyr	300
				305					310				Leu	315
				320					325				Thr	330
G1y	Gly	Met	Tyr	335 Val	Phe	Gln	Leu	Phe	340 Asp	Tyr	Tyr	Ala	Ala	345 Ser
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Ala	Trans.			365			ATG	IIe	Phe	Glu	ser	Leu	Cys	V CA.
3.6 - 1-	TTD	Val	Tyr		Ala				370				Cys	375
мес				Gly 380 Arg		Lys	Arg	Phe	370 Tyr 385	Asp	Asn	Ile		375 Asp 390
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Phe	Ile Leu	Gly Thr	Tyr Pro	Gly 380 Arg 395 Ala 410 Pro	Pro Val	Lys Trp Cys	Arg Pro Thr	Phe Leu Ala	370 Tyr 385 Ile 400 Thr 415 Lys	Asp Lys Phe	Asn Tyr Leu	Ile Cys Phe	Glu Trp	375 Asp 390 Leu 405 Leu 420 Pro
Phe Ile	Ile Leu Lys	Gly Thr Tyr	Tyr Pro Thr	Gly 380 Arg 395 Ala 410 Pro 425 Ala	Pro Val Leu	Lys Trp Cys Thr	Arg Pro Thr Tyr	Phe Leu Ala Asn	370 Tyr 385 Ile 400 Thr 415 Lys 430 Leu	Asp Lys Phe Lys	Asn Tyr Leu Tyr	Ile Cys Phe Thr	Glu Trp Ser	375 Asp 390 Leu 405 Leu 420 Pro 435 Met
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Phe Ile Trp Val	Ile Leu Lys Trp Cys	Gly Thr Tyr Gly Ile	Tyr Pro Thr Asp Pro	Gly 380 Arg 395 Ala 410 Pro 425 Ala 440 Ala 455	Pro Val Leu Leu Trp	Lys Trp Cys Thr Gly Ser	Arg Pro Thr Tyr Trp Leu	Phe Leu Ala Asn Leu Tyr	370 Tyr 385 Ile 400 Thr 415 Lys 430 Leu 445 Arg 460	Asp Lys Phe Lys Ala Leu	Asn Tyr Leu Tyr Leu Gly	Ile Cys Phe Thr Ser	Glu Trp Ser Tyr	375 Asp 390 Leu 405 Leu 420 Pro 435 Met 450 Lys 465 Glu
Phe Ile Trp Val Gly	Ile Leu Lys Trp Cys Pro	Gly Thr Tyr Gly Ile Phe	Tyr Pro Thr Asp Pro	Gly 380 Arg 395 Ala 410 Pro 425 Ala 440 Ala 455 Glu 470	Pro Val Leu Leu Trp Arg	Lys Trp Cys Thr Gly Ser	Arg Pro Thr Tyr Trp Leu Arg	Phe Leu Ala Asn Leu Tyr	370 Tyr 385 Ile 400 Thr 415 Lys 430 Leu 445 Arg 460 Leu 475	Asp Lys Phe Lys Ala Leu Met	Asn Tyr Leu Tyr Leu Gly Cys	Ile Cys Phe Thr Ser Thr	Glu Trp Ser Tyr Ser Leu	375 Asp 390 Leu 405 Leu 420 Pro 435 Met 450 Lys 465 Glu 480

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Gln Ser Phe Leu Gly Ser Glu Gly Ser His Gly Arg Pro Gly Gly
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Pro Gly Pro Gly Thr Gly Arg Gly Lys Tyr Arg Thr Ile Ser Gln
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Ile Pro Gln Phe Thr Leu Asn Phe Val Glu Phe Asn Leu Glu Lys
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His Arg Ser Ser Ser Thr Thr Glu Ile Glu Ile Ile Ala Pro His
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Lys Val Val Glu Arg Thr Gln Asn Val Thr Glu Lys Val Thr Gln
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                                   220
Val Leu Ser Leu Gly Ala Asp Val Leu Pro Glu Tyr Lys Leu Gln
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Ala Pro Arg Ile His Arg Trp Thr Ile Leu His Tyr Ser Pro Phe
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Lys Ala Val Trp Asp Trp Leu Ile Pro Leu Leu Val Ile Tyr Thr
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Ala Val Phe Thr Pro Tyr Ser Ala Ala Phe Leu Leu Ser Asp Gln
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Asp Glu Ser Arg Arg Gly Ala Cys Ser Tyr Thr Cys Ser Pro Leu
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Val Ile Asn Phe Arg Thr Thr Tyr Val Asn Thr Asn Asp Glu Val
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Val Ser His Pro Arg Arg Ile Ala Val His Tyr Phe Lys Gly Trp
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Phe Leu Ile Asp Met Val Ala Ala Ile Pro Phe Asp Leu Leu Ile
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			Arg	380					385					390
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Leu	Thr	Ser	Val	Gly 425	Phe	G1y	Asn	Val	Ser 430	Pro	Asn	Thr	Asn	Ser 435
G1u	Lys	Va1	Phe	Ser 440	Ile	Cys	Val	Met	Leu 445	Ile	Gly	Ser	Leu	Met 450
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Leu Trp Pro Pro Leu Ala Ser Pro Leu His Pro Leu Glu Val Gln
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Arg Ser Ser Arg Pro Ile Arg Trp Trp Ser Gly His Arg Thr Ser
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Leu Arg Arg Ser Pro Arg Ser Cys Pro Trp Ala Arg Met Cys Cys
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Arg Ser Thr Ser Cys Arg Arg Arg Ala Ser Thr Ala Gly Pro Ser
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Cys Thr Thr Ala Pro Phe Lys Ala Val Trp Asp Trp Leu Ile Leu
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Tyr Thr Cys Ser Pro Leu Thr Val Val Asp Leu Ile Val Asp Ile
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Asn Thr Asn Asp Glu Val Val Ser His Pro Arg Arg Ile Ala Val
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Leu	Glu	His	Lys	Ile 275	Gly	Trp	Leu	Asp	Ser 280	Leu	G1y	Val	Gln	Leu 285
Gly	Lys	Arg	Tyr	Asn 290	Gly	Ser	Asp	Pro	Ala 295	Ser	Gly	Pro	Ser	Val 300
Gln	Asp	Lys	Tyr	Val 305	Thr	Ala	Leu	Tyr	Phe 310	Thr	Phe	Ser	Ser	Leu 315
Thr	Ser	Val	Gly	Phe 320	Gly	Asn	Val	Ser	Pro 325	Asn	Thr	Asn	Ser	Glu 330
Lys	Val	Phe	Ser	I1e 335	Сув	Val	Met	Leu	Ile 340	Gly	Ser	Leu	Met	Tyr 345
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			His	425					430					435
			Ala	440					445					450
			Thr	455					460					465
			Leu	470					475					480
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			Ser	545					550					555
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			Ser	575					580					585
			Trp	590					595					600
			Ser	605					610					615
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Pro Ala Ser Asn Asp Leu Ala Leu Val Pro Ile Ala Ser Glu Thr
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Thr Ser Pro Gly Pro Arg Leu Pro Gln Gly Phe Leu Pro Pro Ala
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                                   685
Gln Thr Pro Ser Tyr Gly Asp Leu Asp Asp Cys Ser Pro Lys His
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                695
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                710
Lys Thr Leu Ala Pro Ser Ser Glu Gln Glu Gln Pro Glu Gly Leu
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Trp Pro Pro Leu Ala Ser Pro Leu His Pro Leu Glu Val Gln Gly
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Leu Ile Cys Gly Pro Cys Phe Ser Ser Leu Pro Glu His Leu Gly
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Lys Val Ala Glu Asp Glu Ala Glu Ala Ala Ala Ala Lys Phe
Thr Gly Leu Ser Lys Glu Glu Leu Leu Lys Val Ala Gly Ser Pro
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Gly Trp Val Arg Thr Arg Trp Ala Leu Leu Leu Phe Trp Leu
Gly Trp Leu Gly Met Leu Ala Gly Ala Val Val Ile Ile Val Arg
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Ala Pro Arg Cys Arg Glu Leu Pro Ala Gln Lys Trp Trp His Thr
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Gly Ala Leu Tyr Arg Ile Gly Asp Leu Gln Ala Phe Gln Gly His
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Gly Ala Gly Asn Leu Ala Gly Leu Lys Gly Arg Leu Asp Tyr Leu
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Ser Ser Leu Lys Val Lys Gly Leu Val Leu Gly Pro Ile His Lys
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Ala Lys Lys Ser Ile Arg Val Ile Leu Asp Leu Thr Pro Asn
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Tyr Arg Gly Glu Asn Ser Trp Phe Ser Thr Gln Val Asp Thr Val
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                                   220
Ala Thr Lys Val Lys Asp Ala Leu Glu Phe Trp Leu Gln Ala Gly
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                                   235
Val Asp Gly Phe Gln Val Arg Asp Ile Glu Asn Leu Lys Asp Ala
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               245
Ser Ser Phe Leu Ala Glu Trp Gln Asn Ile Thr Lys Gly Phe Ser
               260
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Glu Asp Arg Leu Leu Ile Ala Gly Thr Asn Ser Ser Asp Leu Gln
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               275
Gln Ile Leu Ser Leu Leu Glu Ser Asn Lys Asp Leu Leu Leu Thr
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Ser Ser Tyr Leu Ser Asp Ser Gly Ser Thr Gly Glu His Thr Lys
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Ser Leu Val Thr Gln Tyr Leu Asn Ala Thr Gly Asn Arg Trp Cys
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Ser Trp Ser Leu Ser Gln Ala Arg Leu Leu Thr Ser Phe Leu Pro
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Ala Gln Leu Leu Arg Leu Tyr Gln Leu Met Leu Phe Thr Leu Pro
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Gly Thr Pro Val Phe Ser Tyr Gly Asp Glu Ile Gly Leu Asp Ala
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Ala Ala Leu Pro Gly Gln Gly Gln Ser Glu Asp Pro Gly Ser Leu
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Leu Ser Leu Phe Arg Arg Leu Ser Asp Gln Arg Ser Lys Glu Arg
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Val Val Leu Asn Phe Gly Asp Val Gly Leu Ser Ala Gly Leu Gin
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Met	Glu	Arg	Pro	Leu 35	Leu	Asn	Gln	Glu	His 40	Leu	Glu	Glu	Leu	Gly 45
Arg	Trp	Gly	Ser	Ala 50	Pro	Arg	Thr	His	Gln 55	Trp	Arg	Thr	Trp	Leu 60
G1n	Cys	Ser	Arg	Ala 65	Arg	Ala	Tyr	Ala	Leu 70	Leu	Leu	Gln	His	Leu 75
Pro	Val	Leu	Val	Trp 80	Leu	Pro	Arg	Tyr	Pro 85	Val	Arg	Asp	Trp	Leu 90
Leu	Gly	Asp	Leu		Ser	Gly	Leu	Ser		Ala	Ile	Met	Gln	
Pro	Gln	Gly	Leu		Tyr	Ala	Leu	Leu		Gly	Leu	Pro	Pro	
Phe	Gly	Leu	Tyr		Ser	Phe	Tyr	Pro		Phe	Ile	Tyr	Phe	
Phe	Gly	Thr	Ser		His	Ile	Ser	Val		Thr	Phe	Ala	Val	
Ser	Val	Met	Val		Ser	Va1	Thr	G1u		Leu	Ala	Pro	Gln	
Leu	Asn	Asp	Ser		Ile	Asn	Glu	Thr		Arg	Asp	Ala	Ala	
Val	Gln	Va1	Ala		Thr	Leu	Ser	Va1		Val	Gly	Leu	Phe	
Val	Gly	Leu	Gly		Ile	His	Phe	Gly	Phe	Val	Va1	Thr	Tyr	
Ser	Glu	Pro	Leu		Arg	Gly	Tyr	Thr		Ala	Ala	Ala	Val	
Val	Phe	Val	Ser		Leu	Lys	Tyr	Val		Gly	Leu	His	Leu	
Ser	His	Ser	Gly		Leu	Ser	Leu	Ile		Thr	Val	Leu	Glu	
Сув	Trp	Lys	Leu		Gln	Ser	Lys	Leu		Gly	Ala	Thr	Gly	
Ser	Tyr	Gly	Met		Leu	Lys	His	Arg		Glu	Val	Asp	Val	
Gly	Asn	Ile	Pro		Gly	Leu	Val	Pro		Va1	Ala	Pro	Asn	Thr 300
Gln	Leu	Phe	Ser		Leu	Val	Gly	Ser	Ala 310	Phe	Thr	Ile	Ala	Val 315
Va1	G1 _Y	Phe	Ala	Ile 320	Ala	Ile	Ser	Leu	Gly 325	Lys	Ile	Phe	Ala	Leu 330
Arg	His	Gly	Tyr	Arg 335	Val	Asp	Ser	Asn	Gln 340	Glu	Leu	Va1	Ala	Leu 345
Gly	Leu	Ser	Asn	Leu 350	Ile	Gly	Gly	Ile	Phe 355	Gln	Cys	Phe	Pro	Val 360
Ser	Суз	Ser	Met	Ser 365	Arg	Ser	Leu	Val	Gln 370	Glu	Ser	Thr	Gly	Gly 375
Asn	Ser	Gln	Val	Ala 380	Gly	Ala	Ile	Ser	Ser 385	Leu	Phe	Ile	Leu	Leu 390
Ile	Ile	Val	Lys		Gly	Glu	Leu	Phe		Asp	Leu	Pro	Lys	
Val	Leu	Ala	Ala		Ile	Ile	Val	Asn	Leu 415	Lys	Gly	Met	Leu	Arg 420
Gln	Leu	Ser	qaA	Met	Arg	Ser	Leu	Trp	Lys	Ala	Asn	Arg	Ala	Asp

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Ala Lys Glu Val Arg Gly Val Lys Val Phe Arg Ser Ser Ala Thr
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Val Tyr Phe Ala Asn Ala Glu Phe Tyr Ser Asp Ala Leu Lys Gln
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Arg Cys Gly Val Asp Val Asp Phe Leu Ile Ser Gln Lys Lys
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Leu Lys Lys Gln Glu Gln Leu Lys Leu Lys Gln Leu Gln Lys
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Glu Glu Lys Leu Arg Lys Gln Ala Ala Ser Pro Lys Gly Ala Ser
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Val Ser Ile Asn Val Asn Thr Ser Leu Glu Asp Met Arg Ser Asn
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Asp Gly Ser Thr Leu Lys Ala Leu Gly Leu Pro Gln Pro Asp Phe
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His Ser Leu Ile Leu Asp Leu Gly Ala Leu Ser Phe Val Asp Thr
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Val Cys Leu Lys Ser Leu Lys Asn Ile Phe His Asp Phe Arg Glu
                                    655
Ile Glu Val Glu Val Tyr Met Ala Ala Cys His Ser Pro Val Val
                                    670
Ser Gln Leu Glu Ala Gly His Phe Phe Asp Ala Ser Ile Thr Lys
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Leu	Leu	Tyr	Trp	Met 50	Pro	G1u	Trp	Arg	Va1 55	Lys	Ala	Thr	Cys	Va1 60
Arg	Ala	Ala	Ile	Lys 65	Asp	Cys	Glu	Val	Val 70	Leu	Leu	Arg	Thr	Thr 75
Asp	Glu	Phe	Lys	Met 80	Trp	Phe	Cys	Ala	Lys 85	Ile	Arg	Va1	Leu	Ser 90
Leu	Glu	Thr	Tyr	Pro 95	Val	Ser	Ser	Pro	Lys 100	Ser	Met	Ser	Asn	Lys 105
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				Phe 140					145					150
Asp	Thr	Ile	His	Asn 155					160					165
			Thr	170					175				Leu	180
				Ala 185					190					195
	•			Val 200					205					210
				Phe 215					220					225
				Glu 230					235					240
				Ser 245					250	_				255
				Leu 260					265					270
				Cys 275					280					285
		-		Val 290			-		295					300
				Pro 305					310					315
				Ser 320					325					330
				Pro 335					340					345
				Asn 350					355					360
				Val 365					370					375
				Val 380					385					390
				Ser 395					400					405
				Ala 410					415					420
				Phe 425					430					435
Glu	Val	Gln	Val	Gly 440	Val	Ile	Ile	Ile	Glu 445	Ser	Leu	Asp	Ile	Ile 450

Thr	Ile	Thr	Val	Pro 455	Pro	Ala	Leu	Pro	Ala 460	Ala	Met	Thr	Ala	Gly 465
Ile	Val	Tyr	Ala	Gln 470	Arg	Arg	Leu	ГЛа	Lys 475	Ile	G1y	Ile	Phe	Cys 480
Ile	Ser	Pro	Gln	Arg 485	Ile	Asn	Ile	Сув	Gly 490	Gln	Leu	Asn	Leu	Va1 495
Cys	Phe	Asp	Lys	Thr 500	Gly	Thr	Leu	Thr	G1u 505	Asp	Gly	Leu	Asp	Leu 510
Trp	Gly	Ile	Gln	Arg 515	Val	Glu	Asn	Ala	Arg 520	Phe	Leu	Ser	Pro	Glu 525
			Cys	530					535					540
_			Thr	545					550			_		555
			Pro	560					565					570
			Glu	575					580					585
			Thr	590					595					600
			Ala	605					610					615
	•	_	Glu	620	_				625					630
			Arg	635					640					645
			Ala	650					655					660
			Tree	O7	mb	T 7 7	Dage	T r 7	7	Dh a	~1 ~	7	T 7 7	T
				665					Asp 670					675
Glu	Asp	Phe	Thr	665 Lys 680	Gln	Gly	Phe	Arg	670 Val 685	Ile	Ala	Leu	Ala	675 His 690
Glu Arg	Asp Lys	Phe Leu	Thr Glu	665 Lys 680 Ser 695	Gln Lys	Gly Leu	Phe Thr	Arg Trp	670 Val 685 His 700	Ile Lys	Ala Val	Leu Gln	Ala Asn	675 His 690 Ile 705
Glu Arg Ser	Asp Lys Arg	Phe Leu Asp	Thr Glu Ala	665 Lys 680 Ser 695 Ile 710	Gln Lys Glu	Gly Leu Asn	Phe Thr Asn	Arg Trp Met	670 Val 685 His 700 Asp 715	Ile Lys Phe	Ala Val Met	Leu Gln Gly	Ala Asn Leu	675 His 690 Ile 705 Ile 720
Glu Arg Ser Ile	Asp Lys Arg Met	Phe Leu Asp Gln	Thr Glu Ala Asn	665 Lys 680 Ser 695 Ile 710 Lys 725	Gln Lys Glu Leu	Gly Leu Asn Lys	Phe Thr Asn Gln	Arg Trp Met Glu	670 Val 685 His 700 Asp 715 Thr	Ile Lys Phe Pro	Ala Val Met Ala	Leu Gln Gly Val	Ala Asn Leu Leu	675 His 690 Ile 705 Ile 720 Glu 735
Glu Arg Ser Ile Asp	Asp Lys Arg Met Leu	Phe Leu Asp Gln His	Thr Glu Ala Asn Lys	665 Lys 680 Ser 695 Ile 710 Lys 725 Ala 740	Gln Lys Glu Leu Asn	Gly Leu Asn Lys Ile	Phe Thr Asn Gln Arg	Arg Trp Met Glu Thr	670 Val 685 His 700 Asp 715 Thr 730 Val 745	Ile Lys Phe Pro Met	Ala Val Met Ala Val	Leu Gln Gly Val Thr	Ala Asn Leu Leu Gly	675 His 690 Ile 705 Ile 720 Glu 735 Asp 750
Glu Arg Ser Ile Asp Ser	Asp Lys Arg Met Leu Met	Phe Leu Asp Gln His	Thr Glu Ala Asn Lys Thr	665 Lys 680 Ser 695 Ile 710 Lys 725 Ala 740 Ala 755	Gln Lys Glu Leu Asn Val	Gly Leu Asn Lys Ile Ser	Phe Thr Asn Gln Arg Val	Arg Trp Met Glu Thr	670 Val 685 His 700 Asp 715 Thr 730 Val 745 Arg 760	Ile Lys Phe Pro Met	Ala Val Ala Val Cys	Leu Gln Gly Val Thr	Ala Asn Leu Leu Gly Met	675 His 690 Ile 705 Ile 720 Glu 735 Asp 750 Ile 765
Glu Arg Ser Ile Asp Ser Leu	Asp Lys Arg Met Leu Met	Phe Leu Asp Gln His Leu Gln	Thr Glu Ala Asn Lys	665 Lys 680 Ser 695 Ile 710 Lys 725 Ala 740 Ala 755 Lys 770	Gln Lys Glu Leu Asn Val	Gly Leu Asn Lys Ile Ser Ile	Phe Thr Asn Gln Arg Val Ile	Arg Trp Met Glu Thr Ala Ala	670 Val 685 His 700 Asp 715 Thr 730 Val 745 Arg 760 Glu 775	Ile Lys Phe Pro Met Asp	Ala Val Ala Val Cys Leu	Leu Gln Gly Val Thr Gly Pro	Ala Asn Leu Leu Gly Met Pro	675 His 690 Ile 705 Ile 720 Glu 735 Asp 750 Ile 765 Lys 780
Glu Arg Ser Ile Asp Ser Leu Asp	Asp Lys Arg Met Leu Met Pro	Phe Leu Asp Gln His Leu Gln Lys	Thr Glu Ala Asn Lys Thr	665 Lys 680 Ser 695 Ile 710 Lys 725 Ala 740 Ala 755 Lys 770 Ala 785	Gln Lys Glu Leu Asn Val Val	Gly Leu Asn Lys Ile Ser Ile Ile	Phe Thr Asn Gln Arg Val Ile Asn	Arg Trp Met Glu Thr Ala Ala Trp	670 Val 685 His 700 Asp 715 Thr 730 Val 745 Arg 760 Glu 775 His 790	Ile Lys Phe Pro Met Asp Ala Tyr	Ala Val Ala Val Cys Leu Ala	Leu Gln Gly Val Thr Gly Pro Asp	Ala Asn Leu Leu Gly Met Pro Ser	675 His 690 Ile 705 Ile 720 Glu 735 Asp 750 Ile 765 Lys 780 Leu 795
Glu Arg Ser Ile Asp Ser Leu Asp	Asp Lys Arg Met Leu Met Pro Gly Gln	Phe Leu Asp Gln His Leu Gln Lys	Thr Glu Ala Asn Lys Thr Asp Val	665 Lys 680 Ser 695 Ile 710 Lys 725 Ala 740 Ala 755 Lys 770 Ala 785 His 800	Gln Lys Glu Leu Asn Val Val Lys Pro	Gly Leu Asn Lys Ile Ser Ile Ile Ser	Phe Thr Asn Gln Arg Val Ile Asn Ala	Arg Trp Met Glu Thr Ala Ala Trp Ile	670 Val 685 700 Asp 715 Thr 730 Val 745 Arg 760 Glu 775 His 790 Asp 805	Ile Lys Phe Pro Met Asp Ala Tyr Pro	Ala Val Ala Val Cys Leu Ala Glu	Leu Gln Gly Val Thr Gly Pro Asp	Ala Asn Leu Gly Met Pro Ser Ile	675 His 690 Ile 705 Ile 720 Glu 735 Asp 750 Ile 765 Lys 780 Leu 795 Pro 810
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Ser Glu Leu Glu Ala Ser Val Ala Ser Pro Phe Thr Ser Lys Thr
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              905
Pro Ser Ile Ser Cys Val Pro Asn Leu Ile Arg Glu Gly Arg Ala
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Ala Leu Ile Thr Ser Phe Cys Val Phe Lys Phe Met Ala Leu Tyr
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Ser Ile Ile Gln Tyr Phe Ser Val Thr Leu Leu Tyr Ser Ile Leu
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Ser Asn Leu Gly Asp Phe Gln Phe Leu Phe Ile Asp Leu Ala Ile
              965
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Ile Leu Val Val Val Phe Thr Met Ser Leu Asn Pro Ala Trp Lys
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Glu Leu Val Ala Gln Arg Pro Pro Ser Gly Leu Ile Ser Gly Ala
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Leu Leu Phe Ser Val Leu Ser Gln Ile Ile Ile Cys Ile Gly Phe
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Gln Ser Leu Gly Phe Phe Trp Val Lys Gln Gln Pro Trp Tyr Glu
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His Asn Ile Gln Asn Tyr Glu Asn Thr Thr Val Phe Phe Ile Ser
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Pro Phe Arg Gln Pro Cys Tyr Lys Asn Tyr Phe Phe Val Phe Ser
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Val Ser Ile Thr Val Glu Glu Ser Val Asp Arg Trp Gly Lys Cys
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                                    445
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Cys Ser Cys Ile Lys Glu Gly Leu Ile Thr Ala Asp Ala Glu Gly
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Val Pro Asp Lys Leu Arg Ser Leu Ala Leu Gly Val Ser Tyr Val
Ile Leu Arg Ile Phe Gly Thr Ile Pro Gly Pro Ser Ile Phe Lys.
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                                    565
Gly His Thr Gly Arg Cys Trp Ile Tyr Asn Lys Thr Lys Met Ala
                575
                                    580
Phe Leu Leu Val Gly Ile Cys Phe Leu Cys Lys Leu Cys Thr Ile
                590
                                    595
Ile Phe Thr Thr Ile Ala Phe Phe Ile Tyr Lys Arg Arg Leu Asn
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                                   610
Glu Asn Thr Asp Phe Pro Asp Val Thr Val Lys Asn Pro Lys Val
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Lys Lys Glu Glu Thr Asp Leu
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Ala Pro Ala Ala Gly Gly Leu Asn Ala Gly Pro Val Pro Pro Ala
Ala Leu Ser Thr Gln Arg Leu Arg Asn Glu Asp Tyr His Asp Tyr
                 35
                                     40
Ser Ser Thr Asp Val Ser Pro Glu Glu Ser Pro Ser Glu Gly Leu
                 50
                                     55
Asn Asn Leu Ser Ser Pro Gly Ser Tyr Gln Arg Phe Gly Gln Ser
                 65
                                     70
Asn Ser Thr Thr Trp Phe Gln Thr Leu Ile His Leu Leu Lys Gly
                 80
                                     85
```

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Asn Ile Gly Thr Gly Leu Leu Gly Leu Pro Leu Ala Val Lys Asn
                    •
                                   100
                95
Ala Gly Ile Val Met Gly Pro Ile Ser Leu Leu Ile Ile Gly Ile
                                    115
Val Ala Val His Cys Met Gly Ile Leu Val Lys Cys Ala His His
                125
Phe Cys Arg Arg Leu Asn Lys Ser Phe Val Asp Tyr Gly Asp Thr
                                   145
Val Met Tyr Gly Leu Glu Ser Ser Pro Cys Ser Trp Leu Arg Asn
                                   160
                155
His Ala His Trp Gly Arg Arg Val Val Asp Phe Phe Leu Ile Val
                170
                                    175
Thr Gln Leu Gly Phe Cys Cys Val Tyr Phe Val Phe Leu Ala Asp
                                    190
                185
Asn Phe Lys Gln Val Ile Glu Ala Ala Asn Gly Thr Thr Asn Asn
                                    205
Cys His Asn Asn Glu Thr Val Ile Leu Thr Pro Thr Met Asp Ser
                215
                                    220
Arg Leu Tyr Met Leu Ser Phe Leu Pro Phe Leu Val Leu Leu Val
                230
                                    235
Phe Ile Arg Asn Leu Arg Ala Leu Ser Ile Phe Ser Leu Leu Ala
                                   250
                245
Asn Ile Thr Met Leu Val Ser Leu Val Met Ile Tyr Gln Phe Ile
                                    265
                260
Val Gln Arg Ile Pro Asp Pro Ser His Leu Pro Leu Val Ala Pro
                                    280
                275
Trp Lys Thr Tyr Pro Leu Phe Phe Gly Thr Ala Ile Phe Ser Phe
                290
                                    295
Glu Gly Ile Gly Met Val Leu Pro Leu Glu Asn Lys Met Lys Asp
                                   310
Pro Arg Lys Phe Pro Leu Ile Leu Tyr Leu Gly Met Val Ile Val
                                    325
                320
Thr Ile Leu Tyr Ile Ser Leu Gly Cys Leu Gly Tyr Leu Gln Phe
Gly Ala Asn Ile Gln Gly Ser Ile Thr Leu Asn Leu Pro Asn Cys
                350
                                   355
Trp Leu Tyr Gln Ser Val Lys Leu Leu Tyr Ser Ile Gly Ile Phe
                                    370
                365
Phe Thr Tyr Ala Leu Gln Phe Tyr Val Pro Ala Glu Ile Ile Ile
                                    385
                380
Pro Phe Phe Val Ser Arg Ala Pro Glu His Cys Glu Leu Val Val
                395
                                    400
Asp Leu Phe Val Arg Thr Val Leu Val Cys Leu Thr Cys Ile Leu
                                    415
                410
Ala Ile Leu Ile Pro Arg Leu Asp Leu Val Ile Ser Leu Val Gly
                                    430
                425
Ser Val Ser Ser Ser Ala Leu Ala Leu Ile Ile Pro Pro Leu Leu
                                    445
Glu Val Thr Thr Phe Tyr Ser Glu Gly Met Ser Pro Leu Thr Ile
                                    460
                455
Phe Lys Asp Ala Leu Ile Ser Ile Leu Gly Phe Val Gly Phe Val
                470
                                    475
Val Gly Thr Tyr Glu Ala Leu Tyr Glu Leu Ile Gln Pro Ser Asn
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Ala Pro Ile Phe Ile Asn Ser Thr Cys Ala Phe Ile
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                 20
Met Pro Ala Lys Phe Val Arg Cys Ala Tyr Val Ile Ile Leu Met
                                     40
Ala Ile Tyr Trp Cys Thr Glu Val Ile Pro Leu Ala Val Thr Ser
                                     55
                 50
Leu Met Pro Val Leu Leu Phe Pro Leu Phe Gln Ile Leu Asp Ser
                 65
Arg Gln Val Cys Val Gln Tyr Met Lys Asp Thr Asn Met Leu Phe
                 80
                                     85
Leu Gly Gly Leu Ile Val Ala Val Ala Val Glu Arg Trp Asn Leu
                                    100
                 95
His Lys Arg Ile Ala Leu Arg Thr Leu Leu Trp Val Gly Ala Lys
                110
                                    115
Pro Ala Arg Leu Met Leu Gly Phe Met Gly Val Thr Ala Pro Leu
                                    130
                125
Ser Met Trp Ile Ser Asn Thr Ala Thr Thr Ala Met Wet Val Pro
                                    145
Ile Val Glu Ala Ile Leu Gln Gln Met Glu Ala Thr Ser Ala Ala
                                    160
Thr Glu Ala Gly Leu Glu Leu Val Asp Lys Gly Lys Ala Lys Glu
                170
                                    175
Leu Pro Gly Ser Gln Val Ile Phe Glu Gly Pro Thr Leu Gly Gln
                185
                                    190
Gln Glu Asp Gln Glu Arg Lys Arg Leu Cys Lys Ala Met Thr Leu
                                    205
                200
Cys Ile Cys Tyr Ala Ala Ser Ile Gly Gly Thr Ala Thr Leu Thr
                215
                                    220
Gly Thr Gly Pro Asn Val Val Leu Leu Gly Gln Met Asn Glu Leu
                230
                                    235
Phe Pro Asp Ser Lys Asp Leu Val Asn Phe Ala Ser Trp Phe Ala
                                    250
Phe Ala Phe Pro Asn Met Leu Val Met Leu Leu Phe Ala Trp Leu
                                    265
Trp Leu Gln Phe Val Tyr Met Arg Phe Asn Phe Lys Lys Ser Trp
                275
                                    280
Gly Cys Gly Leu Glu Ser Lys Lys Asn Glu Lys Ala Ala Leu Lys
                290
                                    295
Val Leu Gln Glu Glu Tyr Arg Lys Leu Gly Pro Leu Ser Phe Ala
                                    310
Glu Ile Asn Val Leu Ile Cys Phe Phe Leu Leu Val Ile Leu Trp
                320
                                    325
Phe Ser Arg Asp Pro Gly Phe Met Pro Gly Trp Leu Thr Val Ala
                335
                                    340
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Trp Val Glu Glu Glu Thr Lys Tyr Val Ser Asp Ala Thr Val Ala
                                   355
Ile Phe Val Ala Thr Leu Leu Phe Ile Val Pro Ser Gln Lys Pro
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Lys Phe Asn Phe Arg Ser Gln Thr Glu Glu Glu Arg Lys Thr Pro
                                   385
               380
Phe Tyr Pro Pro Pro Leu Leu Asp Trp Lys Val Thr Gln Glu Lys
               395
                                   400
Val Pro Trp Gly Ile Val Leu Leu Gly Gly Phe Ala Leu
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                                   415
Ala Lys Gly Ser Glu Ala Ser Gly Leu Ser Val Trp Met Gly Lys
               425
                                   430
Gln Met Glu Pro Leu His Ala Val Pro Pro Ala Ala Ile Thr Leu
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                                   445
Ile Leu Ser Leu Leu Val Ala Val Phe Thr Glu Cys Thr Ser Asn
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                                    460
Val Ala Thr Thr Leu Phe Leu Pro Ile Phe Ala Ser Met Ser
                                    475
Arg Ser Asn Gly Leu Asn Pro Leu Tyr Ile Met Leu Pro Cys Thr
                                   490
                485
Leu Ser Ala Ser Phe Ala Phe Met Leu Pro Val Ala Thr Pro Pro
                                   505
                500
Asn Ala Ile Val Phe Thr Tyr Gly His Leu Lys Val Ala Asp Met
                515
                                    520
Val Lys Thr Gly Val Ile Met Asn Ile Ile Gly Val Phe Cys Val
                                    535
Phe Leu Ala Val Asn Thr Trp Gly Arg Ala Ile Phe Asp Leu Asp
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                                   550
His Phe Pro Asp Trp Ala Asn Val Thr His Ile Glu Thr
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Ala Leu Phe Ile Gly Phe Ser Gln Phe Ser Asp Ser Phe Leu Leu
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                 20
Asp Gln Pro Asn Phe Trp Cys Arg Gly Ala Gly Lys Gly Thr Glu
Leu Ala Gly Val Thr Thr Gly Arg Gly Gly Asp Met Gly Asn
                50
                                     55
Trp Thr Ser Leu Pro Thr Thr Pro Phe Ala Thr Ala Pro Trp Glu
                65
                                    70
Ala Ala Gly Asn Arg Ser Asn Ser Ser Gly Ala Asp Gly Asp
                80
                                    85
Thr Pro Pro Leu Pro Ser Pro Pro Asp Lys Gly Asp Asn Ala Ser
                95
                                    100
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Asn Cys Asp Cys Arg Ala Trp Asp Tyr Gly Ile Arg Ala Gly Leu

				110					115					120
				125			Trp		130					135
Trp	Lys	Val	His	Ile 140	Ala	Lys	Phe	Ser	Leu 145	Leu	Val	Gly	Leu	Ile 150
Phe	Gly	Tyr	Leu	Ile 155	Thr	Gly	Cys	Ile	Ala 160	Asp	Trp	Va1	Gly	Arg 165
Arg	Pro	Va1	Leu	Leu 170	Phe	Ser	Ile	Ile	Phe 175	Ile	Leu	Ile	Phe	Gly 180
Leu	Thr	Va1	Ala	Leu 185	Ser	Va1	Asn	Val	Thr 190	Met	Phe	Ser	Thr	Leu 195
Arg	Phe	Phe	Glu	Gly 200	Phe	Сув	Leu	Ala	Gly 205	Ile	Ile	Leu	Thr	Leu 210
Tyr	Ala	Leu	Arg	Ile 215	Glu	Leu	Cys	Pro	Pro 220	Gly	Lys	Arg	Phe	Met 225
Ile	Thr	Met	Val		Ser	Phe	Va1	Ala	Met 235	Ala	Gly	Gln	Phe	Leu 240
Met	Pro	Gly	Leu	Ala 245	Ala	Leu	Cys	Arg	Asp 250	Trp	Gln	Val	Leu	Gln- 255
Ala	Leu	Ile	Ile	Cys 260	Pro	Phe	Leu	Leu	Met 265	Leu	Leu	Tyr	Trp	Ser 270
Ile	Phe	Pro	Glu	Ser 275	Leu	Arg	Trp	Leu	Met 280	Ala	Thr	Gln	Gln	Phe 285
Glu	Ser	Ala	Lys	Arg 290	Leu	I1e	Leu	His	Phe 295	Thr	Gln	Lys	Asn	Arg 300
Met	Asn	Pro	Glu	Gly 305	Asp	Ile	Lys	Gly	Val 310	Ile	Pro	Glu	Leu	Glu 315
Lys	Glu	Leu	Ser	Arg 320	Arg	Pro	Lys	Lys	Val 325	Cys	Ile	Va1	Lys	Val 330
Val	Gly	Thr	Arg	Asn 335	Leu	Trp	Lys	Asn	Ile 340	Va1	Val	Leu	Cys	Val 345
Asn	Ser	Leu	Thr	Gly 350	Tyr	Gly	Ile	His	His 355	Cys	Phe	Ala	Arg	Ser 360
Met	Met	Gly	His	Glu 365	Val	Lys	Va1	Pro	Leu 370	Leu	Glu	Asn	Phe	Tyr 375
Ala	Asp	Tyr	Tyr	Thr 380	Thr	Ala	Ser	Ile	A1a 385	Leu	Val	Ser	Cys	Leu 390
Ala	Met	Cys	Val	Val 395	Val	Arg	Phe	Leu	Gly 400	Arg	Arg	Gly	Gly	Leu 405
Leu	Leu	Phe	Met	Ile 410		Thr	Ala	Leu	Ala 415		Leu	Leu	Gln	Leu 420
Gly	Leu	Leu	Asn	Leu 425		Gly	Lys	Tyr	Ser 430		His	Pro	Asp	Ser 435
Gly	Met	Ser	Asp	Ser 440		Lys	Asp	Lys	Phe 445		Ile	Ala	Phe	Ser 450
Ile	Val	Gly	Met	Phe 455		Ser	His	Ala	Val 460		Ser	Leu	Ser	Val 465
Phe	Phe	Суз	Ala	·G1u 470		Thr	Pro	Thr	Va1 475		Arg	Суз	Gly	Gly 480
Leu	Gly	Leu	Val	Leu 485		Ser	Ala	Gly	Phe 490		Met	Leu	Thr	Ala 495
Pro	Ile	Ile	Glu	Leu 500		Asn	Gln	Lys	Gly 505		Phe	Leu	His	His 510
Ile	Ile	Phe	Ala	Cys 515		Thr	Leu	Ile	Cys 520		Ile	Cys	Ile	Leu 525
Leu	Leu	Pro	Glu	Ser	Arg	Asp	Gln	Asn	Leu	Pro	Glu	Asn	Ile	Ser

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530
                                    535
Asn Gly Glu His Tyr Thr Arg Gln Pro Leu Leu Pro His Lys Lys
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                545
Gly Glu Gln Pro Leu Leu Leu Thr Asn Ala Glu Leu Lys Asp Tyr
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Ser Gly Leu His Asp Ala Ala Ala Ala Gly Asp Thr Leu Pro Glu
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Gly Ala Thr Ala Asn Gly Met Lys Ala Met
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Val Phe Ala Leu Pro Pro Lys Glu Gly Arg Cys Leu Phe Val Ile
                 35
                                     40
Leu Leu Met Ala Val Tyr Trp Cys Thr Glu Ala Leu Pro Leu Ser
                                     55
                 50
Val Thr Ala Leu Leu Pro Ile Val Leu Phe Pro Phe Met Gly Ile
                                     70
Leu Pro Ser Asn Lys Val Cys Pro Gln Tyr Phe Leu Asp Thr Asn
                                     85
Phe Leu Phe Leu Ser Gly Leu Ile Met Ala Ser Ala Ile Glu Glu
                 95
                                    100
Trp Asn Leu His Arg Arg Ile Ala Leu Lys Ile Leu Met Leu Val
               110
                                   115
Gly Val Gln Pro Ala Arg Leu Ile Leu Gly Met Met Val Thr Thr
                                    130
                125
Ser Phe Leu Ser Met Trp Leu Ser Asn Thr Ala Ser Thr Ala Met
                140
                                   145
Met Leu Pro Ile Ala Asn Ala Ile Leu Lys Ser Leu Phe Gly Gln
                                    160
                155
Lys Glu Val Arg Lys Asp Pro Ser Gln Glu Ser Glu Glu Asn Thr
                170
                                    175
Ala Ala Val Arg Arg Asn Gly Leu His Thr Val Pro Thr Glu Met
                                    190
Gln Phe Leu Ala Ser Thr Glu Ala Lys Asp His Pro Gly Glu Thr
Glu Val Pro Leu Asp Leu Pro Ala Asp Ser Arg Lys Glu Asp Glu
                215
                                    220
Tyr Arg Arg Asn Ile Trp Lys Gly Phe Leu Ile Ser Ile Pro Tyr
                230
                                    235
Ser Ala Ser Ile Gly Gly Thr Ala Thr Leu Thr Gly Thr Ala Pro
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                                    250
Asn Leu Ile Leu Cly Gln Leu Lys Ser Phe Pro Gln Cys
                260
                                    265
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Asp Val Val Asn Phe Gly Ser Trp Phe Ile Phe Ala Phe Pro Leu
                                    280
Met Leu Leu Phe Leu Leu Ala Gly Trp Leu Trp Ile Ser Phe Leu
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Tyr Gly Gly Leu Ser Phe Arg Gly Trp Arg Lys Asn Lys Ser Glu
                305
                                    310
Ile Arg Thr Asn Ala Glu Asp Arg Ala Arg Ala Val Ile Arg Glu
                                   325
                320
Glu Tyr Gln Asn Leu Gly Pro Ile Lys Phe Ala Glu Gln Ala Val
                                   340
                335
Phe Ile Leu Phe Cys Met Phe Ala Ile Leu Leu Phe Thr Arg Asp
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                350
Pro Lys Phe Ile Pro Gly Trp Ala Ser Leu Phe Asn Pro Gly Phe
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                365
Leu Ser Asp Ala Val Thr Gly Val Ala Ile Val Thr Ile Leu Phe
Phe Phe Pro Ser Gln Arg Pro Ser Leu Lys Trp Trp Phe Asp Phe
                                    400
                395
Lys Ala Pro Asn Thr Glu Thr Glu Pro Leu Leu Thr Trp Lys Lys
                                    415
                410
Ala Gln Glu Thr Val Pro Trp Asn Ile Ile Leu Leu Gly Gly
                425
                                   430
Gly Phe Ala Met Ala Lys Gly Cys Glu Glu Ser Gly Leu Ser Val
                                    445
                440
Trp Ile Gly Gly Gln Leu His Pro Leu Glu Asn Val Pro Pro Ala
                                    460
                455
Leu Ala Val Leu Leu Ile Thr Val Val Ile Ala Phe Phe Thr Glu
                                    475
                470
Phe Ala Ser Asn Thr Ala Thr Ile Ile Ile Phe Leu Pro Val Leu
                                    490
Ala Glu Leu Ala Ile Arg Leu Arg Val His Pro Leu Tyr Leu Met
                                    505
Ile Pro Gly Thr Val Gly Cys Ser Phe Ala Phe Met Leu Pro Val
                515
                                    520
Ser Thr Pro Pro Asn Ser Ile Ala Phe Ala Ser Gly His Leu Leu
                530
                                   535
Val Lys Asp Met Val Arg Thr Gly Leu Leu Met Asn Leu Met Gly
                545
                                    550
Val Leu Leu Ser Leu Ala Met Asn Thr Trp Ala Gln Thr Ile
                560
                                   565
Phe Gln Leu Gly Thr Phe Pro Asp Trp Ala Asp Met Tyr Ser Val
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                575
Asn Val Thr Ala Leu Pro Pro Thr Leu Ala Asn Asp Thr Phe Arg
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Thr Leu

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<213> Homo sapiens

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Gln Asp Asp Gly Gln Asp Arg Glu Arg Leu Thr Tyr Phe Gln Asn
                 35
                                     40
Leu Pro Glu Ser Leu Thr Ser Leu Leu Val Leu Leu Thr Thr Ala
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                                    55
Asn Asn Pro Asp Val Met Ile Pro Ala Tyr Ser Lys Asn Arg Ala
                65
                                     70
Tyr Ala Ile Phe Phe Ile Val Phe Thr Val Ile Gly Ser Leu Phe
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                                    85
Leu Met Asn Leu Leu Thr Ala Ile Ile Tyr Ser Gln Phe Arg Gly
                 95
                                   100
Tyr Leu Met Lys Ser Leu Gln Thr Ser Leu Phe Arg Arg Arg Leu
                                    115
Gly Thr Arg Ala Ala Phe Glu Val Leu Ser Ser Met Val Gly Glu
Gly Gly Ala Phe Pro Gln Ala Val Gly Val Lys Pro Gln Asn Leu
                                    145
                140
Leu Gln Val Leu Gln Lys Val Gln Leu Asp Ser Ser His Lys Gln
                155
                                   160
Ala Met Met Glu Lys Val Arg Ser Tyr Gly Ser Val Leu Leu Ser
                                   175
                170
Ala Glu Glu Phe Gln Lys Leu Phe Asn Glu Leu Asp Arg Ser Val
                185
                                   190
Val Lys Glu His Pro Pro Arg Pro Glu Tyr Gln Ser Pro Phe Leu
                                    205
                200
Gln Ser Ala Gln Phe Leu Phe Gly His Tyr Tyr Phe Asp Tyr Leu
                                    220
                215
Gly Asn Leu Ile Ala Leu Ala Asn Leu Val Ser Ile Cys Val Phe
                                    235
Leu Val Leu Asp Ala Asp Val Leu Pro Ala Glu Arg Asp Asp Phe
                245
                                    250
Ile Leu Gly Ile Leu Asn Cys Val Phe Ile Val Tyr Tyr Leu Leu
                260
                                    265
Glu Leu Leu Lys Val Phe Ala Leu Gly Leu Arg Gly Tyr Leu
                275
                                    280
Ser Tyr Pro Ser Asn Val Phe Asp Gly Leu Leu Thr Val Val Leu
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                290
Leu Glu Ala Gly Asp Gly Gly Pro Ala Val Ala Val Gly His Asp
                                    310
                305
Pro His Ala Glu His Ala His Arg Val Pro Leu Pro Ala Tyr His
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                                    325
Pro Gln His Glu Ala Asp Gly Arg Gly Gly Gln Tyr Arg Pro Gly
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Pro Gly Ala Glu His Ala Cys Val Trp Arg Asp Pro Gly Gly Gly
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                                    355
Leu Leu Arg Ile Cys His His Trp Asp Gln Leu Val
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                 20
Gln Arg Gln Ala Ile Lys Gly Ala Phe Tyr Arg Arg Ala Gln Arg
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                                     40
Leu Arg Pro Gln Asp Glu Pro Arg Gln Gly Cys Gln Pro Glu Asp
                                     55
                 50
Arg Arg Arg Ile Ile Ile Asn Val Gly Gly Ile Lys Tyr Ser
                 65
                                    70
Leu Pro Trp Thr Thr Leu Asp Glu Phe Pro Leu Thr Arg Leu Gly
                 80
                                     85
Gln Leu Lys Ala Cys Thr Asn Phe Asp Asp Ile Leu Asn Val Cys
                                    100
Asp Asp Tyr Asp Val Thr Cys Asn Glu Phe Phe Asp Arg Asn
                                   115
                110
Pro Gly Ala Phe Gly Thr Ile Leu Thr Phe Leu Arg Ala Gly Lys
                                   130
               125
Leu Arg Leu Leu Arg Glu Met Cys Ala Leu Ser Phe Gln Asp Ser
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Asp Ile Leu Phe Gly Ser Ala Ser Ser Asp Thr Arg Asp Asn Asn
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Gly Arg Ala Ser Arg Gln Leu Val Leu Val Val Val Phe Val Ala
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                                     25
Leu Leu Leu Asp Asn Met Leu Phe Thr Val Val Val Pro Ile Val
                 35
Pro Thr Phe Leu Tyr Asp Met Glu Phe Lys Glu Val Asn Ser Ser
                 50
Leu His Leu Gly His Ala Gly Ser Ser Pro His Ala Leu Ala Ser
                                    70
                 65
Pro Ala Phe Ser Thr Ile Phe Ser Phe Phe Asn Asn Asn Thr Val
                                    85
Ala Val Glu Glu Ser Val Pro Ser Gly Ile Ala Trp Met Asn Asp
                 95
                                    100
Thr Ala Ser Thr Ile Pro Pro Pro Ala Thr Glu Ala Ile Ser Ala
                110
                                    115
His Lys Asn Asn Cys Leu Gln Gly Thr Gly Phe Leu Glu Glu Glu
```

				105					130					135
mե	mlo se	7	77-7	125	17-1	T 011	Dho	71-		Lys	7.1 -	₹7±1	Mot	
TIIT	TIIT	Arg	vaı	140	vaı	Leu	LIIC	пта	145	пур	ALG	V CL.	TICC	150
Len	Leu	Va1	Asn		Phe	Val	Glv	Pro		Thr	Asn	Ara	Ile	
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Tyr	His	Ile	Pro	Met	Phe	Ala	Gly	Phe	Va1	Ile	Met	Phe	Leu	Ser
_				170					175					180
Thr	Va1	Ser	Leu	Gly	Met	Leu	Ala	Ser	Val	Tyr	Thr	Asp	Asp	His
				185					190					195
Glu	Arg	${\tt Gly}$	Arg	Ala	Met	Gly	Thr	Ala	Leu	Gly	Gly	Leu	Ala	
			_	200					205	=		_		210
Gly	Leu	Leu	Val		Ala	Pro	Phe	Gly		Val	Met	Tyr	Glu	
TT 7	a1	т	G	215	D	Dla =	T 011	т1 -	220	71-	T2h o	T 011	71 -	225
vaı	GIY	гàг	ser	230	Pro	Pne	ьeu	тте	235	Ala	Pne	цец	Ата	240
T.em	Δen	G1v	Δla		G1n	T.e.u	Cvs	T1e		Gln	Pro	Ser	Tvs	
шси	тър	ريي	2114	245	0111		0,0		250					255
Ser	Pro	Glu	Ser		Lys	G1y	Thr	Pro		Phe	Met	Leu	Leu	Lys
				260	_				265					270
Asp	Pro	Tyr	Ile	Leu	Val	Ala	Ala	Gly	Ser	Ile	Cys	Phe	Ala	Asn
				275					280					285
Met	${ t Gly}$	Va1	Ala		Leu	Glu	Pro	Thr		Pro	Ile	Trp	Met	
	_,		_	290		_		~1	295	~ 1	-	77 .	T)]	300
Gln	Thr	Met	Cys		Pro	Lys	.I.r.b	Gin		Gly	Leu	Ата	Pne	ьец 315
Dwo	71 -	Cox	₹ <i>7</i> ~ 1	305	Th rac	T. 611	т1 о	Glar	310	Asn	T.011	Dhe	Glv	
PIO	ALA	Ser	val	320	T X T	дец	116	O T Y	325	22011	БСС	1110	C±1	330
Leu	Ala	Asn	Lvs		Glv	Ara	Trp	Leu		Ser.	Leu	Ile	Gly	
				335			-		340				_	345
Leu	Val	Val	Gly	Thr	Ser	Leu	Leu	Cys	Val	Pro	Leu	Ala	His	Asn
				350					355					360
Ile	Phe	Gly	Leu	Ile	${ t G1y}$	Pro	Asn	Ala		Leu	Gly	Leu	Ala	
				365					370			>	_	375
Gly	Met	Val	Asp		Ser	Met	Met	Pro		Met	GIY	His	Leu	
3	T	7	TT -	380	G	77_7	(Th. 220	G1	385	₹ <i>7</i> ~ 1	The wave	7 T ~	T10	390
Asp	ьeu	Arg	HIS	395	Ser	val	тАт	GTA	400	Val	тАт	мта	TTE	405
Agn	va 1	Δla	Phe		Met	Glv	Phe	Δla		Gly	Pro	Ser	Thr	
ıı.p	V CC.11	1110	1110	410		2			415	1				420
G1y	Ala	Ile	Val		Ala	Ile	Gly	Phe	Pro	Trp	Leu	Met	Val	Ile
_				425					430					435
Thr	Gly	Val	Ile	Asn	Ile	Va1	Tyr	Ala	Pro	Leu	Cys	Tyr	Tyr	Leu
				440					445					450
Arg	Ser	Pro	Pro		Lys	Glu	Glu	Lys		Ala	Ile	Leu	Ser	
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Asp	Cys	Pro	Met			Arg	Met	лЛх		Thr	GIn	ьуs	Pro	
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